



Biocide effect of *Fusarium solani* and *Penicillium spp* against the cereals cyst nematode “*Heterodera avenae*”

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ABSTRACT

Strategic Cultures by excellence, cereals are a valuable food in the world, unfortunately they are subject to various pest attacks in the fields among which cereals cyst nematodes “*Heterodera*” serious, parasite that its damage greatly reduces yields. In order to reduce its infestations and preserve the health of the consumer, we tried to use biological control using two cultures filtrate of fungus namely *Fusarium solani* and *Penicillium spp* against the nematode. After exposing the eggs to various concentrations of the two filtrates for a period of 6, 24 and 48 hours, the results are highly significant and shows the ovicidal effect of the two filtrates. This effect increases with the immersion time and the concentration of the two filtrates. A mortality rate of eggs of 93.62% and 82.45% is recorded respectively for *F. solani* and *Penicillium spp* after 48h of exposure. By comparing the efficiency of the two filtrates cultures on nematode, the results revealed that the ovicidal action of pure filtrate of *F. solani*, reached 94.39% after 48 hours against (83.39%) for the pure filtrate of *Penicillium spp*. So, the culture filtrate of *F. solani* is much more effective than the filtrate of *Penicillium spp*. Also our experimentations have shown that nematode eggs are very sensitive to low pH (3.5).

Key words: Fungi, *Heterodera avenae*, inhibitory effect.

INTRODUCTION

Cereals are considered the most cultivated cultures by man and have long been prominent in human nutrition. Although these cultures have benefited from some development programs but national production, did not know the expected improvement. The causes of this are mainly of technical orders, and phytosanitary. Of these, nematode induce 10% loss of global production [38]. For cereals, the complex *Heterodera avenae*: cereal cyst nematode is by far the most to fear. It is present in temperate cereal regions, [25-26, 32]. This wide distribution follows the expansion of cereal surfaces. The *Heterodera* complex is represented by twelve species in the world. However, the specific composition varies from one region to another. In Algeria, *Heterodera* group was the subject of several investigations, the results of which revealed the presence of several species in cereal plots, [24, 30]. There the use of synthetic nematicides were unsatisfactory in addition to their adverse effects on the flora and fauna that have led to their reduction. Thus, new solutions must be found quickly to solve the nematode problems. Among these means, the use of natural antagonists including fungal metabolites are an interesting way forward, easy to use and environmentally friendly. In this concordance many fungi are known to produce nematicidal compounds or nematostatics [3-4, 6, 15, 21, 23]. Thus secondary metabolites of *Fusarium oxysporum* are toxic to *Meloidogyne incognita*. So many toxins of *Fusarium spp*, reduce the viability of nematodes. [8, 3, 28]. Other studies have shown the inhibitory action of hatching eggs or Nematicide from some fungi isolated from oil nematodes and plants [1, 7, 5, 10, 15, 22, 27, 32, 35-36]. Our contribution aims to highlight the effect of toxic and biocide filtrate of cultures of *Fusarium solani* and *Penicillium spp* by bio-assays sustainability of *Heterodera avenae* eggs.

EXPERIMENTAL SECTION

The populations of the cereal cyst nematode "*Heterodera avenae*" were taken in cereal plots in Mascara region (western Algeria). Soil samples thus recovered were dried prior to the extraction of cysts. For this, we used the technique FENWICK (1940), based on the floating cysts. This process involves separating the cysts of the mineral fraction of the soil. recovered cysts will be subject to a morphometric and anatomical study (microscopic observation of the perineal region refers to vulva, cone and the constituents of the structure such as the presence or absence of the bullae and/or bridge, the shape of fenestrae and the length of the vulvar slit). Both fungi used in this case *Fusarium solani* and *Penicillium spp* do not grow on cereals, They were obtained from the Laboratory of Plant Pathology. After incubation, the mycelial mass *Penicillium spp* appears, on which we collected cuttings for aseptically drop the center of a Czapeck cast medium in Petri dishes. The cultures were then incubated at 28 °C in an oven, in order to obtain pure cultures. Successive subcultures of the previous crop are carried again on the Czapeck medium. This strain is stored in a tube containing Czapeck medium set at a temperature of 4 °C. Regarding the *Fusarium solani*, we realized a culture by taking a small sample of the mycelial mass of pure culture we file into a tube containing 10 ml of sterile distilled water. When adjusted, the solution should contain about 100 spores per ml. A drop of this solution (after stirring) is filed under sterile with a Pasteur pipette on a PDA cast medium in Petri dishes with sterile talc. This drop is spread across the surface. After 48 h of incubation at 28 °C, the spores which have developed a mycelium, are transplanted one by one on a PDA medium [19].

Pathogenicity test:

Almost the saprophytic *Fusarium* lives in the soil and to differentiate them from parasitic forms, morphological appearance is not enough. Both forms do not show any differences that may be distinguished from each other. To do this, experiment by inoculation is necessary [11]. Thus *Henzet* tomato variety is sensitive to the strain of *Fusarium solani*. Three weeks after planting, the young tomato plants (stage appearance of the first two true leaves) are dug; their roots were washed with water, then for 15 minutes in a mistaken conidia suspension from an old culture 4 days obtained by uniform spreading on the surface of the medium in a Petri dish. The witnesses are mistake in distilled water for the same duration. The inoculated seedlings were transplanted in pots of 30 cm diameter containing potting soil. As of three tomato plants is used to inoculate each strain, [17].

Estimation of symptoms:

External symptoms are estimated twice per week during two to three weeks after inoculation [18]. In vitro tests we have done are that described by [12, 16].

Methods of filtrate cultures: A cutting taken from the mycelial growth area of a culture of the antagonist is placed in flasks of 500 ml Erlenmeyer containing 300 ml GYP liquid nutrient medium. These flasks were incubated for 3 weeks in the dark at 26 °C. After this period, the mycelium is separated from the culture medium by filtration on different diameters (11, 12, 5, and 13), which avoid the passage of all contaminating spores

pH measurement

The pH is measured before the incubation of the fungus and adjusted to 5.8. Then the acidic effect was tested with the filtrate then diluted in sterile distilled water to P/2, P/4 and P/8. The pH of the two filtrates of the fungus is measured. Two control media are used for comparison, one is sterile distilled water, the other is the non-inoculated culture medium, Adjusted to the same pH as the test filtrate.

Test of filtrate on *H.avenae* eggs.

Eggs are deposited in hemolysis tubes, each tube receives about 300 eggs which we added a droplet of liquid tested 100 µl. Each treatment was repeated 5 times. The toxicity of the filtrate was estimated by the mortality percentage after 6, 24 and 48 hours of immersion and after coloring eggs with new blue-R [13]. Dead eggs are dark blue and viable eggs remain colorless. All tests are carried to ambient temperature of laboratory.

Statistical analysis

All the results obtained are subjected to the treatment of the analysis of the variance to a criterion of classification for all the parameters study with the use of software Statist, version 5.

RESULTS AND DISCUSSION

Heterodera avenae: is a species often found on cereals it is characterized by the presence of dense and massive bullae, two semi-fenestrae widely separated by the vulva which has a length less than 20 µm. This last is in the vulvar bridge which is free of ramification. In this study, we confirmed the presence of this species in several cereal plots in the Mascara region (western Algeria).

Study of pathogenicity test:

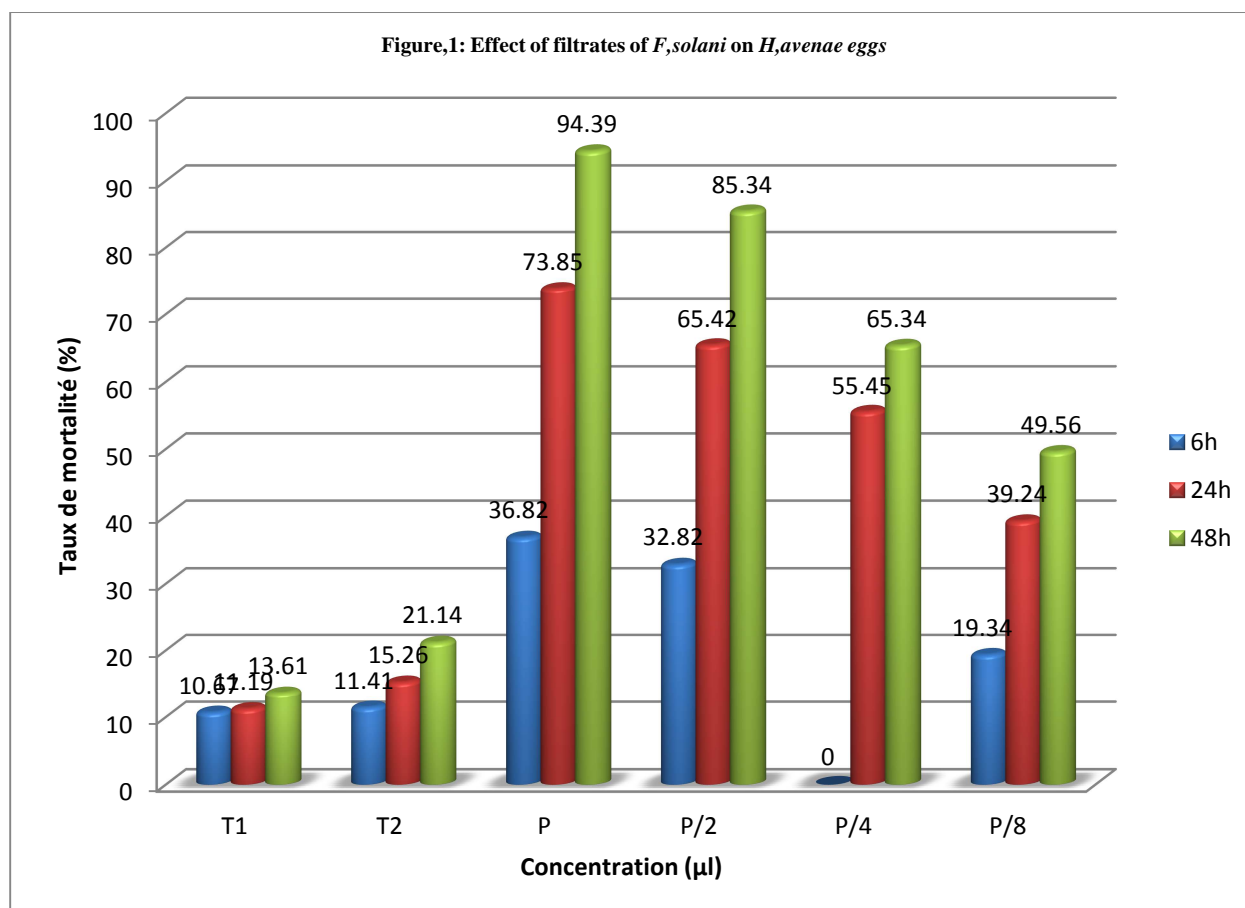
Pathogenicity of *Fusarium solani* is estimated using external symptoms expressed as follows: Healthy plant and diseased plant. Thus, after three weeks of incubation, it appears that our strain used is non-pathogenic for tomato. If we compared pots inoculated with the witness, we find that all plants are identical to the control plants. To confirm the absence of the fungus in the plant, we seeded small fragments of plants inoculated on PDA. After 7 days of incubation at 28 °C, microscopic observation revealed the absence of *Fusarium solani* within the plant. The use of saprophytic strains is necessary to avoid the risk of having additional damage to the plant. Therefore our strain is a saprophyte for the tomato. We appreciate the influence of aging on the pathogenicity of culture of *F. solani*. For the strain *Penicillium spp.*, used in this test, it is also a saprophyte for tomato.

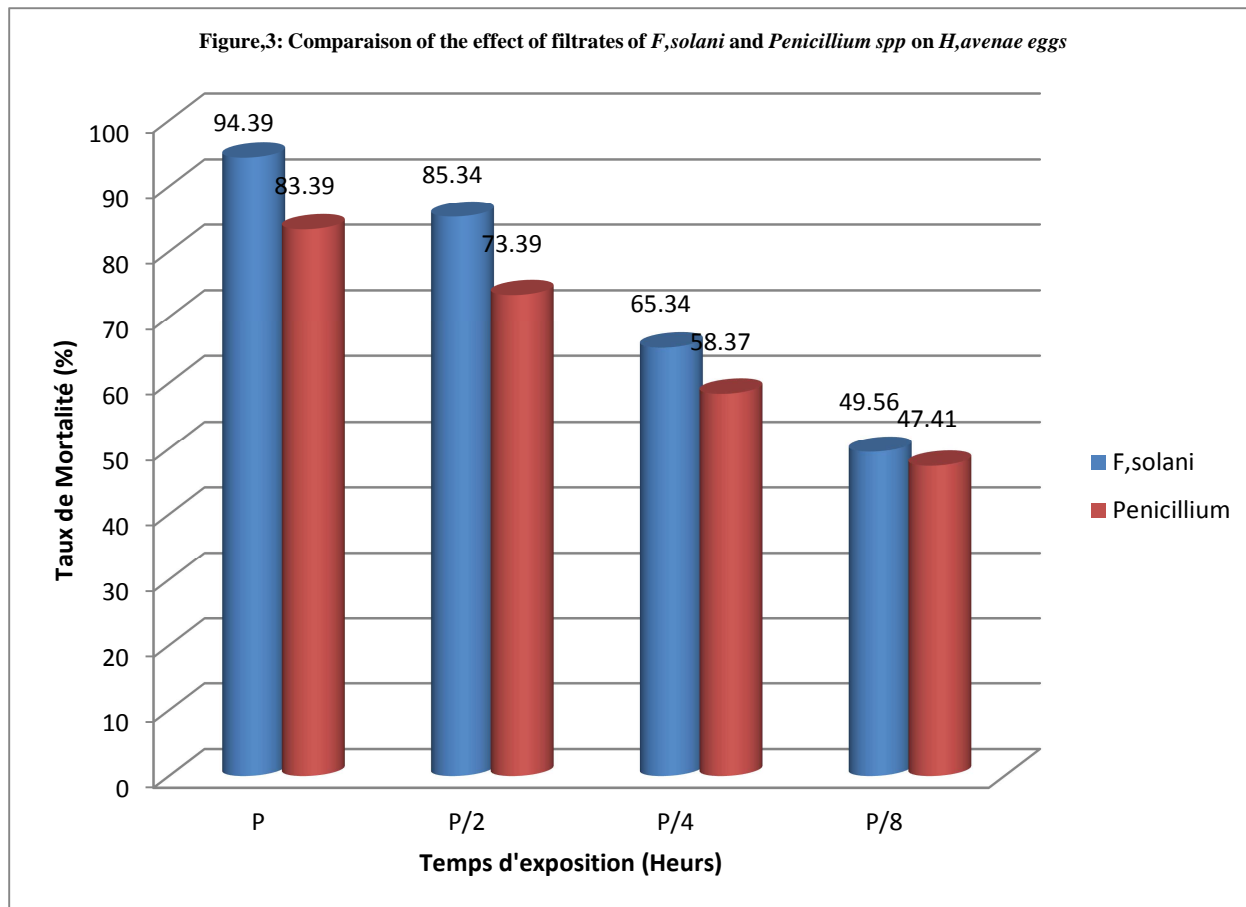
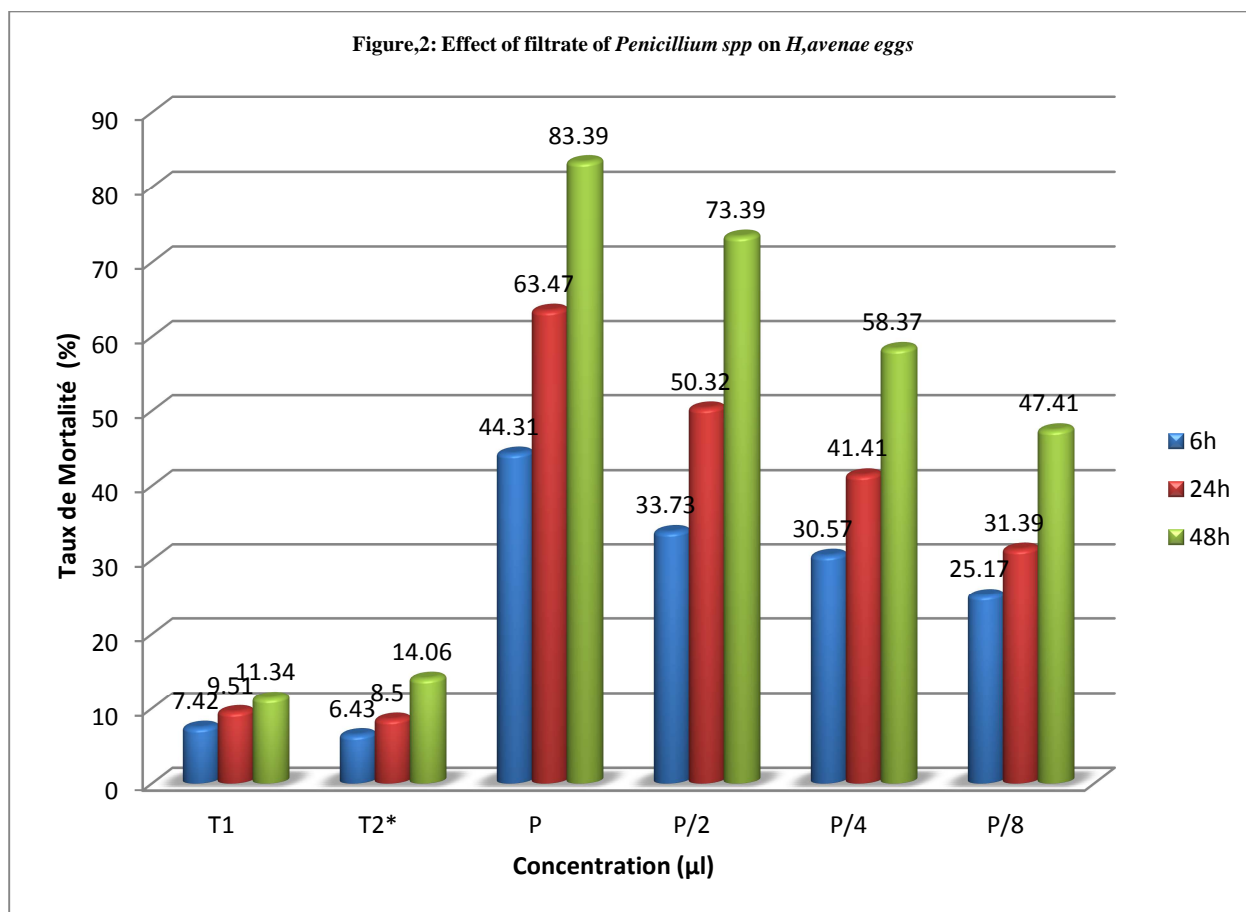
Effect of filtrate culture of *F. solani* on egg of *Havenae*: According to the results (Figure 1), we find that the percentage of dead eggs increases with the concentration of the filtrate and the immersion time. After 6 hours of immersion, there has been a death rate of 35.33% with pure filtrate of *F. solani*. This rate reached 72.20% after 24 hours and 100% after 48 hours. Regarding the filtrate diluted to 50% (P/2), the mortality rate increases with the egg immersion time (85.34%) after 48 hours. Even at lower dilutions (P/8), the filtrate is still effective, the mortality percentages are 19.34, 39.24 and 49.56%, respectively, after 6 hours, 24 hours and 48 hours of immersion. These results are highly significant ($F_c=31.59$ and $F_{th}=2.32$) compared to controls after 48 hours of immersion (distilled water: 13.61%) and (uninoculated medium: 25.14%).

Effect of filtrate culture of *Penicillium spp* on egg of *H. avenae*:

The results of the *Penicillium spp* effect on egg of *H. avenae* are mentioned in the (Figure 2). And since the first hour of immersion (6h), pure filtrate was effective with a mortality rate of 44.31%. This value reaches 83.39% after 48 h, very significant result with ($F_c=55.72$ and $F_{th}=2.32$). Even with dilutions (P/2, P/4, and P/8), the filtrate remains effective against egg viability. Thus the inhibition rate of 73.39% is achieved after 48 hours under the effect of the lowest dilution (P/8). Compared to the two witnesses with whom we recorded weak inhibition rate 10.24 and 13.49% respectively for distilled water and uninoculated medium.

Our results are consistent with those of [8, 3, 28], which highlighted the toxicity of secondary metabolites produced by endophytic fungi such as *Fusarium oxysporum* on *Meloidogyne incognita* and other toxins from various *Fusarium sp.*, which can reduce the viability of nematodes.





	T1	T2	P	P/2	P/4	P/8
6h	10.67	11.41	36.82	32.82	32,8229,69	19.34
24h	11.19	15.26	73.85	65.42	55.45	39.24
48h	13.61	21.14	94.39	85.34	65.34	49.56

	T1	T2*	P	P/2	P/4	P/8
6h	7.42	6.43	44.31	33.73	30.57	25.17
24h	9.51	8.5	63.47	50.32	41.41	31.39
48h	11.34	14.06	83.39	73.39	58.37	47.41

	F,solani	Penicillium
P	94.39	83.39
P/2	85.34	73.39
P/4	65.34	58.37
P/8	49.56	47.41

The effect of pH on the mortality of eggs:

According to the results, we note that the low pH (3.5) for the witness (T2) influenced the mortality of *H. avenae* eggs under the action of *F. solani* after 48 hours, by registering a mortality rate of 21.14%, but by pH=5 (T2) under *Penicillium* spp, we noticed a very small effect on the viability of the eggs (mortality rate after 48 hours was 14.21%), this rate is nearly similar to that obtained under the effect of the distilled water (11.34%). Our results agree with those of [6], who showed that the culture filtrates are toxic on nematodes at a lower pH.

Comparison of the effect of the two filtrates: The two filtrates culture of *F. solani* and *Penicillium* spp. exhibit a prominent toxicity on *H. avenae* eggs. This toxicity increases with both the immersion time and with the different concentrations (P, P/2, P/4 and P/8).

The ovicide action of pure filtrate of *F. solani*, reached 94.39% after 48 hours against (83.39%) for the pure filtrate of *Penicillium* spp. at lower dilutions P/8, the two filtrates also showed efficacy with inhibitory rate of hatching (49.56 and 47.41%) respectively for the culture filtrate of *F. solani* and *Penicillium* spp. Eggs of *Heterodera* appear to be more sensitive to culture of *F. solani* filtrate but statistically there is no significant difference ($F_{cal}=0.12$ and $F_{th}=4.19$) (Figure 3).

CONCLUSION

The phytoparasitic nematodes have a very significant economic impact on a global scale, much more in developing countries than industrial countries. Considering their extreme resistance, due to their great physiological variability and the fact that they are soil pests, it is very difficult to combat them. In this context, the nematode problems will not be solved by the adoption of a single method but by the combination of all available means used together: crop rotation, resistant varieties, nematophagous fungi, mycorrhizal plants, nematicides plants, organic amendments... That set of measures can only replace the destructive performance offered by synthetic nematicides. This is why research work on other natural nematicides molecules has helped to identify toxic substances produced by fungi that can act on nematodes. These substances are present in the filtrates.

In that regard our test allowed us to confirm the high significant efficacy of the two filtrates on *H. avenae* eggs. Ovicide action after 48 hours of immersion reached 94.39% and (83.39%) respectively for pure filtrate of *F. solani*, and pure filtrate of *Penicillium* spp. All the results obtained show that it is now possible to consider a biological control against cyst nematodes "Heterodera" through *F. Solani* and *Penicillium* spp while respecting certain precautions: Evaluation of the growth of the fungus so that they have time to secrete substances metabolite and to be effectively against the eggs of phytophagous nematodes. Regarding soil

protection, we are moving toward the use of saprophytic fungi (*F. solani* and *Penicillium spp*). Then the soil pH should be below the pH 5. The safeguarding of the food value of this invaluable food product is very important thus any attempt at study in this field by the use of natural substances and fungi filtrates is very justified.

REFERENCES

- [1] VSB Alcantara; JL de Azevedo, *Phytopatologia Brasileira.*, **1980**, 5, 201–206.
- [2] HH Ameen, *Bulletin of Faculty of Agriculture, University of Cairo.*, **1991**, 42, 963-970.
- [3] H Anke; O Sterner, *Current Organic Chemistry.*, **1997**, 1, 361-374.
- [4] H Anke; M Stadler; A Mayer; O Sterner, *Canadian Journal of Botany.*, **1995**, 73, S932-S939.
- [5] S Caroppo; B Perito; O Pelagatti, *Redia.*, **1990**, 73, 451–462.
- [6] SY Chen; DW Dickson; DJ Mitchell, *Journal of Nematology.*, **2000**, 32, 190-197
- [7] A Ciancio; A Logrieco; F Lamberti; A Bottalico, *Nematol. Mediterranea.*, **1988**, 16, 137–138.
- [8] A Ciancio, *Fundamental and Applied Nematology.*, **1995**, 18, 451-454.
- [9] MJN Costa; VP Campos; LH Pfenning; DF Oliveira, *Nematol. Brasileira.*, **2000**, 24, 219-226.
- [10] JS Dahiya; DP Singh, *Plant and Soil.*, **1985**, 86, 145–146.
- [11] P Davet ; F Rouxel, Eds, INRA, Paris, France., **1997**, 194.
- [12] JC Cayrol, *PHM Revue Horticole.*, **1989**, 293, 55-58.
- [13] Shepherd, **1962** in DeGuiran, G., **1979.** *Rev.Nématology.*, **1979**, 2, 65-77.
- [14] C Djian; L Pijarowski; M Ponchet; N Arpin; J Favre-Bonvin, *Nematologica.*, **1991**, 37, 101-112.
- [15] J Hallmann ; RA Sikora, *European Journal of Plant Pathology.*, **1996**, 102, 155-162.
- [16] J Henni, *Cryptogamie, Mycologie.*, **1987**, 8 (3), 203-207.
- [17] J Henni ; C Boisson ; JP Geiger, *Phytopathol. Mediterr.*, **1994a**, 33, 51-58.
- [18] J Henni ; C Boisson ; JP Geiger, *Phytopathol. Mediterr.*, **1994b** 33, 10-16.
- [19] A Johnson; C Booth, Commonwealth Mycological Institut. Kew, Surrey., **1983**, 439p.
- [20] TA Khan, *Bionotes.*, 1999, 1, 38-39.
- [21] B Köpcke; D Wolf; H Anke; O Sterner, British Mycological Society International Symposium, Bioactive Fungal Metabolites .Impact and Exploitation, UW Swansea, UK, 22-27 April **2001**, 72. [Abstr.]
- [22] A Mani; C L Sethi, *Nematropica.*, **1984**, 14, 121–129.
- [23] SLF Meyer; SI Massoud ; DJ Chitwood ; DP Roberts, *Nematology.*, **2000**, 14(2), 871-879.
- [24] A Mokabli, Thèse Doctorat. INA. El Harrach. Alger., **2002**.
- [25] JM Nicol., **2002.** in BC Curtis; S Radjaram; G Macpherson, FAO Series, Rome, Italy., **2002**, 345-366
- [26] J Nicol; R Rivoal; S Taylor; M Zaharieva, *Nematology Monographs and Perspectives.*, **2003**, 2, 1-19.
- [27] JK Nitao; SLF Meyer; D Chitwood, *Journal of Nematology.*, **1999**, 31:172–183.
- [28] JK Nitao; SLF Meyer; WF Schmidt; JC Fettinger; DJ Chitwood, *Journal of Chemical Ecology.*, **2001**, 27, 859-869.
- [29] N Randhawa; P Singh; KS Sandhu; A Bhatia, *Plant Disease Research.*, **2001**, 16, 280-282.
- [30] K Righi ; A Mokabli ; F Assia Righi ; H Gourari, *Actes du Séminaire International sur la Protection des végétaux, ENSA. El Harrach.*, **2011**, 97-102.
- [31] R Rivoal; R Cook, in K Evans; DL Trudgill; JM Webster. Wallingford, UK, CAB International., **1993**, 259–303.
- [32] PK Sakhuja; I Singh; SK Sharma, *Indian Phytopathology.*, **1979**, 31, 376–377.
- [33] DD Sharma, *Indian Journal of Sericulture.*, **1999**, 38, 152-154.
- [34] AM Shepherd, **1962**, in G De Guiran, *Revue of Nématology.*, **1979**, 2, 65-77.
- [35] KP Singh; K Bihari; VK Singh, *Current Nematology.*, **1991**, 2, 9–14.
- [36] K Walia; G Swarup, *Indian Journal of Nematology.*, **1985**, 15, 174–176.
- [37] LF Wang; BJ Yang; CD Li, *Scientia Silvae Sinicae.*, **1999**, 35, 41-47.
- [38] AG Whitehead. Plant nematode control, CAB International, Wallingford, England, **1998**, 384pp