



Research Article

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Oregonin from the stems and leaves of Korean *Alnus* species (Betulaceae)

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ABSTRACT

A diarylheptanoid, (5S)-1,7-bis-(3,4-dihydroxyphenyl)-5-hydroxyheptane-3-on-5-O-β-D-xylopyranoside, named as oregonin (**1**), was isolated from the bark of *Alnus tinctoria* Sarg. which is a species of the genus *Alnus* species, growing throughout Korea. The structure elucidation was accomplished by various spectroscopic methods including Negative FAB-MS, ¹H-NMR and ¹³C-NMR techniques or comparison with authentic samples.

Keywords: *Alnus tinctoria*, Betulaceae, oregonin, Chemotaxonomy

INTRODUCTION

Genus *Alnus* refers to deciduous broad-leaved trees or shrubs found in damp areas, mountains and comprises of more than 17 species, *Alnus borealis* Koidzumi, *Alnus firma* Sieb. et Zucc., *Alnus hirtella* Koidz., *Alnus hirsuta* Turcz., *Alnus hirsuta* Turcz. var. *sibirica* Fischer, *Alnus japonica* Steudel, *Alnus maritima* var. *arguta* Regel., *Alnus japonica* Thunb. var. *koreana* Callier, *Alnus japonica* Thunb. var. *reginosa* Nakai, *Alnus japonica* Thunb. var. *serrata* Nakai, *Alnus mandshurica* Callier, *Alnus mandschurica* Thunb. for *barbinervis*, *Alnus mandschurica* Thunb. for *pubescens* Kitagawa, *Alnus maximowiczii* Call, *Alnus mayrii* Call, *Alnus pendula* Matsum and *Alnus vermicularis* Nakai are growing in Korea[1]. The species examined in this study were *Alnus japonica* (stem, leaf), *Alnus hirsuta* (stem, leaf) (collected in Seoul National University Gwanak Arboretum, Anyang-si, Gyeonggi-do, Republic of Korea, 8 June 2009, vouchers AJS2009-01, AJL2009-01, AHS2009-01 and AHL2009-01), *Alnus tinctoria* Sarg. (stem, leaf) and *Alnus hirsuta* Turcz var. *sibirica* Fischer (stem, leaf) (collected in Korea National Arboretum, Pocheon-si, Gyeonggi-do, Republic of Korea, 5 June 2009, vouchers ATS2009-01, ATL2009-01, AHTS2009-01 and AHTL2009-01) and they were certificated by Dr. Jin (Division of Horticulture and Education, Korea National Arboretum).

Diarylheptanoids are characteristic components of the *Alnus* species[2,3]. Several interesting biological activities of diarylheptanoids including their anti-inflammatory[4-7], anti-oxidant properties[8] and anti-atopic dermatitis[16] have previously been reported. In a previous study conducted in our lab, quantitative analysis of diarylheptanoids including oregonin (**1**) was conducted using HPLC on *Alnus japonica*, *Alnus hirsuta*, *Alnus hirsuta* var. *sibirica*[9], *Alnus pendula*, *Alnus firma* and *Alnus maximowiczii*[18]. Here, as part of our continuous search for diarylheptanoids from new natural sources, we describe the isolation and identification of oregonin (**1**) from the stems and leaves of *Alnus tinctoria* Sarg. and screening of oregonin (**1**) from some other *Alnus* species [*Alnus japonica* (stem, leaf), *Alnus hirsuta* (stem, leaf) and *Alnus hirsuta* Turcz var. *sibirica* Fischer (stem, leaf)].

EXPERIMENTAL SECTION

General experimental procedure

The stationary phases for the column chromatographic isolation were performed on Sephadex LH-20 (10-25 μm, GE Healthcare Bio-Science AB, Uppsala, Sweden), MCI-gel CHP 20P (75-150 μm, Mitsubishi Chemical, Tokyo,

Japan) and ODS-B gel (40-60 μm , Daiso, Osaka, Japan). ODS-B gel was used as stationary phase on middle pressure liquid chromatography (MPLC) system. Sample injector was Waters 650E (Waters, Seoul, Korea), detector was 110UV/VIS detector (Gilson, Middleton, WI) and pump was TBP5002 (Tauto Biotech, Sanghai, China). Thin layer chromatography (TLC) was carried out using a pre-coated silica gel 60 F₂₅₄ plate (Merck, Darmstadt, Germany) on chloroform, methanol and water (70:30:4, volume ratio). The spots were detected under UV radiation (254 nm) and by spraying with FeCl_3 and 10% H_2SO_4 followed by heating.

The components from the *Alnus* species were identified by several instrumental analyses. The 1D NMR such as ^1H - (300 or 600MHz) and ^{13}C - (75 or 150MHz) nuclear magnetic resonance (NMR) experiments were recorded with Gemini 2000 and VNS (Varian, Palo Alto, CA, USA) at center for research facilities on Chung-Ang University. Low resolution fast atom bombardment mass spectrum (LRFAB-MS) were recorded with JMSAX505WA (JEOL, Tokyo, Japan) at National Center for Inter-University Research Facilities on Seoul National University.

Extraction and isolation (stems)

Fresh, chopped stems (350 g) of *Alnus tinctoria* Sarg. were extracted using 80% aqueous MeOH at room temperature for 3 days. The filtrate was concentrated and applied to a Sephadex LH-20 column (10-25 μm , GE Healthcare Bio-Science AB, Uppsala, Sweden) containing increasing proportions of MeOH (30~100%) afforded 4 fractions (1-4). Repeated column chromatography of fraction 2 on MCI-Gel CHP 20P (75-150 μm , Mitsubishi Chemical, Tokyo, Japan) using a H_2O : methanol gradient has resulted in oregonin (**1**).

Extraction and isolation (leaves)

Fresh, chopped leaves (200 g) of *Alnus tinctoria* Sarg. were extracted using 80% aqueous MeOH at room temperature for 3 days. The filtrate was concentrated and applied to a Sephadex LH-20 column (10-25 μm , GE Healthcare Bio-Science AB, Uppsala, Sweden) containing increasing proportions of MeOH (30~100%) afforded 4 fractions (1-4). Repeated column chromatography of fraction 2 on the MCI-Gel CHP 20P (75-150 μm , Mitsubishi Chemical, Tokyo, Japan) and then fraction 2-2 on ODS-B gel (40-60 μm , Daiso, Osaka, Japan) with 30%-100% methanol gradient in middle pressure liquid chromatography (MPLC) system (5 ml/min, 280 nm) has resulted in oregonin (**1**).

Oregonin (**1**)

Brown amorphous powder, Negative FAB MS: m/z 477 $[\text{M}-\text{H}]^-$, ^1H -NMR (600MHz, $\text{DMSO}-d_6+\text{D}_2\text{O}$): δ 6.67-6.60 (4H in total, H-2',2'',5',5''), 6.48-6.45 (2H in total, H-6'', 6'), 4.19 (1H, br d, $J=7.8\text{Hz}$, xyl-1), 4.03 (1H, m, H-5), 3.76 (1H, dd, $J=11.4, 6\text{Hz}$ xyl-5e), 3.35(1H, m, xyl-4), 3.08-2.56 (8H in total, H-1,2,4,7), 1.74-1.68 (2H in total, m, H-6)[10-12]. ^{13}C -NMR (150 MHz, $\text{DMSO}-d_6 + \text{D}_2\text{O}$): see Table 1[10-12].

RESULTS AND DISCUSSION

Quantitative analysis of Oregonin (**1**) of *Alnus tinctoria* Sarg. using high pressure liquid chromatography (HPLC)

HPLC was used for the quantitative analysis of the oregonin (**1**) contents. An Waters 600 series HPLC system (Milford, MA, USA), was employed and equipped with a vacuum degasser, a binary pump, a UV detector and column compartment. Oregonin (**1**) was separated on Kromasil 100-5 C18 (4.6 \times 250 mm, 5 μm particle) with a linear gradient water: acetonitrile = 90:10 to 60:40 for 24 min. The column temperature was maintained at room temperature and the flow rate was 1.0 ml/min. The system was monitored at 280 nm (λ_{max} of **1**) eluting at 18.54 ± 0.01 min. Oregonin (**1**) was detected in the extracts of *Alnus tinctoria* Sarg.(stem, leaf), *Alnus japonica* (stem, leaf), *Alnus hirsuta* (stem, leaf) and *Alnus hirsuta* Turcz var. *sibirica* Fischer (stem, leaf). We were able to quantify oregonin (**1**) from the stems and leaves extracts of *Alnus tinctoria* Sarg. ($4.24 \pm 0.002\%$), ($0.85 \pm 0.001\%$) using a calibration equation ($y=5201.9x-47967$; $R^2=0.9985$).

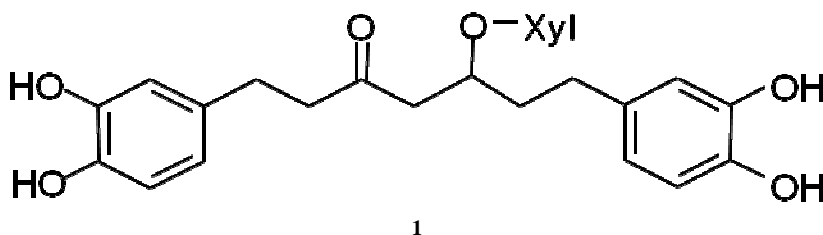
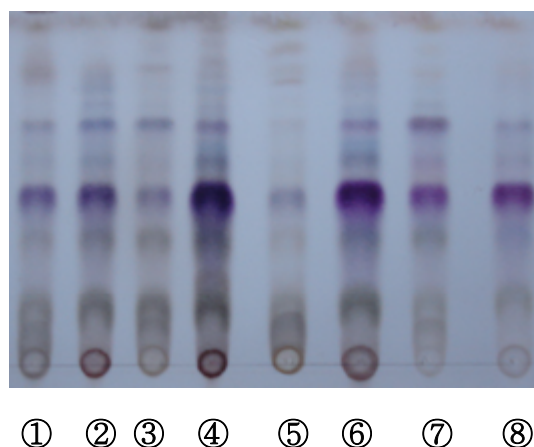


Fig. 1. Chemical structures of Oregonin (**1**) isolated from *Alnus tinctoria* Sarg.

Table 1. ^{13}C -NMR spectra of Oregonin (1)

Carbon No.	Oregonin (1)
C-1	28.7
C-2	45.1
C-3	209.6
C-4	47.5
C-5	76.9
C-6	39.7
C-7	30.5
C-1'	132.4
C-1''	133.3
C-2'	115.8
C-2''	115.8
C-3'	145.1
C-3''	145.1
C-4'	143.2
C-4''	143.4
C-5'	116.0
C-5''	116.0
C-6'	119.2
C-6''	119.3
Xyl-1	102.8
Xyl-2	74.7
Xyl-3	77.0
Xyl-4	69.8
Xyl-5	66.0

* 150 MHz (DMSO- d_6 + D_2O)**Fig. 2.** Comparative result of TLC chromatogram (10% sulfuric acid test)

TLC conditions: stationary phase (Silica gel 60 F₂₅₄), developing solvent[(CHCl₃/MeOH/H₂O(70:30:4)] and detection (diluted sulfuric acid test solution for spraying 105 °C, 5min)

① *A. japonica* (leaf), ② *A. japonica* (stem), ③ *A. hirsuta* (leaf), ④ *A. hirsuta* (stem), ⑤ *A. tinctoria* Sarg. (leaf), ⑥ *A. tinctoria* Sarg. (stem), ⑦ *A. hirsuta* Turcz. var. *sibirica* Fischer (leaf) and ⑧ *A. hirsuta* Turcz. var. *sibirica* Fischer (stem)

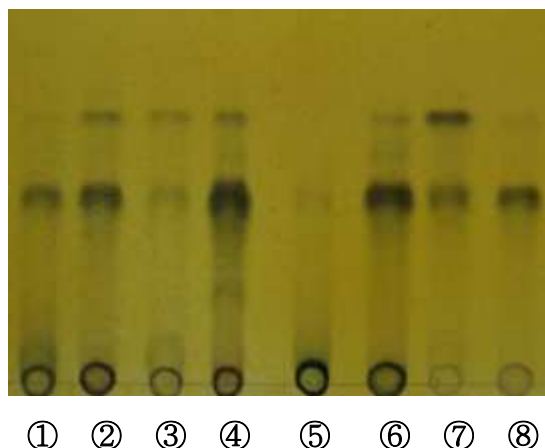


Fig. 3. Comparative result of TLC chromatogram (FeCl₃ test)

TLC conditions: stationary phase (Silica gel 60 F₂₅₄), developing solvent[CHCl₃/MeOH/H₂O(70:30:4)] and detection (FeCl₃ test solution for spraying, dry)

① *A. japonica* (leaf), ② *A. japonica* (stem), ③ *A. hirsuta* (leaf), ④ *A. hirsuta* (stem), ⑤ *A. tinctoria* Sarg. (leaf), ⑥ *A. tinctoria* Sarg. (stem), ⑦ *A. hirsuta* Turcz var. *sibirica* Fischer (leaf) and ⑧ *A. hirsuta* Turcz var. *sibirica* Fischer (stem)

Table 2. Retention time of Oregonin (1) from *Alnus* species

Material	Retention time (min)
	Oregonin (1)
Standard	18.54 ± 0.01
<i>A. japonica</i> (leaf)	18.22 ± 0.15
<i>A. japonica</i> (stem)	18.17 ± 0.02
<i>A. hirsuta</i> (leaf)	18.37 ± 0.15
<i>A. hirsuta</i> (stem)	18.60 ± 0.15
<i>A. tinctoria</i> Sarg. (leaf)	18.71 ± 0.15
<i>A. tinctoria</i> Sarg. (stem)	18.84 ± 0.15
<i>A. hirsuta</i> Turcz var. <i>sibirica</i> Fischer (leaf)	18.33 ± 0.15
<i>A. hirsuta</i> Turcz var. <i>sibirica</i> Fischer (stem)	18.31 ± 0.15

The results are expressed as means ± S.D (n=3).

Table 3. Contents of Oregonin (1) from *Alnus* species

Material	Contents (%)
	Oregonin (1)
<i>A. japonica</i> (leaf)	1.37 ± 0.002
<i>A. japonica</i> (stem)	5.13 ± 0.001
<i>A. hirsuta</i> (leaf)	2.20 ± 0.001
<i>A. hirsuta</i> (stem)	3.56 ± 0.001
<i>A. tinctoria</i> Sarg. (leaf)	0.85 ± 0.001
<i>A. tinctoria</i> Sarg. (stem)	4.24 ± 0.002
<i>A. hirsuta</i> Turcz var. <i>sibirica</i> Fischer (leaf)	1.27 ± 0.002
<i>A. hirsuta</i> Turcz var. <i>sibirica</i> Fischer (stem)	3.44 ± 0.001

The results are expressed as means ± S.D (n=3).

CONCLUSION

Chemotaxonomic significance

This is a report on the isolation and identification of oregonin (1) from *Alnus tinctoria* Sarg. Since the initial isolation of oregonin (1) from *Alnus rubra*[13], and oregonin (1) has only been distributed among *Alnus hirsuta*[3], *Alnus cordata*, *Alnus incana*, *Alnus viridis* and *Alnus glutinosa*[14] *Alnus japonica*[15], *Alnus serrulatoides*[11,12], *Pinus flexilis*[10] and *Alnus pendula*[17,18].

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REFERENCES

- [1] WT Lee. Lineamenta Florae Koreae, *Academy Press*, 1996, 1, 154.

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- [2] Y Asakawa. *B. Chem. Soc. Jpn.*, **1971**, 44, 2761.
- [3] MW Lee; T Tanaka; GI Nonaka; I Nishioka. *Phytochemistry*, **1992**, 31, 2835.
- [4] MW Lee; NY Kim; MS Park; KH Ahn; SH Toh; DR Hahn; YC Kim; HT Chung. *Planta. Med.*, **2000**, 66, 551.
- [5] MW Lee; JH Kim; DW Jeong; KH Ahn; SH Toh; YJ Surh. *Biol. Pharm. Bull.*, **2000**, 23, 517.
- [6] JM Han; WS Lee; JR Kim; JS Son; KH Nam; SC Choi; JS Lim; TS Jeong. *J. Agr. Food. Chem.*, **2007**, 55, 9457.
- [7] JM Han; WS Lee; JR Kim; JS Son; OH Kwon; HJ Lee; JJ Lee; TS Jeong. *J. Agr. Food. Chem.*, **2008**, 56, 92.
- [8] YA Lee; DW Jeong; KH Kim; JS Kim; SW Kim; MW Lee. *Yakhak Hoeji*, **2000**, 47, 193.
- [9] HW Lim; MK Kim; HJ Kim; JG Shim; GH Kim; HK Choi; MW Lee. *Kor. J. Pharmacog.*, **2004**, 35, 384.
- [10] KK Lee; BD Bahler; GA Hofman; MR Mattern; RK Johnson; DGI. Kingston. *J. Nat. Prod.*, **1998**, 61, 1407.
- [11] S Ohta; T Aoki; T Hirata; T Suga. *J. Chem. Soc.*, **1984**, 1, 1635.
- [12] T Suga; S Ohta; T Hirata; T Aoki; *Chem. Lett.*, **1982**, 6, 895.
- [13] JJ Karchesy; ML Laver; DF Barofsky; E Barofsky. *Chem. Commun.*, **1974**, 649.
- [14] NR Guz; P Lorenz; JP Metraux. *Biochem. Syst.*, **2002**, 30, 471.
- [15] T Aoki; S Ohta; T Suga. *Phytochemistry*, **1990**, 29, 3611.
- [16] SE Choi; MS Jeong; MJ Kang; DI Lee; SS Joo; CS Lee; H Bang; MK Lee; SC Myung; YW Choi; K Lee; SJ Seo; MW Lee. *Exp. Dermatol*, **2010**, 19, e37.
- [17] SE Choi; KH Park; MH Kim; JH Song; HY Jin; MW Lee. *Natural Product Sciences.*, **2012**, 18, 106.
- [18] SE Choi. *Asian Journal of Chemistry*, **2013**, 25, 6989.