



Research Article

ISSN : 0975-7384
CODEN(USA) : JCPRC5

Comparative studies for the cyclisation reactions of 2-alkynylaniline derivatives to generate the indole ring systems on polystyrene solid supports

Chinh T. Bui

Walter & Eliza Hall Institute of Medical Research, 4 Research Ave, Latrobe R & D Park, Bundoora, Vic 3086 and Department of Medical Biology, The University of Melbourne, Parkville Vic 3010, Melbourne, Australia

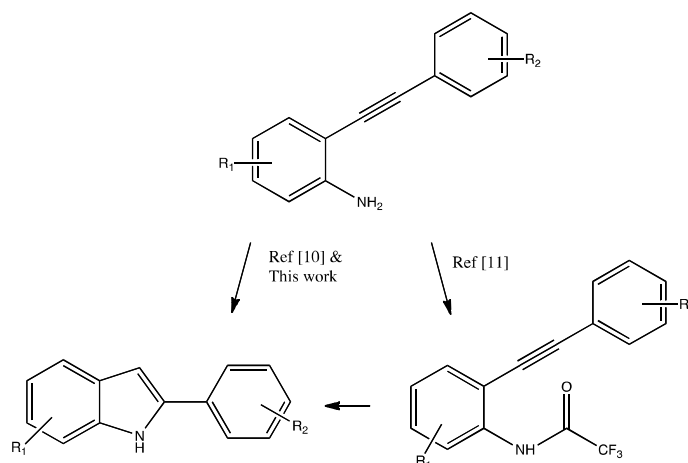
ABSTRACT

Generation of indole ring systems based on cyclisation of 2-alkynylaniline moieties on solid supports have been investigated. The cyclisation reactions were studied in 3 different conditions: (i) iodonium-mediated cyclisation (iodo-cyclisation) (ii) base-promoted cyclisation, and (iii) palladium (II)-mediated cyclisation. Only the latter with palladium dichloride gave satisfactory results on polystyrene resin solid supports.

Keywords Synthesis of indole, solid-phase synthesis, polystyrene, cyclisation of 2-alkynylaniline

INTRODUCTION

The indole ring system is regarded as a privileged set of structures used in medicinal chemistry as it has been found in many natural products [1] as well as pharmaceutical agents [2-4] to have a remarkable range of biological activities. Various synthetic routes have been developed in solution phase chemistry to obtain the heterocyclic indole rings, including Fischer's pathway [5], based on the coupling reactions of hydrazine with aldehyde or ketone, Wittig's indole synthesis between phosphonium salt of benzylamides [6], Nenitzescu's synthesis of indoles [7], gold-mediated condensation reactions between N-arylhydroxylamines and alkynes [8], etc. Among them, cyclisation of 2-alkynylaniline derivatives [9] with or without the presence of palladium as a catalyst has attracted our attention for a number of reasons: (i) The starting materials 2-alkynylanilines are readily derived from 2-iodoanilines via Sonogashira coupling reactions with a wide range of alkynes [10, 11] (ii) It has been established that the amino groups adjacent to the alkyne functional groups can undergo an intramolecular cyclisation in the presence of iodonium reagents (iodo-cyclisation process) [9] and (iii) Simplicity of the base-promoted cyclisations of 2-alkynylanilines has also been thoroughly investigated in solution-phase [12] and the reaction conditions have been carried out in the presence of strong bases such as potassium hydride, potassium *t*-butoxides [13] etc. However, for both solution and solid-phase synthesis of indoles, the amino group of 2-alkynylanilines required to be protected with trifluoroacetic acid [13] and the following cyclisation condition occurred with concomitant loss of the TFA group to generate the indole rings (Scheme 1). In addition, cyclisation of unprotected 2-alkynylanilines has not been fully investigated in solid-phase chemistry. This reason prompted us to investigate the solid-phase synthesis of indole ring systems from 2-alkynylanilines using three common reaction conditions: (i) iodonium-mediated cyclisation (iodocyclisation) (ii) base-promoted cyclisation, and (iii) palladium (II)-mediated cyclisation reactions.



Scheme 1: Cyclisation of 2-alkynylanilines with and without protection of amine moieties.

EXPERIMENTAL SECTION

Solvents and fine chemicals were purchased from Aldrich chemical company (Castle Hill, Australia) and used as supplied. Polystyrene resins derivatized with Fmoc-protected Rink Amide linker were purchased from Novabiochem (100 – 200 mesh, loading capacity: 0.47 mmol/g).

LC-MS analysis: LC analysis was conducted with a Phenomenex C18- (20 x 4.00 mm) column, using isocratic mobile-phase containing 10 % acetonitrile, 89.9 % water and 0.1 % of formic acid solution. Detection was measured at 214 nm and a flow rate was constant at 0.5 ml/min. MS analysis was conducted with Finnigan LCQ Avantage Max system.

Proton (^1H) and Carbon (^{13}C) NMR spectra were recorded with Bruker-300 spectrometer operating at 300 MHz for proton. All spectra were recorded in DMSO- d_6 or MeOD- d_4 at 25 °C.

Solid-phase reactions were carried out in 10-ml screw-cap vials (Wheaton vials, Aldrich). The reaction vials were incubated in a heat block and gently shaken at specified temperatures. After cleavage (20 % TFA in DCM) the residue was dried under a stream of N_2 gas. The residue was resuspended in ethyl acetate, passed through a plug of silica and the eluent was concentrated and submitted for LC-MS & NMR analysis without any further purification.

Preparation of resin 1:

Rink amide resin (1 g, polystyrene, 100 – 200 mesh, Novabiochem, 0.55 mmol/g) was suspended in 10 ml of 20 % piperidine in DMF for 30 min (Fmoc-deprotection) and carefully washed with DMF (3 x 5 mL) and DCM (3 x 5 mL). The resulting resin was gently mixed on a shaker with a mixture of 2-amino-3-iodobenzoic acid (0.26 g, 1.0 mmol), HOBt (0.135g, 1.0 mmol) and 1,3-diisopropylcarbodiimide (0.126 g, 1 mmol) in DCM/DMF (1:1, 10 mL) at 40 °C for 16h. The resins were carefully washed with DCM/DMF (1:1, 3 x 5 mL) and DCM (3 x 5 mL) and dried under reduced pressure for 2-3 hours to afford **1**.

Preparation of resin 2:

Resins **1** (0.5 g, 0.27 mmol) were incubated with a mixture of 4-methylphenylacetylene (115 mg, 1 mmol), $\text{PdCl}_2(\text{PPh}_3)_2$ (60 mg, 0.084 mmol) and CuI (10 mg, 0.10 mmol) in piperidine/ THF (1:1, 4 mL) at 60 °C for 16 h. The resulting resins were filtered and carefully washed with DMF (3 x 5 mL), methanol (5 mL) and DCM (3 x 5 mL) and dried under reduced pressure for 30 minutes to afford **2**.

Preparation of resin 3:

Resins **2** (0.5 g, 0.27 mmol) were incubated with PdCl_2 (3.4 mg, 0.02 mmol) in acetonitrile (2 mL) at 60 °C for 1-2 hours. The resulting resins were filtered and carefully washed with DMF (3 x 5 mL), methanol (3 x 5 mL) and DCM (3 x 5 mL) and dried under reduced pressure for 30 minutes to afford **3**.

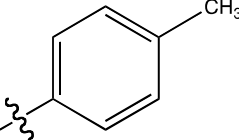
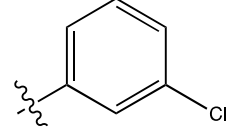
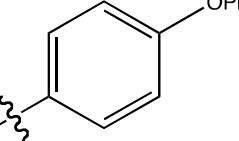
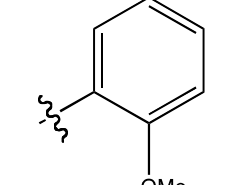
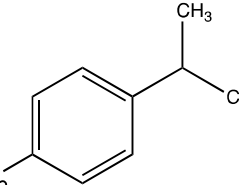
Compound **4a**: Resins **3** were finally incubated with 20 % TFA in DCM for 1 h. The cleavage solution was separated and dried under a stream of N₂ gas (15 min). The resulting crude products were dissolved in ethyl acetate (0.2 ml) and filtered through a plug of silica and the filtrate was concentrated and subjected to LC-MS and ¹H & ¹³C-NMR analysis to confirm structure of the desired product **4a**.

4a. R_t = 5.93 min. by LC; MS: m/z = 251.0 [M + H]⁺. ¹H-NMR (300 MHz, DMSO-d₆): 8.12 (s, 1H), 7.78 (d, J = 8.2 Hz, 2H), 7.66 (d, J = 8.8 Hz, 1H), 7.40 (d, J = 8.6 Hz 1H), 7.29 (d, J = 8.6 Hz, 2H), 6.92 (s, 1H), 2.4 (s, 3H). ¹³C-NMR (75 MHz, DMSO-d₆): 169.41, 139.54, 139.05, 137.57, 129.95, 129.56, 128.51, 125.51, 121.72, 120.51, 110.97, 99.40, 21.27.

4b. R_t = 5.0 min. by LC; MS: m/z = 271.2 [M + H]⁺. ¹H-NMR (300 MHz, DMSO-d₆): 8.15 (s, 1H), 8.06 (s, 1H), 7.88 (d, J = 7.8 Hz, 1H), 7.70 (d, J = 7.8 Hz, 1H), 7.50 (t, J = 8.3 Hz 1H), 7.6-7.3 (m, 2H), 7.12 (s, 1H). ¹³C-NMR (75 MHz, DMSO-d₆): 169.30, 139.32, 137.75, 134.48, 134.30, 131.24, 128.23, 127.74, 126.40, 125.08, 124.21, 122.37, 120.93, 111.30, 101.26.

4c. R_t = 5.28 min. by LC; MS: m/z = 329.5 [M + H]⁺. ¹H-NMR (300 MHz, DMSO-d₆): 8.14 (s, 1H), 7.91 (d, J = 9.2 Hz, 2H), 7.68 (d, J = 9.2 Hz, 1H), 7.46-7.38 (m, 2H), 7.18 (t, J = 6.4 Hz, 1H), 7.1 - 6.5 (m, 5H), 6.93 (s, 1H). ¹³C-NMR (75 MHz, DMSO-d₆): 169.42, 156.89, 156.82, 139.14, 138.98, 130.59, 128.54, 127.72, 127.36, 126.22, 124.16, 121.79, 120.54, 119.40, 119.30, 111.01, 99.56.

Table 2: LC-MS Analysis of the Compounds 4a-e

Compounds	R	% Purity	Rt (min)	LC-MS [M+H] found	LC-MS [M+H] Expected
4a		99	4.94	251.0	251.1
4b		98	5.00	271.2	271.1
4c		99	5.28	329.5	329.1
4d		99	4.83	267.7	267.5
4e		99	5.23	279.6	279.5

4d. R_t = 4.83 min. by LC; MS: m/z = 267.7 [M + H]⁺. ¹H-NMR (300 MHz, MeOD-d₄): 8.17 (s, 1H), 7.78 (d, J = 7.5 Hz, 1H), 7.68 (d, J = 8.9 Hz, 1H), 7.49 (d, J = 8.4 Hz 1H), 7.31 (t, J = 7.2 Hz, 1H), 7.11 (s, 1H), 7.04 (t, J = 8.2 Hz,

1H), 6.98 (s, 1H), 3.97 (3H, s). ¹³C-NMR (75 MHz, MeOD-d₄): 172.69, 156.40, 138.69, 137.02, 128.67, 128.02, 127.83, 124.22, 120.70, 120.66, 120.55, 120.19, 111.50, 110.49, 101.38, 54.67.

4e. R_t = 5.23 min. by LC; MS: m/z = 279.6 [M + H]⁺. ¹H-NMR (300 MHz, MeOD-d₄): 8.16 (s, 1H), 7.73 (d, J = 8.0 Hz, 2H), 7.68 (d, J = 8.6 Hz, 1H), 7.44 (d, J = 7.8 Hz, 1H), 7.32 (t, J = 7.5 Hz, 2H), 6.86 (s, 1H), 3.85 – 3.0 (m, 1H), 1.28 (d, J = 7.1, 6H). ¹³C-NMR (75 MHz, MeOD-d₄): 172.62, 148.51, 139.93, 139.42, 129.83, 128.81, 126.56, 124.96, 124.41, 120.77, 120.23, 110.34, 98.76, 33.76, 22.91.

RESULTS AND DISCUSSION

Scheme 2 outlines our solid-phase synthesis of a small group of 2-substituted indoles, using polystyrene resin as a solid support. Firstly, resin bound Rink amide linker was derivatized with 2-amino-3-iodobenzoic acid under standard coupling conditions (HOBt/DIC). The resulting resin was allowed to react with acetylene via Sonogashira coupling reaction to afford the substrate **2**. The resin **2** was subjected to a series of optimizing conditions (Table 1) for the cyclisation reactions. These conditions have proven to be effective in solution phase but they have not been analysed on solid supports. At first, the intramolecular cyclisation process mediated with iodonium or bromonium (TBAB) were carried out (entries 1 & 2, Table 1). This work followed the published procedure of cyclisation where the *N*-protected amine reacted rapidly with acetylenes, (either with methyl group [9] or TFA moiety [13]). However, no product was observed on solid-phase synthesis with unprotected amino groups in this study. Alternative methods using strong bases (potassium *t*-butoxide or NaH) were subsequently applied to the cyclisation reactions, but did not produce the desired result either (entries 3 & 4, table 1). It should be noted that the previously published reaction conditions [12] on Rink-MBHA-resins was not suitable for Rink-polystyrene-resins in this study. In fact, the polystyrene did not swell sufficiently in toluene, possibly explaining the poor yield in entry 3 (table 1) Surprisingly, Palladium-mediated cyclisation reactions with PdCl₂ as a catalyst worked well under very mild conditions (60 °C for less than 2 hours). It is notable that the reaction worked much better in acetonitrile than other solvents like dichloromethane or DMF as palladium formed a complex with acetonitrile, making it soluble under solid-phase conditions. Finally, cleavage by 20% TFA in DCM released target **4**, which was subjected to LC-MS analysis without further purification. The proton NMR of the compound **4** confirmed a single peak at around 6.9 ppm as the typical C-3 proton of indole ring. The reaction conditions were applied to several acetylenes and all were obtained in high yield (yield > 85%) and purity (Table 2).

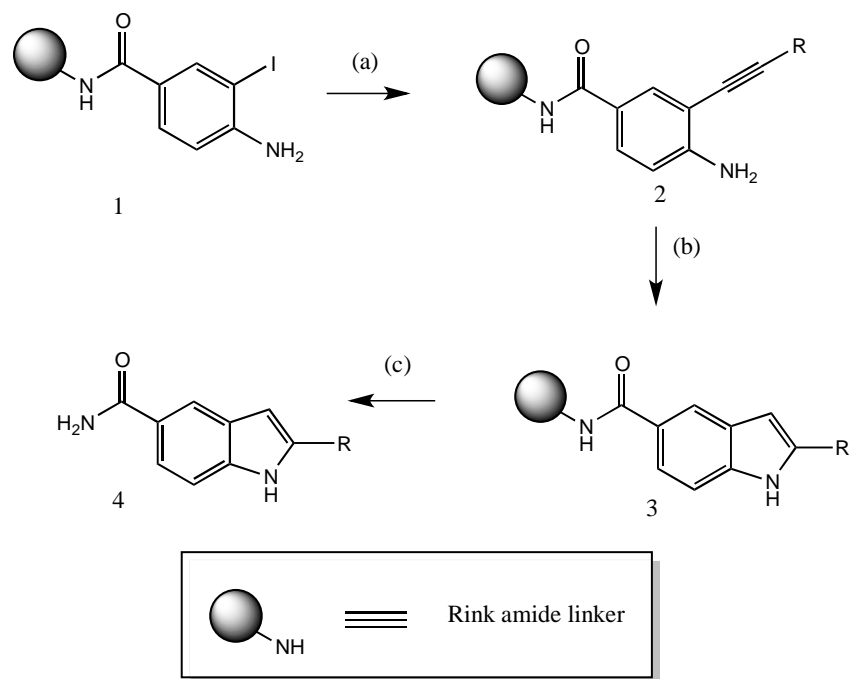


Figure 2: Solid-phase synthesis of 2-substituted indoles (R = substituted phenyl rings)

Table 1: Cyclisation Reactions of 2-Alkynylaniline Derivatives on Polystyrene Resin

Entry	R	Reaction conditions	% Purity	% Yield
1	4-Me-Ph	TBAB/THF/ 60 °C /12h	-	NR
2	4-Me-Ph	I ₂ in DCM/60 °C /4h	-	NR
3	4-Me-Ph	K ⁺ <i>t</i> -BuO ⁻ /NMP/toluene or DCM/ 25 °C 4h then 60 °C /12h	-	<10
4	4-Me-Ph	NaH/NMP/toluene or DCM/60 °C /12h	-	NR
5	4-Me-Ph	PdCl ₂ /AcCN/ 60 °C/2h	99	95

CONCLUSION

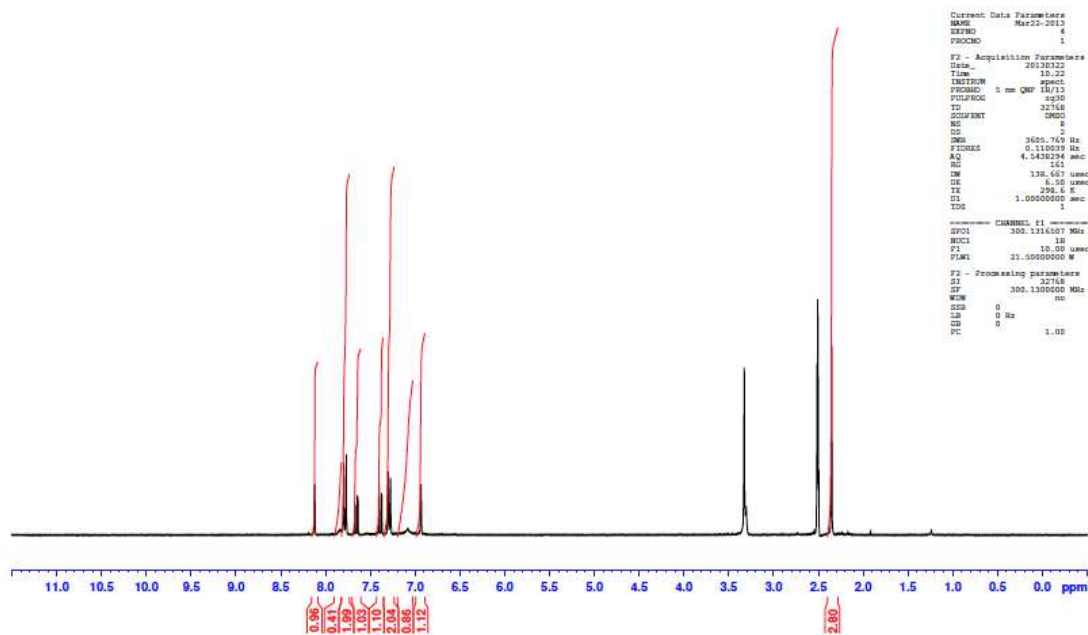
This study focuses on the preparation of 2-substituted indoles, which has never been fully investigated on polystyrene resin solid supports so far. Here, we reported the successful conditions for cyclisation reaction of 2-alkynylaniline derivatives in the presence of PdCl₂ catalyst/acetonitrile solvent.

Acknowledgement

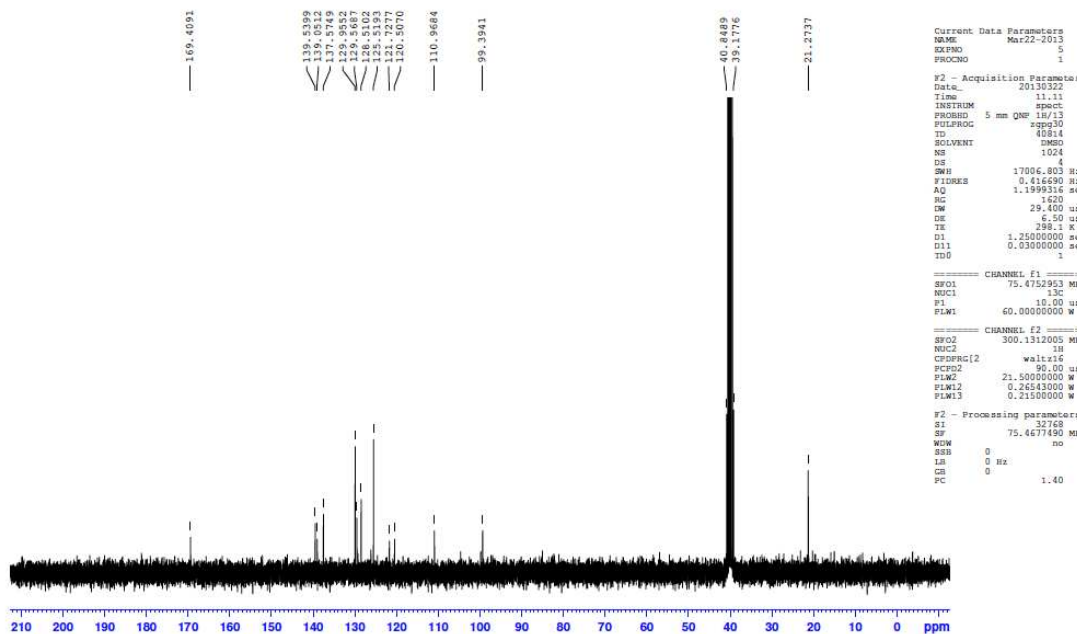
Author would like to thank Ms. Danuta Buczek for accumulation of LC-MS Data

REFERENCES

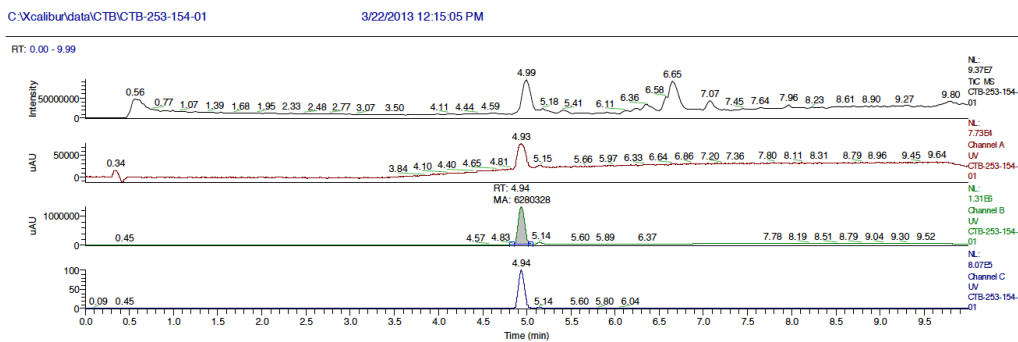
- [1] M Lounasmaa, A Tolvanen, *Nat. Prod. Rep.*, **2000**, 17, 175 - 191.
- [2] D, Ramesh, A. Avinash, S. K. Salenaagina, M. V. Saicharan Teja, P. Masthanaiah, P. Raja Rathnam and V.Chandana silpa, *J. Chem. Pharm. Res.*, **2011**, 3(5), 519 - 523.
- [3] K. Swathi , A. Srinivas and M. Sarangapani, *J. Chem. Pharm. Res.*, **2010**, 2(2), 220-225.
- [4] D. D. Bhargual,; N. Kumar, S. Drabu, *J. Chem. Pharm. Res.*, **2010**, 2(2), 345-349.
- [5] SM Hutchins; KT Chapman, *Tetrahedron Lett.* **1996**, 37, 4869 - 4872.
- [6] I Hughes, *Tetrahedron Lett.* **1996**, 37, 7595 - 7598.
- [7] DM Ketcha, LJ Wilson, DE Portlock, *Tetrahedron Lett.*, **2000**, 41, 6253 – 6257.
- [8] A Arcadi, G Bianchi, F Marinelli, *Synthesis*, **2004**, 4: 610-618.
- [9] Halim R, Scammells PJ, Flynn BL *Org. Lett.*, **2008**, 10, 1967–1970.
- [10] CCG Boojamra, KM Burow, JA Ellman, *J. Org. Chem.*, **1995**, 60, 5742–5743.
- [11] CT Bui, BL Flynn, *J. Comb. Chem.*, **2006**, 8,163–167.
- [12] C Koradin, W Dohle, AL Rodriguez, B Schmid, P Knochel, *Tetrahedron*, **2003**, 59, 1571 - 1587.
- [13] MD Collini, JW Ellingboe, *Tet. Lett.*, **1997**, 38 (46), 7963-7966.



¹H NMR spectrum of 4a

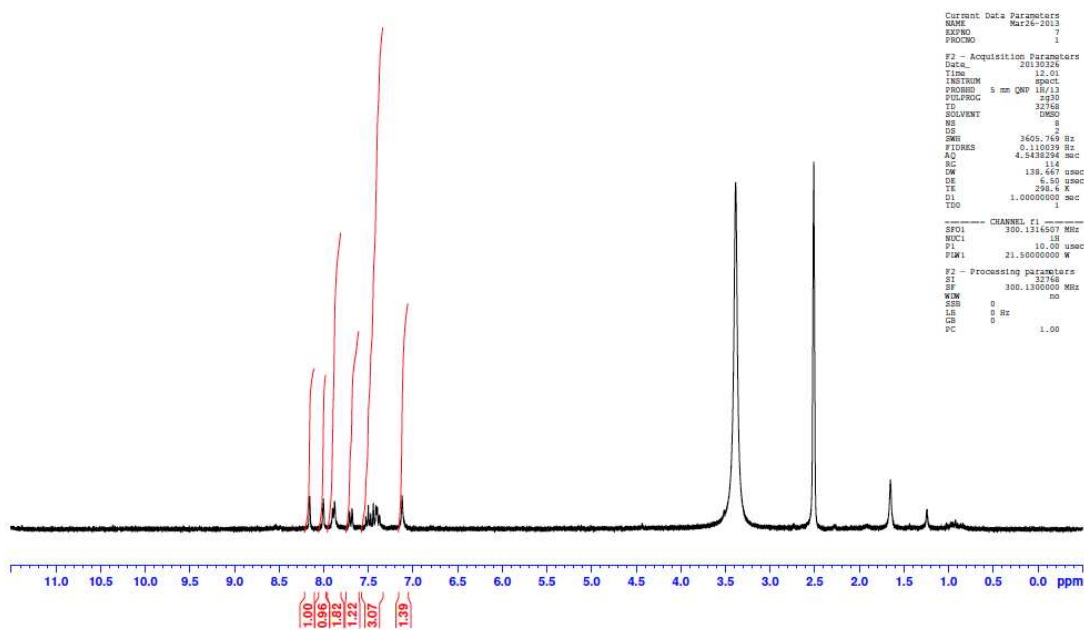


¹³C NMR spectrum of 4a

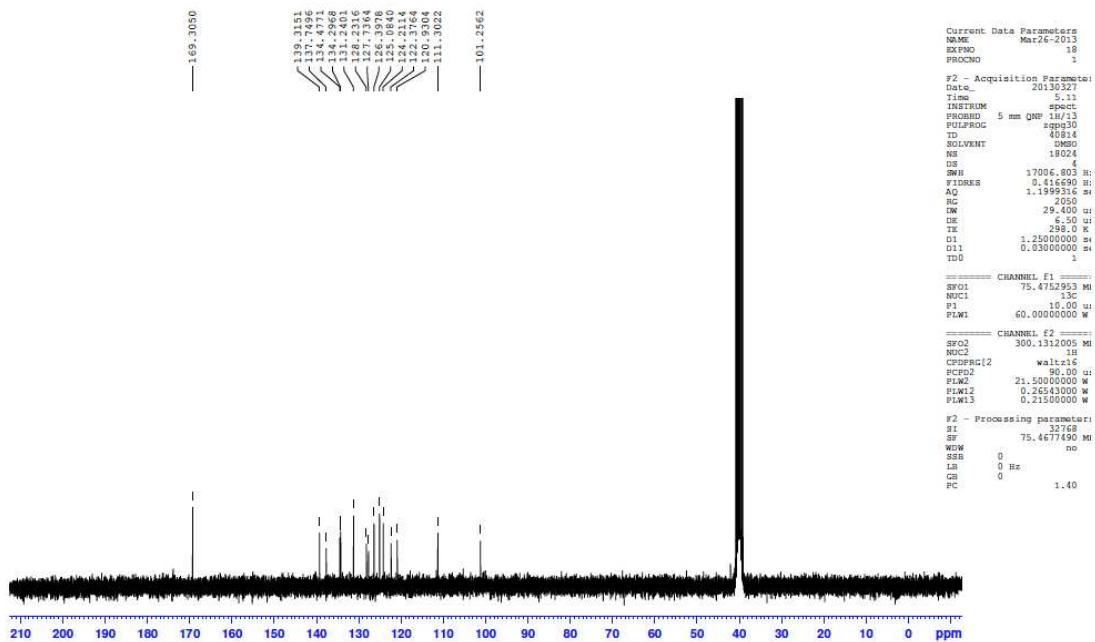


HPLC-RP: (254nm) : 99.0% : rt: 4.94

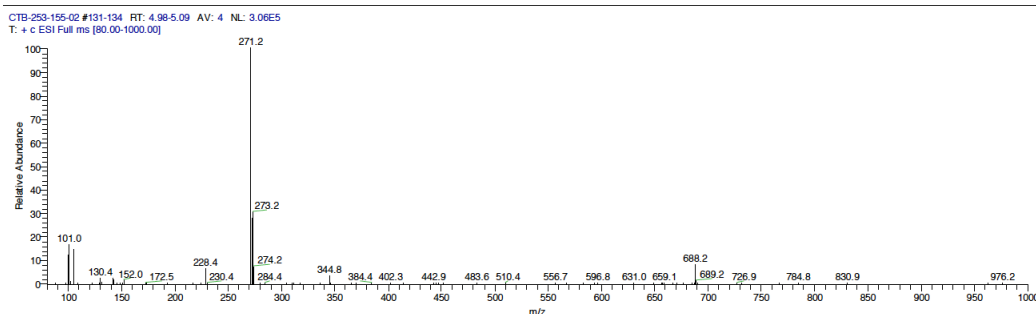
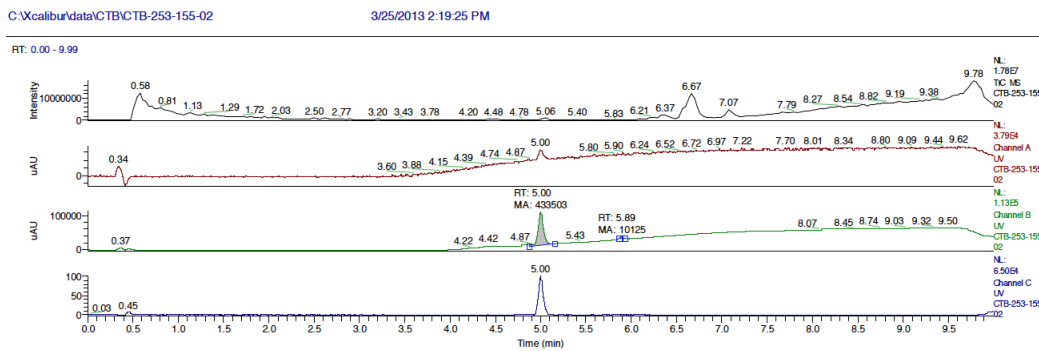
LC-MS Data of 4a



¹H NMR spectrum of 4b

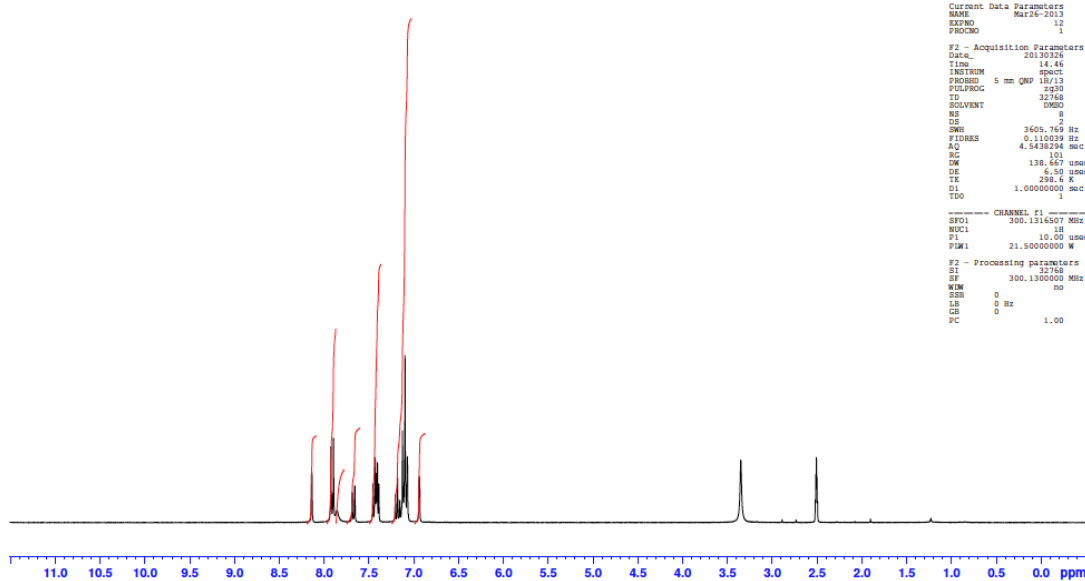


¹³CNMR spectrum of 4b



HPLC-RP: (254nm) : 98.0% : rt: 5.00

LC-MS Data of 4b



```

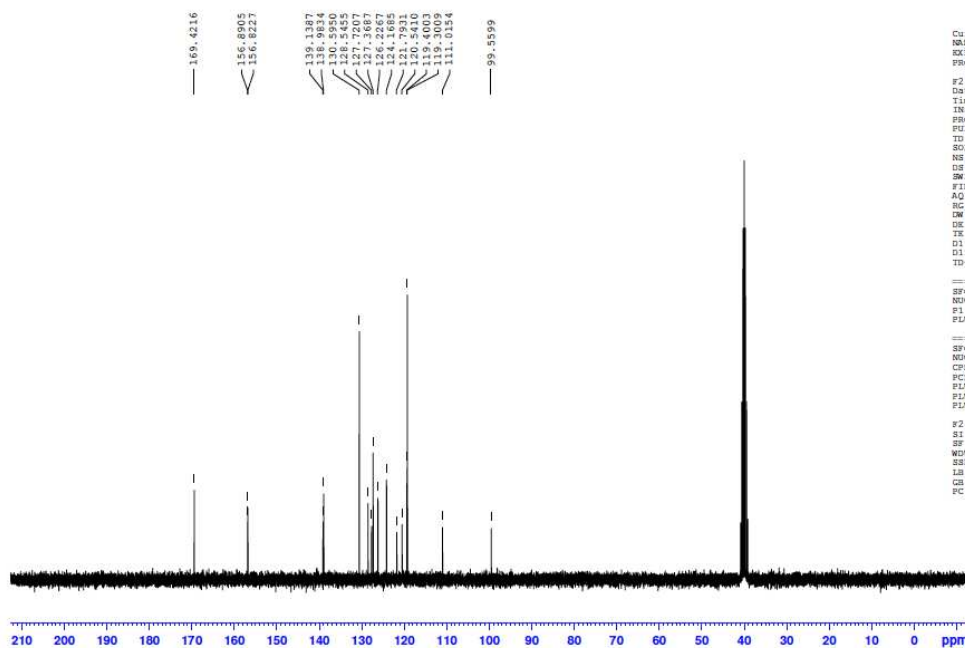
Current Data Parameters
NAME      Mar26-2013
EXPNO    12
PROCNO   1

F2 - Acquisition Parameters
Date_    20130326
Time     14.46
INSTRUM  spect
PROBHD   5 mm QNP 1H/13
PULPROG  zgpg30
TD       32768
SOLVENT  DMSO
NS       8
DS       2
SWH      3605.769 Hz
FIDRES   0.110039 Hz
AQ       4.8438294 sec
RG       101
DW       138.667 usec
DE       6.50 usec
TE       298.6 K
D1       1.00000000 sec
TDO      1

===== CHANNEL f1 =====
SFO1    300.1314697 MHz
NUC1    1H
P1       10.00 usec
P1M1    21.50000000 W

F2 - Processing parameters
SI       32768
SF       300.1300000 MHz
WDW      no
SSB      0
LB       0 Hz
GB       0
PC       1.00
    
```

¹H NMR spectrum of 4c



```

Current Data Parameters
NAME      Mar26-2013
EXPNO    13
PROCNO   1

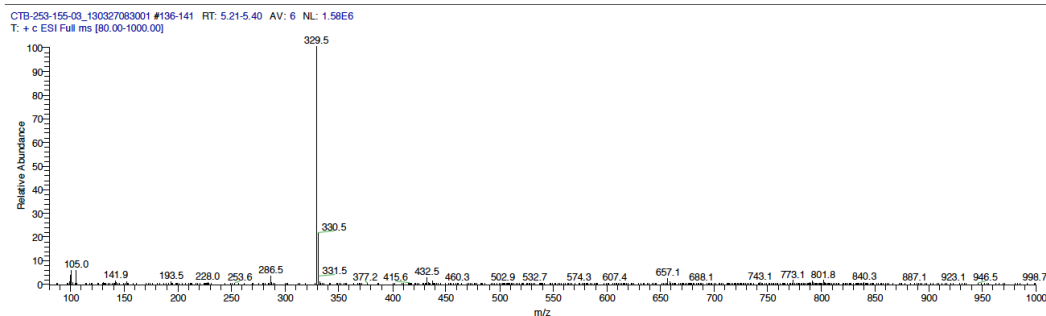
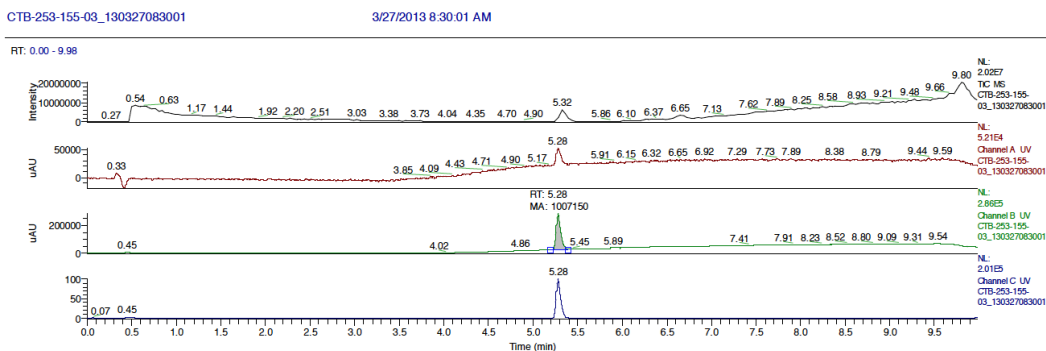
F2 - Acquisition Parameters
Date_    20130326
Time     15.11
INSTRUM  spect
PROBHD   5 mm QNP 1H/13
PULPROG  zgpg30
TD       40814
SOLVENT  DMSO
NS       493
DS       2
SWH      17000.805 Hz
FIDRES   0.416690 Hz
AQ       1.1999216 sec
RG       2000
DW       29.400 usec
DE       6.50 usec
TE       298.0 K
D1       1.25000000 sec
D11      0.03000000 sec
TDO      1

===== CHANNEL f1 =====
SFO1    75.4752863 MHz
NUC1    13C
P1       10.00 usec
P1M1    60.00000000 W

===== CHANNEL f2 =====
SFO2    300.1312005 MHz
NUC2    1H
CPDPRG2  waltz16
PCPD2    96.00 usec
PLM2    21.50000000 W
PLM12   0.26543000 W
PLM13   0.21500000 W

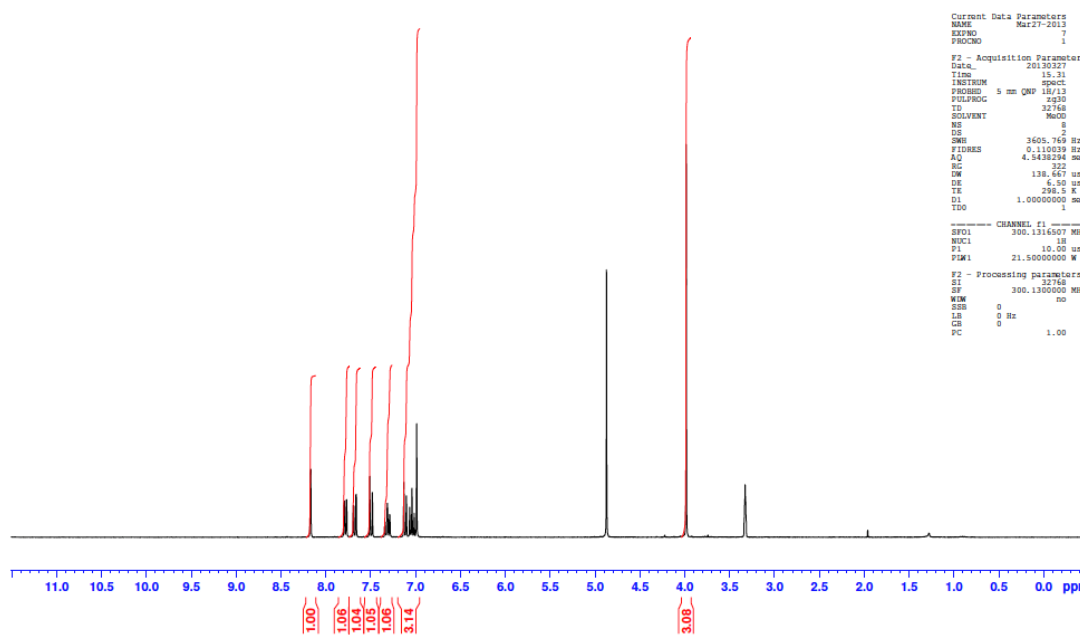
F2 - Processing parameters:
SI       32768
SF       75.4677490 MHz
WDW      no
SSB      0
LB       0 Hz
GB       0
PC       1.40
    
```

¹³C NMR spectrum of 4c



HPLC-RP: (254nm) : 99.0% : rt: 5.28

LC-MS Data of 4c



```

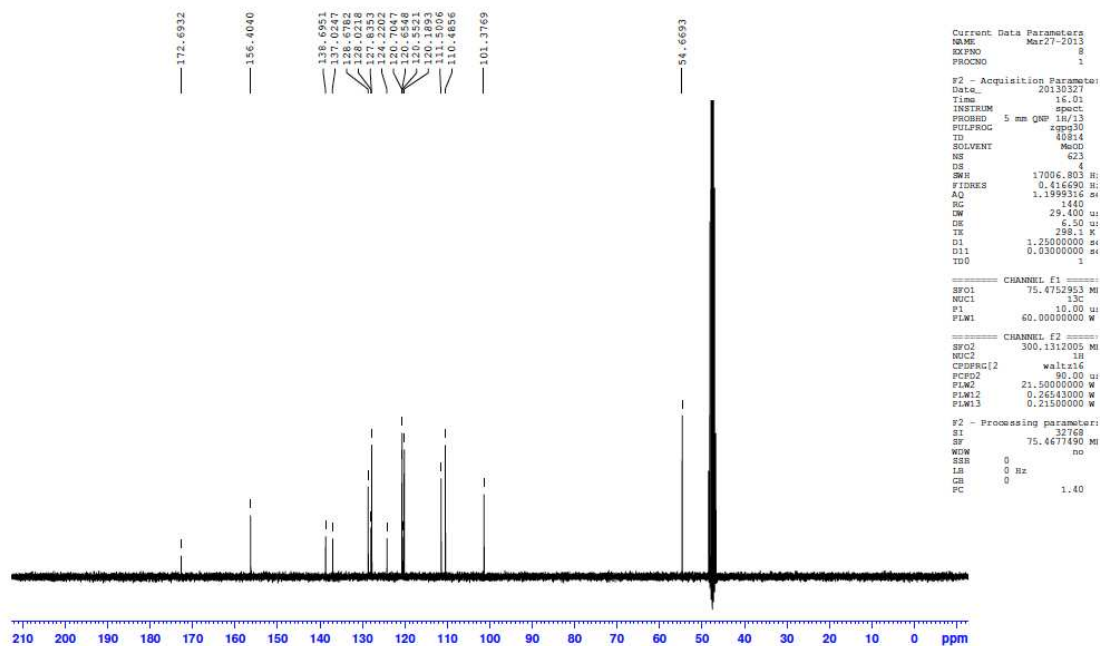
Current Data Parameters
NAME      Mar27-2013
EXPNO    1
PROCNO   1

F2 - Acquisition Parameters
Date_    20130327
Time     15:31
INSTRUM  spect
PROBHD   5 mm QNP 1H/13
PULPROG  zgpg30
TD       32768
SOLVENT  MeOD
NS       8
DS       4
SWH      3605.769 Hz
FIDRES   0.118039 Hz
AQ       4.9438294 sec
RG       322
AQ       138.667 usec
DE       6.90 usec
TE       298.2 K
D1       1.00000000 sec
TDO

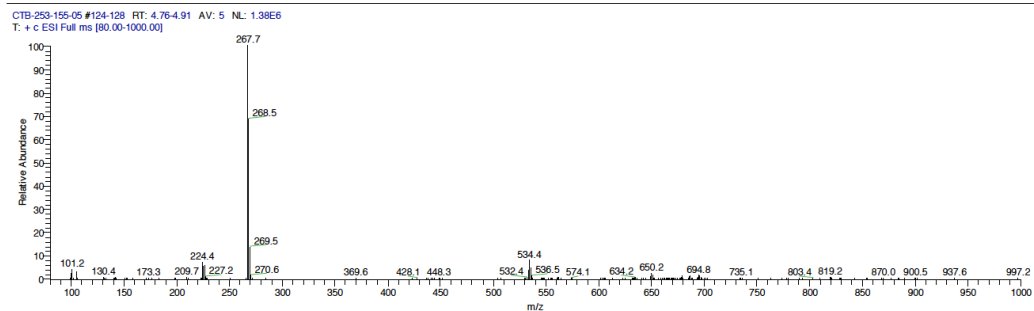
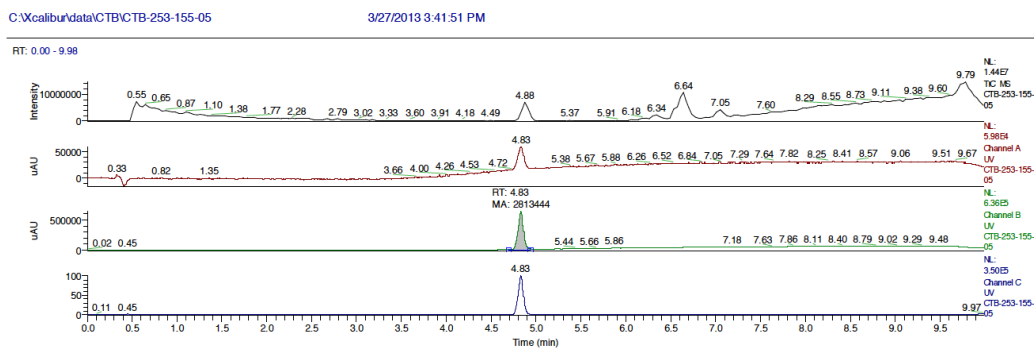
===== CHANNEL f1 =====
SFO1    300.1314507 MHz
NUC1    13
P1       10.00 usec
PLW1    21.50000000 W

F2 - Processing parameters
SI       32768
SF       300.1300000 MHz
WDW      EM
SSB      0
LB       0 Hz
GB       0
PC       1.00
    
```

¹H NMR spectrum of 4d

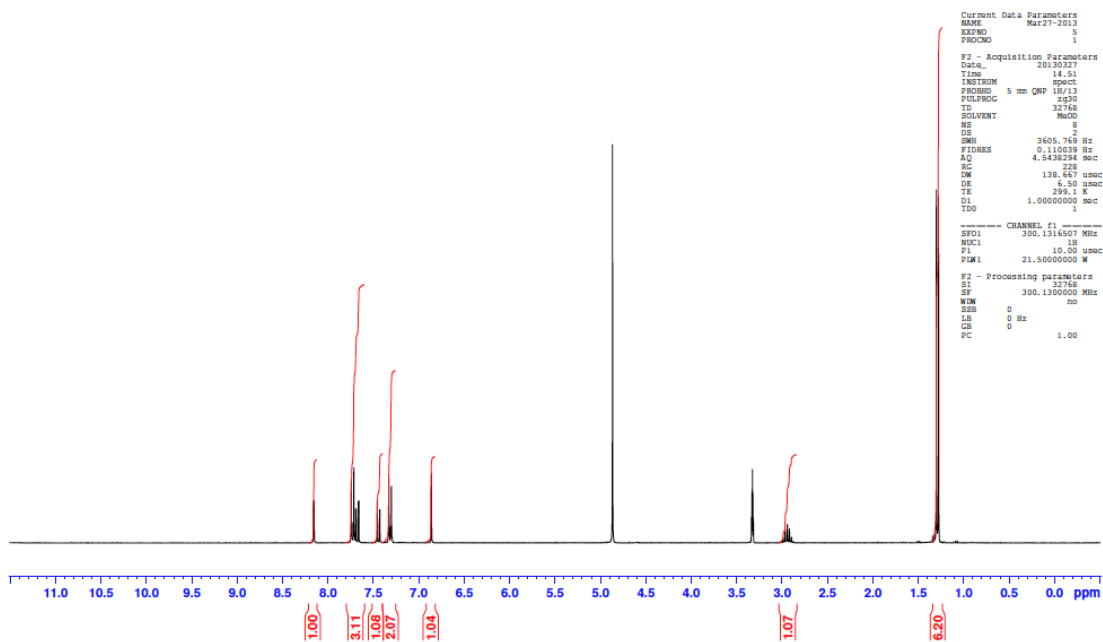


¹³CNMR spectrum of 4d

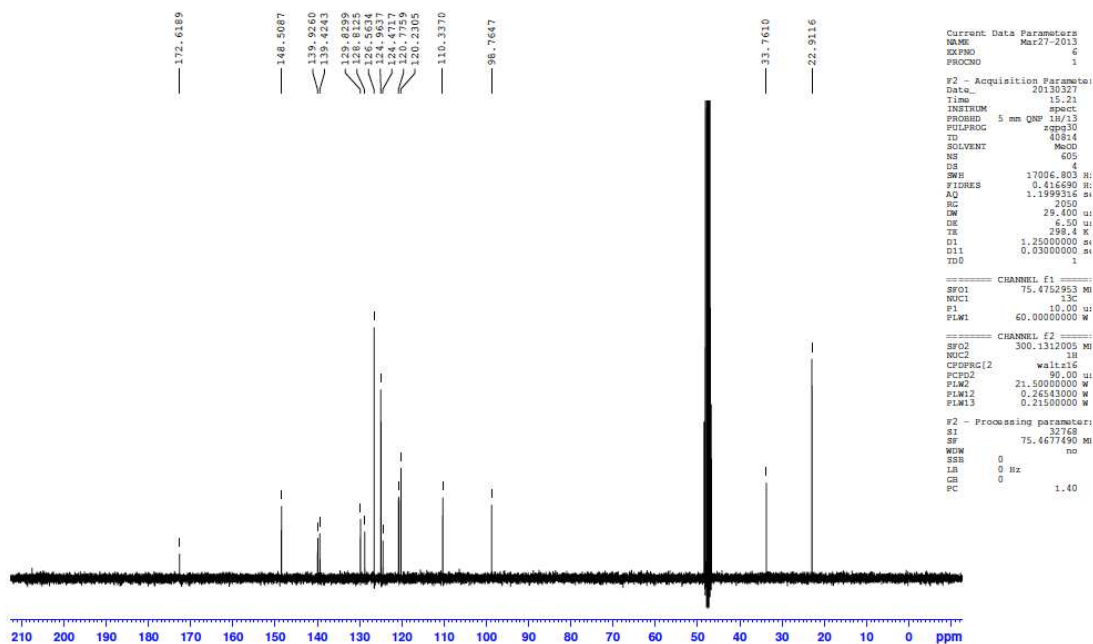


HPLC-RP: (254nm) : 99.0% : rt: 5.23

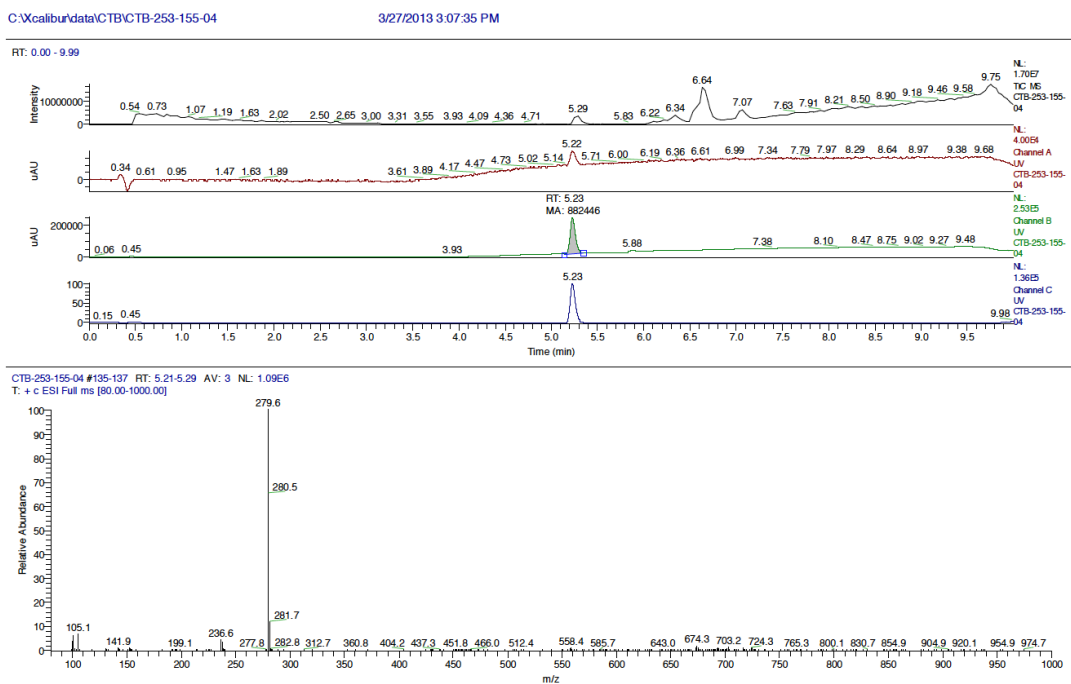
LC-MS Data of 4d



¹H NMR spectrum of 4e



¹³C NMR spectrum of 4e



HPLC-RP: (254nm) : 99.0% : rt: 5.23

LC-MS Data of 4e