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Research Article

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Lead removal from aqueous solutions by Bacillus subtilis

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ABSTRACT

The dead biomass of Bacillus subtilis has been used for the removal of lead (II) from an aqueous solution. The effects of various parameters such as contact time, adsorbate concentration, pH of the medium and temperature were examined. Optimum removal at 20° C was found to be 98.4 % at pH 6.5, with an initial Pb (II) concentration of 100 mgL⁻¹. Dynamics of the sorption process and mass transfer of Pb (II) to Bacillus subtilis were investigated and the values of rate constant of adsorption, rate constant of intraparticle diffusion and the mass transfer coefficients were calculated. Different thermodynamic parameters viz., changes in standard free energy, enthalpy and entropy were evaluated and it was found that the reaction was spontaneous and exothermic in nature. The adsorption data fitted the Langmuir isotherm. The data were subjected to multiple regression analysis and a model was developed to predict the removal of Pb (II) from an aqueous solution.

Keywords: Adsorption, monolayer, lead (II), Bacillus subtilis, Exothermic, multiple regressions, isotherm.

INTRODUCTION

The presence of heavy metals like Pb, Cd, Hg, Cr, Ni, Cu, Zn and Co are, at elevated concentrations, in the environment specifically in various water resources is of major concern because of their toxicity, non-biodegradable nature and threat to human, animal and plant life. WHO recommend in their guidelines for drinking water quality a threshold limit for lead is $10 \ \mu g \ L^{-1}$. In order to fulfill the threshold criteria and to reduce pollution contaminated waters need to be cleaned. To minimize this problem, biosorption can be part of the solution. Biosorption of heavy metals by bacterial fungal or algal biomass (live or dead cells) and agricultural waste biomass [1-12] has been recognized as a potential alternative to existing technologies such as precipitation, ion exchange, solvent extraction and liquid membrane for the removal of heavy metals from industrial wastewater because all these processes have the limitations of technical and/or economical viability.

The literature reveals two distinct approaches to use of living organisms and the use of a non-viable biomass [13-14]. There are significant practical limitations to systems, which employ living microorganisms. The most significant limitation is that microbial growth is inhibited when the concentrations of metal ions are too high or when significant amount of metal ions are adsorbed by microorganisms [13]. Dead cells or agricultural wastes accumulate heavy metal ions to the same or to a greater extent than living cells [13-14]. The reason for this is that the changes, which occur in the cell structure after the cells are dry- killed, affect adsorption in a positive manner [15]. For metal removal applications, the use of dead biomass or agricultural waste may be preferable as large quantities are readily and cheaply available as a byproduct of various industries [16]. Therefore, *Bacillus subtilis* was used for the removal of Pb (II) from aqueous solution.

EXPERIMENTAL SECTION

2.1 Media and Culture condition

The following medium and culture conditions were used for maintenance of cultures.

2.1.1. Maintenance of Culture

Bacillus subtilis strain was maintained on nutrient agar madia. The medium had the composition: Beef Extract- 1.0 gL⁻¹, Yeast Extract- 2.0 gL⁻¹, Peptone- 5.0 gL⁻¹, Agar- 2.0 gL⁻¹, NaCl- 5.0 gL⁻¹, Distilled water- 1000 mL, pH- 7.0.

2.1.2 Growth Medium

Bacillus subtilis was grown on the broth medium having the composition: D-glucose- 10 gL⁻¹, KH₂PO₄- 2 gL⁻¹, MgSO₄.7H2O- 0.5 gL⁻¹, NH₄Cl- 0.1 gL⁻¹, CaCl₂.2H₂O- 0.1 gL⁻¹, Thiamine- 0.001 gL⁻¹, Distilled water- 1000 mL, pH- 4.5.

2.1.3 Culture Condition

The culture condition of the *Bacillus subtilis* was given as: temperature- 37 °C, pH- 7.0, Aerobic, agitation rate- 200 rpm.

Initially, biomass of microorganisms was obtained through the cultivation in broth media followed by centrifugation, then autoclaved to obtain the dead biomass.

2.2 Experimental Procedure

In the present investigation batch mode of operation was selected in order to measure the progress of adsorption. Thus in a representative experiment 1.0 gm dead biomass of *Bacillus subtilis* was shaken with 50 ml of lead nitrate of desired concentration (100,125 and 150 mg L⁻¹) at different temperatures (20, 30 and 40° C) and pH (3.2, 4.0, 5.0, 5.5, 6.0, 6.5, 7.0, 7.5 and 8.0) in different glass bottles in a shaking thermostat set at 20, 30 and 40° C at a constant speed of 125 rpm. The pH of the adsorbate solution was adjusted by adding 0.1M HCl or 0.1M NaOH. The progress of the adsorption process was observed at different time intervals till the attainment of saturation. At the completion of predetermined time intervals, the adsorbate and adsorbent were separated by centrifugation at 15000 rpm and the supernatant liquid was analysed by AAS to determine the residual concentration of lead ion.

RESULTS AND DISCUSSION

3.1 Effect of contact time and concentration

A series of experiments were performed at different adsorbate concentration viz., 100, 125 and 150 mg L⁻¹ and time interval. The removal of Pb (II) was found to be 96.85, 92.15 and 85.75 % respectively, at 30 $^{\circ}$ C and pH 6.5. The extent of adsorption increased rapidly in the initial stages and became slower at later stages until the attainment of equilibrium. Equilibrium time for the adsorption of Pb (II) on *Bacillus subtilis* at various adsorbate concentrations was found to be 100 minutes, which showed that equilibrium time was independent of the initial adsorbate concentration. The following correlation had been developed between percentage removal and initial adsorbate concentration (C_o) to predict the percentage removal of Pb (II) by *Bacillus subtilis* at any initial concentration. Percentage removal of Pb (II) = 33.5 C_o^{0.63}

3.2 Adsorption dynamics

The rate constant k_{ad} (min⁻¹) for lead (II) adsorption on *Bacillus subtilis* was determined by using the first order rate kinetic equation [17].

$$\log (q_e - q) = \log q_e - \frac{k_{ad}}{2.303}.t$$
 (1)

Where q (mgg⁻¹) and q_e (mgg⁻¹) are the amounts of adsorbate at time t (min.) and at equilibrium respectively. The linear plots of log (q_e-q) versus t (Fig. 1) suggest the first order kinetics of the uptake of Pb (II). The values of k_{ad} (5.675 x 10⁻², 5.148 x 10⁻² and 4.665 x 10⁻² min⁻¹) at different temperatures were calculated from the slopes of these plots.

The data was also tested for pore diffusion using the following equation [18]:

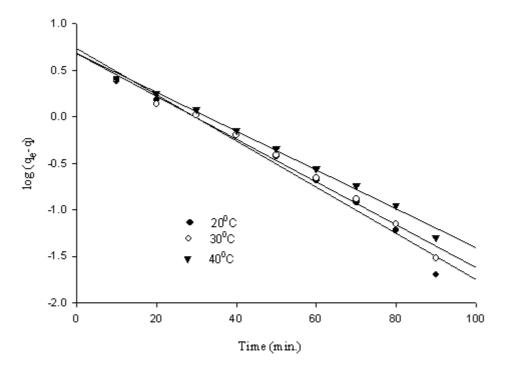
$$q = k_{id} t \frac{1}{2}$$
⁽²⁾

Where, q is the amount sorbed at time t and $t^{\frac{1}{2}}$ is the square root of the time. The values of k_{id} (3.582 x10⁻², 3.245 x10⁻² and 2.825 x10⁻² mgg⁻¹min^{-1/2}) at temperatures 20, 30 and 40°C respectively, were calculated from the slopes of respective plot q versus $t^{\frac{1}{2}}$ (Fig. 2) at later stages. The dual nature of the curves was obtained due to the varying extent of sorption in the initial and final stages of the experiment. This can be attributed to the fact that in the initial stages, sorption was due to boundary layer diffusion effect whereas, in the later stages (linear portion of the curve) was due to the intraparticle diffusion effects. However, these plots indicated that the intraparticle diffusion was not the only rate controlling step because it didn't pass through the origin. This was further supported by calculating the

intraparticle diffusion co-efficient (\overline{D} , cm² sec⁻¹) using the following equation [19].

$$\overline{D} = 0.03r^2 / t_{1/2}$$
(3)

where r (cm) is the average radius of the sorbent particle and $t_{1/2}$ (min.) is the time for half of the sorption. According to the Michelsen et al.[20] a \overline{D} (cm² sec⁻¹) value of the order of 10⁻¹¹ is indicative of intraparticle diffusion as rate determining step. In this investigation, the values of \overline{D} (8.142 x 10⁻⁹, 7.255 x 10⁻⁹ and 5.778 x 10⁻⁹ cm²sec⁻¹ at 20, 30 and 40°C respectively) obtained was in order of 10⁻⁹ cm² sec⁻¹ which was more than two order of magnitude higher, indicated that the intraparticle diffusion was not the only rate controlling step. It was concluded that both boundary layer and intraparticle diffusion might be involved in this removal process.



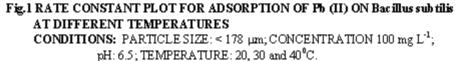
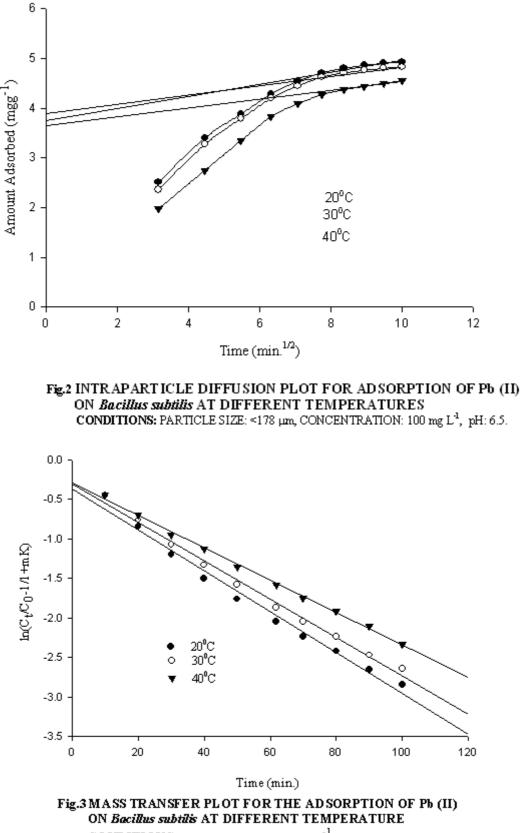
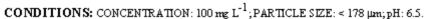


 Table 1: Values of thermodynamic parameters, Langmuir constants and R_L values of Pb (II) adsorption on

 Bacillus subtilis at different temperatures

| Temp. ⁰ C | $-\Delta G^0$ (k cal mol ⁻¹) | -ΔH ⁰ (k cal mol ⁻¹) | $-\Delta S^0$ (cal mol ⁻¹ k ⁻¹) | Q ⁰ (mg g ⁻¹) | b (l mg ⁻¹) | R _L |
|----------------------|--|--|--|---|----------------------------|----------------|
| 20 | 1.470 | | | 142.86 | 0.959 | 0.0103 |
| 30 | 1.067 | 18.344 | 57.019 | 140.85 | 0.542 | 0.0181 |
| 40 | 0.497 | | | 136.98 | 0.204 | 0.0467 |





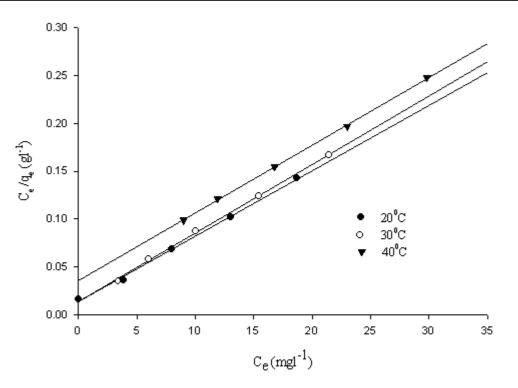


Fig.4 LANGMUIR ISOTHERM PLOT FOR THE ADSORPTION OF Pb (II) ON *Bacillus subtilis* AT DIFFERENT TEMPERATURES. CONDITIONS: PARTICLE SIZE:<178 µm; pH: 6.5; CONCENTRATION: 100, 110, 125, 140 & 150 mgL⁴.

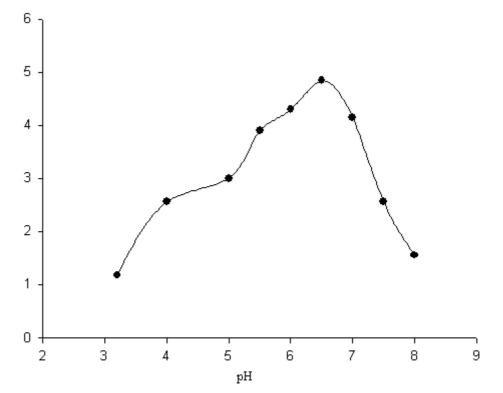


Fig.5 EFFE CT OF pH ON THE REMOVAL OF LEAD BY Bacillus *Bacillus subtilis*. CONDITIONS: CONCENTRATION: 100 mgL⁻¹; PARTICLE SIZE: <178 μm; TEMP.: 30⁰C; pH: 3.2, 4.0, 5.0, 5.5, 6.0, 6.5, 7.0, 7.5 and 8.0.

Table 2: Percentage removal at different conditions (experimental and predicted values at equilibrium time,
pH 6.5 and agitation rate 125 rpm)

| Initial adsorbate concentration (mg L ⁻¹) | Percentage removal | | Temp. | Percentage removal | |
|---|--------------------|-----------------|-------------------|--------------------|-----------------|
| Initial adsorbate concentration (ing L) | Exp. value | Predicted value | (⁰ C) | Exp. value | Predicted value |
| 100 | 96.85 | 97.9 | 20 | 98.64 | 99.8 |
| 125 | 92.15 | 91.0 | 30 | 96.85 | 97.9 |
| 150 | 85.75 | 86.5 | 40 | 90.78 | 93.2 |

3.3 Mass Transfer Study

The uptake of pollutant species from liquid phase (sorbate) to solid surface (sorbent) is carried out by transfer of mass from the former to the latter. A number of steps can be considered participating in the process and out of various models tried for the present studies, overall sorption process is assumed to occur using a three-step [21] model:

1. Mass transfer of sorbate from the aqueous phase on to the solid surface.

2. Sorption of solute on to the surface sites, and

3. Internal diffusion of solute via either a pore diffusion model or homogeneous solid phase diffusion model.

During the present investigation, step (2) has been assumed rapid enough with respect to the other steps and therefore it is not rate limiting in any kinetic study. Taking in to account these probable steps, Mckay et al. model has been used for the present investigation:

$$\ln\left(\frac{C_{A}}{C_{A_{0}}}-\frac{1}{1+mK}\right) = \ln\left(\frac{mK}{1+mK}\right) - \left(\frac{1+mK}{mK}\right)\beta_{1}S_{s}.t$$
(4)

where, m is the mass of the biosorbent per unit volume, K is the constant obtained by multiplying Q^0 and b (Langmuir's constants), β_1 is the mass transfer coefficient, S_s is the outer specific surface of the biosorbent particles per unit volume of particle-free slurry. The values of m and S_s were calculated using the following relations:

$$m = \frac{W}{V},$$
(5)

$$S_{s} = \frac{6m}{d_{p}\delta\rho(1-\varepsilon_{p})},$$
(6)

where, W is the weight of the adsorbent, V the volume of particle-free slurry solution, and dp, $\delta\rho$ and ϵp are the diameter, density and porosity of the adsorbent particles, respectively. The values of β_1 (5.585X 10⁻⁵, 4.965X 10⁻⁵ and 4.126X 10⁻⁵ cm sec⁻¹) calculated from the slopes and intercepts of the plots (Fig. 3) of ln (Ct/C0 – 1/1+ mK) versus t (min.) at different temperatures (20, 30 and 40 °C). The values of β_1 obtained show that the rate of transfer of mass from bulk solution to the biosorbent surface was rapid enough so it cannot be rate controlling step [22]. It can also be mentioned that the deviation of some of the points from the linearity of the plots indicated the varying extent of mass transfer at the initial and final stages of the sorption.

3.4 Thermodynamic evaluation of the process

This was again confirmed by thermodynamic parameters such as free energy (ΔG^0 , k cal mol⁻¹), enthalpy (ΔH^0 , k cal mol⁻¹) and entropy (ΔS^0 , cal mol⁻¹ k⁻¹) changes during the process. These parameters (Table 1) were calculated by the method described by Singh et al. [22] at 20, 30 and 40^oC temperatures. The negative and small values of free energy change (ΔG^0) were an indication of the spontaneous nature of the adsorption process. The negative values of standard enthalpy change (ΔH^0) for the intervals of temperatures was indicative of the exothermic nature of the adsorption process and the negative values of ΔS^0 for the corresponding temperature intervals suggested the probability of favourable adsorption.

3.5 Adsorption isotherm

The experimental data was found to fit the Langmuir isotherm. The basic assumption of Langmuir adsorption isotherm is based on monolayer coverage of the adsorbate on the surface of adsorbent. The saturated monolayer is represented by the following equation.

$$\frac{C_{e}}{q_{e}} = \frac{1}{Q^{0}b} + \frac{C_{e}}{Q^{0}}$$
(7)

Where, $C_e (mg L^{-1})$ is the equilibrium concentration of the adsorbate, $q_e (mgg^{-1})$ is the amount of adsorbate adsorbed at equilibrium; $Q^0 (mgg^{-1})$ and b (lmg^{-1}) is the Langmuir constants related to the capacity and energy of the adsorption respectively. The linearity of the plots C_e/q_e versus C_e (Fig. 4) showed the applicability of the Langmuir isotherm for the present system. Q^0 and b were determined from the slopes and intercepts of the respective plots. The decrease in their values (Table 1) with temperature increase also supported that removal of Pb (II) on *Bacillus subtilis* was exothermic in nature [22]. High Q^0 values (Table 1) also showed that the adsorbent had a good capacity to remove Pb (II).

The equilibrium parameter R_L which is defined as $R_L = 1/(1+bC_0)$ in the range $0 < R_L < 1$ reflects a favourable adsorption process [23] where b (1 mg^{-1}) is the Langmuir's constant and $C_0 (\text{mg } L^{-1})$ is initial adsorbate concentration. In the present investigation the equilibrium parameter (Table 1) was found to be in the range $0 < R_L < 1$ indicating that the adsorption process was favourable and the Langmuir isotherm was applicable [23].

3.6 Effect of pH

Experiments were performed at different pH values (3.2, 4.0, 5.0, 5.5, 6.0, 6.5, 7.0, 7.5 and 8.0). The percentage removal increased from 23.60 to 96.85 % with an increase of pH from 3.2 to 6.5 and thereafter removal decreased from 96.85 to 31.20 % with an increase of pH from 6.5 to 8.0, at 30^{0} C and Pb (II) concentration of 100 mg L⁻¹. The optimum pH for the removal of Pb (II) on *Bacillus subtilis* was found 6.5 (Fig. 5). Furthermore, the adsorption process can be explained on the basis of the nature of adsorbent used which contained several metal oxides. These oxides when mixed up with adsorbate solution undergo surface hydroxylation and form hydroxyl compounds on the surface which gives positively or negatively charged surface as a result of subsequent acid base dissociation [24]. It can be seen that adsorption increased in the pH range 3.0-6.5 and beyond pH 6.5, it started to decrease. The increase in adsorption correlated with the presence of Pb (II) ions up to pH 6.5. Beyond pH 6.5, the formation of hydroxide of Pb (II) occurred which resulted in a decrease in adsorption. This fact was supported by the distribution of lead species at various pH values [25].

MULTIPLE REGRESSION ANALYSIS

The effect of initial adsorbate concentration, contact time, temperature and pH of the system on Pb (II) removal by *Bacillus subtilis* had been examined. The cumulative effect of all these independent variables (lead removal) are given by the following relation:

$$Y = 6.5593 + 0.6385a_1 + 0.5168a_2 - 0.4679a_3 + 0.2835a_4 - 0.0959a_5 \tag{8}$$

Where, Y is the predicted value of Pb (II) removal, a_1 , concentration of adsorbate, a_2 , contact time; a_3 , temperature; a_4 , pH; a_5 , agitation rate of the system. The model values calculated with the help of equation (14) and the experimental values are given in Table 2. It may be seen that predicted values were pretty close to the experimental values. From these results it is concluded that all independent variables have cumulative effect on lead removal by *Bacillus subtilis*.

CONCLUSION

The dead biomass of *Bacillus subtilis* was found to be an effective biosorbent for the removal of Pb (II) from an aqueous solution. The study showed that the temperature and pH of the solution strongly influenced the adsorption process. Adsorption in the initial stages was due to the boundary layer diffusion whereas in the later stages adsorption was due to intraparticle diffusion. Thermodynamic studies confirmed that the process was spontaneous and exothermic. The fit of the adsorption data into the Langmuir isotherm confirmed monolayer adsorption. Mass transfer studies confirmed that the rate of mass transfer from sorbate to adsorbent was rapid enough. The data thus obtained from this investigation would be useful in designing and fabricating an efficient treatment plant for Pb (II) rich effluents.

REFERENCES

- [1] D Mohan; CU Jr Pittaman, J. Hazard. Mater., 2006, B137, 762-811.
- [2] KK Singh; M Talat; SH Hasan, Bioresource Technol., 2006, 97, 2124-2130.
- [3] KL Mandiwana; T Resane; N Panichev; P Ngobeni, J. Hazard. Mater., 2006, B137, 1241-1245.
- [4] YM Khlystov, Atmos. Environ., 2006, 40, 8088 8093.
- [5] SG Tuncel; S Tugrul; T Topal, Water Res., 2007, 41, 365 372.
- [6] H Harmens; DA Norris; GR Koerber; A Buse; E Steinnes; A Ruhling, Atmos. Environ., 2007, 41, 6673 6687.
- [7] SH Hasan; KK Singh; O Prakash; M Talat; YS Ho, J. Hazard. Mater., 2008, 152, 356-365.
- [8] HT Nguyen; KH Kim, Atmos. Environ., 2008, 42, 5015 5031.

[9] H Yin; B He; X Lu; H Peng; J Ye; F Yang, *Water Res.*, 2008, 42, 3981 – 3989.

- [10] YG Bermudez; ILR Rico; OG Bermudez; E Guibal. *Chem. Engg. J.*, **2011**, 166, 122 131.
- [11] MMM Rahmati; P Rabbani; A Abdolali; AR Keshtkar. J. Haz. Mater., 2011, 185, 401-407.
- [12] N Feng; X Guo; S Liang; Y Zhu; J Liu. J. Haz. Mater., 2011, 185, 49-54.

[13] DW Darnall; B Green; M Hosea; RA McPherson; M Henzl; MD Alexander; R Thomson, Editor. Ind.Div.Royal Soc. Chem. Congress, UK, **1986**, 1-24.

- [14] D Khummongkol; GS Canterford; C Fryer, Biotech. Bioeng., 1982, 24, 2643-2660.
- [15] M Nourbakhsh; Y Sag; D Ozer; Z Aksu; T Kutsal; AA Caglar, Process Biochem., 1994, 29, 1-5.
- [16] JM Tobin; B L'homme; JC Roux, Biotech. Tech. 1993, 7, 739-744.

[17] S Lagergren, Handlingar, 1898, 24, 1-39.

[18] WJ Weber; JC Morris, J. San. Eng. Div. Proc. Anal. Soc. Civil Engr., 1963, 89, SA2, 31-38.

- [19] AK Bhattacharya; C Venkobachar, J. Environ. Eng., 1984, 110, 110 120.
- [20] LD Michelsen; PG Gideon; EG Pace; LH Kutsal, USDI, Office of Water Res. and Tech. Bull. No. 74, 1975.
- [21] G Mckay; MS Otterburn; AG Sweeny, Water Res., 1981, 15, 327-339.
- [22] DB Singh; DC Rupainwar; G Prasad; KC Jayaprakas, J. Hazard. Mater., 1998, 60, 29-40.
- [23] GN Manju; MC Gigi; TS Anirudhan, Indian J. Chem. Tech., 1999, 6, 134-141.
- [24] KK Pandey; G Prasad; VN Singh, J. Chem. Technol. Biotechnol., 1984, 34A, 367-374.
- [25] GE Jean; GM Bancroft, Geochimica et Cosmochimica Acta, 1986, 50, 1455-1463.