



Review Article

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## Role of Nanocarriers for Drug Delivery to Brain

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### ABSTRACT

*Noninvasive treatment of Neurological diseases is limited because of poor transport of drug molecules into brain. Almost all drugs and therapeutic agents do not reach to brain due to presence of defensive barrier of brain such as Blood Brain Barrier and Blood – Cerebrospinal Fluid Barrier. Therefore main challenge for CNS drug delivery is how to maintain therapeutic concentration of drug molecules in brain. Nanotechnology gives promising solutions to overcome these obstacles. Different nanocarriers such as Liposomes, Solid lipid nanoparticles, Micelles have been studies for the drug delivery across the brain. Many nanoformulations can transport drug molecules into brain in in vitro and in vivo models. Many studies have been done to access the nanotechnology against CNS disorders such as brain tumour, Alzheimer's disease and acute ischemic stroke. In future nanaocarriers will be promising alternative to increase brain drug concentration using novel drug delivery systems which will improve Blood brain barrier permeability and reduce their neurotoxicity.*

**Keywords:** Nanocarriers; CNS; Brain drug delivery; Blood brain barrier

### ROLE OF NANOCARRIERS FOR DRUG DELIVERY TO BRAIN

Brain is most vital organ of body controlling all the actions. Blood Brain barrier is defense biological barrier which restricts the passage of toxins and pathogens but it also restricts the activity of drug molecule. For a drug to achieve its maximum activity it should cross blood brain barrier. To overcome these barrier different approaches has been made to increase brain drug concentration.

The CNS barriers protect the brain from invading pathogens, neurotoxin molecules and circulating blood cells. These barriers are Blood Cerebrospinal fluid barrier, Blood brain Barrier, Blood retinal barrier and blood spinal cord barrier [1]. Drug delivery to brain is limited due to presence of Blood Brain Barrier (BBB). Blood brain barrier restricts the passage of therapeutic agents, neuropeptides to brain. Many Drug Delivery methods have been developed for CNS diseases targeting but mostly are invasive and lack target specificity. To cross the blood brain barrier many types of nanocarriers systems has been made such as linear polymers, hyper branched polymers, dendimers, liposomes and micelles. Novel administrations has also been approached which includes temporary disruption of BBB to increase permeability, local drug administration by using impregnated polymers; Convection enhanced delivery and intranasal delivery. In this review brief introduction to BBB and role of nanocarriers for drug delivery to brain will be discussed.

#### Blood Brain Barrier

This is anatomic barrier of brain which is developed by coordinated function of multiple cell types to restrict passage of harmful substances to brain and maintains intra cranial pressure. BBB is formed by micro vascular endothelium, basement membrane and glial cells such as pericytes. Intraluminal space of brain capillaries are lined by monolayer of microvascular endothelial cells and closely packed endothelial cells forms the tight junctions limiting substances crossing the BBB [2]. Pinocytic vesicles and high number of mitochondria transport certain

molecules between blood and brain. BBB transport essential protein which are necessary for the brain function. Glucose transport (GLUT) system is essential for brain for Glucose gives proper energy and helps to function it properly. Modification of therapeutics molecules are done to be recognized by GLUT. Multiple Drug resistance Protein (MDR1) is another important transport system of Brain which restricts the transport of certain compounds in the brain [3]. To Cross the BBB, properties of barriers, permeability properties of brain endothelial cells and molecular weight of compound are important parameters. There are mainly four mechanisms of BBB transport [2].

#### Simple diffusion:

Transport through from high to low concentration.

#### Facilitated diffusion:

Type of carrier mediated endocytosis in which solute binds to specific membrane protein same like simple diffusion from high to low concentration.

#### Simple diffusion through an aqueous channel:

Formed within the membrane and mainly charged ions transport.

#### Active transport via protein carrier:

This is active transport occurred by a protein carrier having specific binding site that go through affinity changes. Concentration gradient is responsible for solute transport.

BBB is formed by cerebral endothelium, Blood – CSF barrier is comprised of choroid plexus and CSF-Blood barrier formed by avascular arachnoid epithelium which lies under dura and encloses the brain [4]. Anatomical and Physiologic functions of these barrier controls the toxicity and normal brain functions by regulating concentration and clearance of endogenous and exogenous molecules [5]. Brain endothelial has cells have tight junction complexes that restrict passage through BBB. Capillaries and endothelial cells occupy 1% and 0.1% of brain volume respectively, Surface area of brain microvasculature is  $\sim 20 \text{ m}^2$  and total length is  $\sim 400$  miles. The distance between brain capillaries is  $\sim 40 \text{ }\mu\text{m}$  and brain cells are at  $20 \text{ }\mu\text{m}$  from capillaries which forms spaces that allow small molecules to cross them [6]. But a feature of BBB does not allow molecules to pass through them. Hydrophilic compounds having mass lower than 150 Da and highly hydrophobic compounds having mass lower than 400-600 Da can cross the membrane by passive diffusion. Major Barriers of BBB are listed in Table 1 [7,8].

**Table 1: Barriers of BBB**

|                    | Elements                                                        | Role                                                                                               |
|--------------------|-----------------------------------------------------------------|----------------------------------------------------------------------------------------------------|
| Anatomical Barrier | Tight interendothelial junctions                                | Restricting free exchange of solutes and cells between blood and CNS                               |
|                    | Absence of fenestrations and low number of pinocytotic vesicles |                                                                                                    |
|                    | Luminal glycocalyx                                              |                                                                                                    |
| Transport Barrier  | Diffusion pathways                                              | Supply of sugars, amino acids, lipids, vitamins, minerals, metabolic precursors, peptides, protein |
|                    | Solute Carriers                                                 |                                                                                                    |
|                    | Efflux Pumps                                                    |                                                                                                    |
|                    | Adsorptive and receptor mediated transendothelial transport     |                                                                                                    |
| Metabolic barrier  | Phase 1 and 2 enzymes                                           | Protection from bioactive molecules                                                                |

P- Glycoprotein is drug transport protein which is ATP – dependent located at apical membranes of different epithelial cells which forms BBB [9]. Presence p-gp can restricts the activity of many drugs in brain, including digoxin, dexamethasone, vincristine, taxanes, cyclosporine etc. [10]. For drug transport across the BBB primary physiological pathways which needs to be utilized are listed in Table 2 [8].

**Table 2: Primary physiological pathways**

| Pathway                         | Transfer of                                                    | Methods to Enhance                                                       |
|---------------------------------|----------------------------------------------------------------|--------------------------------------------------------------------------|
| Lipid – mediated diffusion      | Lipophilic small molecules                                     | Increasing lipophilicity                                                 |
| Carrier – mediated transport    | Ligands of Carriers : amino acids, glucose and other nutrients | Development of ligand analogues                                          |
|                                 |                                                                | Conjugation of drugs to ligand- targeted nanoparticles                   |
| Receptor – mediated transport   | Peptides, proteins                                             | Drugs or nanoparticles conjugates to peptide or protein vectors          |
| Adsorptive – mediated transport | Serum Protein                                                  | Drugs or nanoparticles conjugated to cationic protein or peptide vectors |

When neurological disease occurs, properties of BBB changes and these changes can be utilized for passage of drug molecules which are not possible in healthy brain. BBB functioning alters by oxidative stress, inflammatory mediators, lipid mediators, vasogenic agents, infective agents as well as physiological and immunological stimuli [11]. Table 3 contains the pathological conditions their action on BBB and how these changes can be utilized to transport the drug to brain [12].

**Table 3: CNS disorders and their effect on BBB**

| Pathological conditions | Influence on BBB                                                                                                                                        | Effects for Drug crossing BBB                                                                                  | References |
|-------------------------|---------------------------------------------------------------------------------------------------------------------------------------------------------|----------------------------------------------------------------------------------------------------------------|------------|
| Multiple Sclerosis      | Disruption of Tj, Enhanced leukocyte activity, release of inflammatory cytokines/chemokines                                                             | It may enhance paracellular transport of drugs                                                                 | [13-15]    |
| Alzheimer's Disease     | BBB disruption and allowed the greater access of peripheral IgG to CNS                                                                                  | Potentially it may enhance paracellular transport of drugs that have affinity for albumin and IgG into the CNS | [16]       |
|                         | Overexpression of efflux pumps                                                                                                                          | Efflux pump inhibitors may improve drug deliver into the brain.                                                | [17]       |
| Parkinson's Disease     | BBB disruption                                                                                                                                          | It enhanced therapeutic agent concentration in the brain.                                                      | [18]       |
| HIV                     | Increase in diameter of cortical vessels, thinning of basal lamina, loss of glycoproteins, apoptosis of endothelial cells and tight junction disruption | Potentially it may increase drug transport into the brain due to the leaky barrier.                            | [19]       |
| Infectious disease      | Leukocyte Invasion, elevated CSF - to - Serum albumin ratio and BBB impairment                                                                          | It may enhance paracellular transport of drugs and drugs with affinity for albumin.                            | [20,21]    |
| Inflammation            | Increased BBB permeability                                                                                                                              | It may facilitate paracellular drug transportation.                                                            | [22]       |
| Stroke                  | BBB disruption                                                                                                                                          | It enhanced paracellular drug, e.g. Ginkgolide B, passage into the brain                                       | [23]       |
|                         | Upregulation of diphtheria toxin receptor                                                                                                               | It may provide disease-induced specific drug targeting of the BBB and receptor mediated transcytosis.          | [24,25]    |
| Trauma                  | BBB breakdown                                                                                                                                           | It enhanced therapeutic agent concentration in brain                                                           | [26]       |
| Pain                    | Alteration of BBB chemokine receptor due to activated astrocytes                                                                                        | It may lead to astrocyte-targeted therapy.                                                                     | [27]       |
|                         | Decreased Tight junctions proteins and BBB perturbation                                                                                                 | It may facilitate paracellular drug transportation.                                                            | [28]       |
| Brain Tumour            | Loss of Tight Junctions in tumous vascular system, enhanced retention effect                                                                            | Angiogenic vessels are permeable to nano-sized materials                                                       | [29]       |
|                         | Overexpression receptors of folate, insulin and transferrin                                                                                             | It enhanced folic acid, insulin and transferrin-attached nanoparticles across the BBB.                         | [30,31]    |
| Ischemia /Seizures      | Upregulation of Diphtheria Toxin Receptors                                                                                                              | Potentially it may increase disease-induced specific drug targeting of BBB and receptor mediated transcytosis. | [32]       |

To cross the BBB many methods has been developed which includes disruption of BBB, Carrier mediated transport and chemical modification of drug molecule. Intranasal route is being widely studied because of its potential to cross BBB through olfactory receptors. Nanocarriers are important drug delivery systems which can be utilized to cross the BBB and to deliver drugs to brain.

### Approaches of Brain Targeting

#### Invasive approaches:

- Intracerebroventricular infusion
- Convection – enhanced delivery
- Intra cerebral injection or implants
- Disruption of BBB

#### Noninvasive:

- **Chemical techniques**
  - Prodrugs
- **Colloidal techniques**
  - Nanoparticles
  - Liposomes
- **Miscellaneous techniques**

- Intranasal delivery

**Invasive Approach**

In this surgery is done by administering the drug in brain through implants or Intra cerebro ventricular infusion. These methods are complex and require neurosurgeons. Larger and smaller both molecules can be given through these methods.

**ICV Injection**

Drug distribution is limited to brain due to protective behavior of BBB, Intra cerebro ventricular drug administration is method that can cross BBB and high brain drug concentration can be achieved. In this method drug is directly administered into ventricles of brain. Factors which can affect drug efficacy are osmolarity, pH, preservatives used and diluents which are used in drug solution. Drugs can be administered intraventricularly using an Ommaya Reservoir; a plastic reservoir implanted subcutaneously is scalp and connected to the ventricles within brain via an outlet catheter. Drug solution can be injected subcutaneously to implanted reservoir and delivered to ventricles by manual compression of reservoir through scalp [33]. This method is complex and required surgeons.

**Limitations:**

Due to presence of extracellular fluid space of brain drug diffusion is through brain parenchyma is very slow and inversely proportional to Molecular weight of drug [33].

**Convection Enhanced Delivery**

In this catheters of small diameter are directly placed in brain tumor. This reduces the surgical exposure of brain. Therapeutics is infused into tumor to saturate the target tissue. This allows macromolecular drugs to bypass the blood brain barrier. It minimizes the exposure of drug molecules to rest of body and drug can be delivered to targeted site. Convection enhanced delivery gives better drug distribution to brain tumors.

**Limitations:**

Back flow of drug with catheters and drug leakage in non desired areas.

**Intracerebral Implants**

In this technique therapeutic agents comprises of biodegradable polymer matrix or reservoir is implanted intracerebrally. This provides sustained drug release and localized delivery of drugs. Basic mechanism involves in this is diffusion and convection.

**Limitations:**

Drug distribution in brain decrease exponentially by distance to site of action required.

**Blood Brain Barrier Disruption**

By this medication pass through the protective blood brain barrier of brain and large doses can be send to tumor and nearby tissue. Blood Brain Barrier is made of tightly knit cells that line the blood vessels in brain. By shrinking these cells medication can be passed through brain and reach at targeted site. Some of the important techniques are:

**Osmotic Disruptions**

Endothelial cells shrinks by osmotic shock and disrupt tight junctions present in brain. Intracarotid infusion of a hypertonic arabinose or mannitol solution is used to cause vasodilatation and shrinkage of cerebrovascular endothelial cells. This method has been used to target water soluble drugs, peptides, antibodies and viral vectors for gene therapy. Metastatic or primary brain tumors are also treated with this method.

**MRI Guided Targeted Blood Brain Barrier Disruption**

This method is used to disrupt blood brain barrier noninvasively and reversibly at targeted locations. This technique is highly localized and can target wide range of therapeutic agents to brain. In 1942 Lynn et al. finds the potential of Focused ultrasound to produce thermal or mechanical effects in brain

**Nanocarriers for Brain Drug Delivery**

Nanocarriers are colloidal system of nano scale size able to transport drug molecules to different sites of body. They can carry small molecular weight drugs or macromolecules such as genes or proteins. Nanocarriers protect drug

from degradation, reduces the toxicity of molecules, increase solubility and bioavailability of drug molecules and can increase the half life in blood stream. Drug carrier can be made up of carbon, polymeric and magnetic materials. They can be utilized in targeted drug delivery and controlled drug delivery. An appropriate carrier is used in this formulation which is non-toxic, biodegradable, biocompatible and binds with drug without changing its chemical features. Nanocarriers such as liposomes and micelles can enhance the pharmacological properties of drugs because their small size (~100 nm or less) allows them to easily cross the biological membranes. In Table 4 list of nanocarriers which are nanoparticles and in Table 5 list of nanocarriers other than nanoparticles are listed respectively.

Table 4: List of nanocarriers used for CNS drug delivery

| Carrier Type                   | Materials Used                                      | Drugs                         | Results                           | Ref.    |
|--------------------------------|-----------------------------------------------------|-------------------------------|-----------------------------------|---------|
| Polymeric nanoparticle         | PBCA                                                | Methotrexate                  | Increase Brain Drug Concentration | [34-36] |
|                                |                                                     | Dalargin                      |                                   |         |
|                                |                                                     | Temozolomide                  |                                   |         |
|                                | MMA-SPM, PCBA                                       | Lamivudine(3TC)               | Enhanced BBB permeability         | [37]    |
|                                |                                                     | Zidovudine (AZT)              |                                   |         |
|                                | PLGA/Alginate                                       | Dexamethasone                 | Extended drug release             | [38]    |
|                                | PLA/PEG                                             | Vasoactive intestinal peptide | Increase Brain Drug concentration | [39]    |
| Solid lipid nanoparticle (SLN) | PLGA                                                | Superoxide dismutase          | Improved neurological functions   | [40]    |
|                                | Soya phosphatidyl-choline 95%                       | Doxorubicin                   | Improved brain accumulation       | [41,42] |
|                                | Stearic acid/ Soy bean lecithin                     | Camptothecin                  | Increases Drug Release            | [43]    |
|                                | Stearylamine                                        | Paclitaxel                    | Enhanced Brain Drug Concentration | [44]    |
| Lipid nanocapsule              | Triglycerides of Capric and caprylic acids, Solutol | Melatonin                     | Enhanced bioavailability          | [44]    |
|                                |                                                     | Etoposide                     | Improved Bioavailability          | [45,46] |
|                                |                                                     | Paclitaxel                    | Increased Half life               | [44]    |
| Albumin nanoparticle           | Albumin                                             | Loperamide                    | Improved BBB penetration          | [47]    |
|                                |                                                     | Paclitaxel                    | Increase accumulation of Drug     | [48]    |

Table 5: List of nanocarriers other than nanoparticles

| Carrier type    | Material                                   | Drug                                                      | Result                             | Ref     |
|-----------------|--------------------------------------------|-----------------------------------------------------------|------------------------------------|---------|
| Liposomes       | Phospholipids and cholesterol              | Phenytoin, $\gamma$ -Aminobutyric acid                    | Improved local action              | [49]    |
|                 |                                            | Cisplatin                                                 | drug concentration                 | [50]    |
|                 |                                            | Stavudine                                                 | Enhanced Anti HIV Increased effect | [50]    |
|                 |                                            | Amphotericin                                              | Increased Brain Drug concentration | [51]    |
| Micelles        | Pluronic P85                               | Biphalin, Enkephalin, Morphine                            | Enhanced analgesic properties      | [51,52] |
|                 | Core Shell Micelles                        | Doxorubicin, Digoxin, Paclitaxel, Ritonavir, Vinblastine, | Increased brain drug concentration | [52-54] |
|                 |                                            | Antioxidant nitroxyl radicals                             | Improved Neurological functions    | [55]    |
| Dendrimers      | Mannosylated Poly(propyl eneimine)         | Lamivudine                                                | Increased uptake of Lamivudine     | [56]    |
|                 | Polyether-copolyester                      | Methotrexate                                              | Enhanced circulation               | [57]    |
| Nanogel         | N-isopropyl-acrylamide/N-vinyl-pyrrolidone | 5-fluorouracil                                            | Increased Brain accumulation       | [58]    |
| Nano-Emulsion   | Edible Oil                                 | Saquinavir                                                | Increased Brain drug Concentration | [59]    |
|                 | Pine nut oil                               | Paclitaxel, Ceramide                                      | Increased Drug Uptake              | [60]    |
| Nano suspension | Atovaquone Crystal                         | Atovaquone                                                | Enhanced Drug bioavailability      | [61]    |

### Liposomes

These are nano or microsize vesicles which consist of one or more lipid bilayers surrounding an aqueous compartment. These are spherical shape vesicles composed of cholesterol and phospholipids they have ability to incorporate hydrophilic, lipophilic and hydrophobic compounds. Hydrophilic compounds can be entrapped into aqueous core of liposomes or between the lipid layer and water phase. Lipophilic or hydrophobic compounds are

incorporated into hydrophobic core of lipid bilayers. With the incorporation of polymers, polysaccharides, peptides, antibodies their surface can be modified to improve brain drug delivery. Table 6 comprises of advantages and disadvantages of liposomes system. In Tables 7 and 8 applications of liposomes and marketed preparations of liposomes are listed respectively.

**Table 6: Advantages and disadvantages of liposomes [62]**

| Advantages                                                                                                                                   | Disadvantages                                                           |
|----------------------------------------------------------------------------------------------------------------------------------------------|-------------------------------------------------------------------------|
| Liposomes are increased efficacy and therapeutic index of drug (Actinomycin-D).                                                              | Production cost is high.                                                |
| Liposome is increased stability via encapsulation.                                                                                           | Leakage and fusion of encapsulated drug / molecules.                    |
| Liposomes are biocompatible, completely biodegradable, non-toxic, flexible and nonimmunogenic for systemic and non-systemic administrations. | Sometimes phospholipid undergoes oxidation and hydrolysis like reaction |
| Liposomes are reduction in toxicity of the encapsulated agent (Amphotericin B, Taxol).                                                       | Short half-life.                                                        |
| Liposomes help to reduce exposure of sensitive tissues to toxic drugs                                                                        | Low solubility                                                          |
| Site avoidance effect.                                                                                                                       | Fewer stabiles                                                          |
| Flexibility to couple with site-specific ligands to achieve active targeting                                                                 |                                                                         |

### Liposomes Mechanisms of Crossing BBB

#### Cationization of vector:

At blood brain barrier electrostatic interaction is present between positive charge and polyanious which can lead to adsorptive mediated endocytosis [63,64].

#### Tageting ligand:

Receptor mediated transcytosis can occur by using ligand targeting liposomes toward the receptors present on brain endothelial cells. Antibodies and aptamers can bind to liposome surface [65,66].

#### Triggered drug release:

Magnetic field, temperature, ultrasound intensity, light or electric pulses are specific external stimuli which can trigger drug release of liposomes [67,68].

#### Theranostic:

Non invasive contrast agents can also entrapped in liposomes. For the diagnosis, real times monitoring of disease multifunctional theranostic liposomes are used [68,69].

**Table 7: Applications of liposomes [70]**

| Actions                                                                                                    | Examples                                                                                                                                                     |
|------------------------------------------------------------------------------------------------------------|--------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Improved solubility of lipophilic and amphiphilic drugs                                                    | Amphotericin B, porphyrins, minoxidil, some peptides, and anthracyclines, respectively; hydrophilic drugs, such as anticancer agent doxorubicin or acyclovir |
| Passive targeting to the cells of the immune system, especially cells of the mononuclear phagocytic system | Antimoniales, amphotericin B, porphyrins, vaccines, immunomodulators                                                                                         |
| Sustained release system of systemically or locally administered liposomes                                 | Doxorubicin, cytosine arabinoside, cortisones, biological proteins or peptides such as vasopressin                                                           |
| Site-avoidance mechanism                                                                                   | Doxorubicin and amphotericin B                                                                                                                               |
| Site-specific targeting                                                                                    | Anti-inflammatory drugs, anti-cancer, anti-infection                                                                                                         |
| Improved transfer of hydrophilic, charged molecules                                                        | Antibiotics, chelators, plasmids, and genes                                                                                                                  |
| Improved penetration into tissues                                                                          | Corticosteroids, anesthetics, and insulin                                                                                                                    |

**Table 8: Marketed liposomes products [71]**

| Product                  | Drug                 | Formulation     | Company                          | Indication/Target         | Country         |
|--------------------------|----------------------|-----------------|----------------------------------|---------------------------|-----------------|
| Doxil <sup>TM</sup>      | Doxorubicin          | Liposomes (LCL) | Sequus Pharmaceuticals, Inc., CA | Kaposi sarcoma in AIDS    | USA and Europe  |
| Ambisome <sup>TM</sup>   | Amphotericin B       | Liposomes (CL)  | NeXstar Pharmaceutical Inc., CO  | Serious Fungal Infections | In 24 countries |
| DaunoX-ome <sup>TM</sup> | Daunorubicin citrate | Liposomes (LCL) | NeXstar Pharmaceutical Inc., CO  | Kaposi sarcoma in AIDS    | USA and Europe  |
| Amphocil <sup>TM</sup>   | Amphotericin B       | Lipid Complex   | Sequus Pharmaceutical Inc., CA   | Serious Fungal Infections | Asia and Europe |
| Abelcet <sup>TM</sup>    | Amphotericin B       | Lipid Complex   | The Liposome Company, NJ         | Serious fungal Infections | USA and Europe  |

### Solid Lipid Nanoparticles

In 1991 solid lipid nanoparticles were introduced which gives alternative carrier system to traditional colloidal carriers, such as emulsions, liposomes and polymeric micro and nanoparticles. They have many advantages over the traditional drug delivery system but lesser disadvantages as compared to them [72]. Poly(ethylene glycol)- modified SLNs can penetrate BBB and allow great delivery to brain [73]. For the manufacturing of SLN Lipids are used, which are solid at room temperature and also at room temperature, such as triglycerides, Partial glycerides, fatty acids, steroids and waxes.

#### Advantages of SLNs [74]

- Possibility of controlled drug release and drug targeting
- Increased drug stability
- High drug payload
- Incorporation of lipophilic and hydrophilic Drugs
- No biotoxicity of carrier
- Avoidance of organic solvents
- No problems with respect to large scale production and sterilization

#### Disadvantages of SLN

- Poor Drug loading
- Drug Expulsion After polymeric transition during storage
- Relatively high water content of dispersions (70-99.9%)

#### Advantages of SLNs Over Polymeric Nanoparticles

Reticulo Endothelial System cannot take up SLNs ranging 120-200 nm and bypass liver and spleen filtration [75]. For weeks controlled can be achieved and site specific delivery can be possible by attaching ligand and by coating [76,77]. SLNs are very stable as compared to other formulations and have high drug payload [43]. They can entrap both hydrophilic and hydrophobic drugs [73,78]. Mostly carrier lipids are biodegradable and safe [79] and less usage of organic solvent [80].

### Microspheres

Microspheres are also referred as microparticles. These are small spherical particles ranging from 1  $\mu\text{m}$  to 1000  $\mu\text{m}$ . These are free flowing powder consisting of proteins, starches, gums, fats and waxes and synthetic biodegradable polymers. Albumin and Gelatin are natural polymers and Poly lactic acid and polyglycolic acid is synthetic polymer which is used in microspheres. Polymer microspheres comprised of polyethylene and polystyrene are most commonly used. Polystyrene microspheres are used in biomedical applications and polyethylene microspheres as temporary and permanent filler. Glass microspheres have limited applications in medical field. Ceramic microspheres are used as grinding media. In Table 9 Applications of microspheres for CNS delivery are listed

#### Materials Used in Preparation of Microspheres [81,82]

##### Classification:

- Synthetic polymers
- Natural Polymers

##### Synthetic polymers:

**Non biodegradable polymers:** Polymethyl methacrylate (PMMA), Acrolein, Glycidyl methacrylate, Epoxy Polymers.

**Biodegradable Polymers:** Lactides, their glycolides and their copolymers, Polyalkyl Cyano Acrylate, Polyamides.

##### Natural Polymers:

Obtained from Sources like proteins, carbohydrates and chemically modified carbohydrates.

**Proteins:** Albumin, Gelatin, Collagen.

**Carbohydrates:** Agarose, Carrageenan, Chitosan, Starch, Chemically modified Carbohydrates like Poly (Acryl) Dextran, Poly (acryl) Starch.

### Advantages

They provided sustained release of drug and minimize dosing frequency and improve patient compliance. Due to Smaller size and spherical shape can to given intravenously. They enhance the bioavailability and minimize adverse effects of drug.

### Limitations

Incorporation of food and rate transit though gut can alter the release rate of controlled release dosage form. Release rate may be differing from one dose to another in prolonged release. Sustained and controlled release formulations usually contains higher dose of drug which can lead to toxicity.

### Applications

#### Bioadhesive microspheres:

- Biodegradable Starch microspheres for Nasal delivery of Domperidone [83].
- Nasal delivery of insulin by degradable microspheres [84].
- Chitosan – ethylcellulose Microspheres for nasal delivery [85].
- Mucoadhesive microspheres for nasal administration of metoclopramide[86].
- Mucoadhesive microspheres for gastrointestinal tract [86].
- Microspheres for sustained ocular delivery of daunorubicin [87].
- Bioadhesive sulfacetamide sodium microspheres for treatment of bacterial keratitis [88].

#### Magnetic microspheres:

- Thermo- sensitive magnetic hyrogel microspheres for enzyme immobilization [89].
- Low dose of doxorubicin loaded magnetic albumin microspheres for selective targeting [90].
- Magnetic microspheres for labeling and separation of cells [91].

#### Floating microspheres:

- Gastroretentive floating drug delivery of repagalinide by calcium silicate based microspheres [92].
- Prolongation of gastric residence time of Cimetidine by floating microspheres [93].
- Floating microspheres of metformin hydrochloride for controlled release [94].
- Floating bioadhesive microspheres of acetohydroxamic acid for clearance of helicobacter pylori [95].

#### Radioactive microspheres:

- Radioactive microspheres to assess distribution of cardiac output in rabbits [96].
- Targeted delivery of magnetic radioactive 90 Y-microspheres to tumor cells [97].

**Table 9: Application of microspheres for neurodegenerative diseases**

| Polymer                       | Microsphere size                   | Drug                                       | References |
|-------------------------------|------------------------------------|--------------------------------------------|------------|
| Hyaluronane derivative        | 2-21 $\mu\text{m}$                 | Nerve growth factor                        | [98]       |
| Alginate-polylysine           | 50 $\mu\text{m}$                   | Nerver growth factor                       | [99]       |
| Alginate-Chitosan             | 50 $\mu\text{m}$ - 3 $\mu\text{m}$ | BDNF                                       | [100]      |
| Poly(DL-lactide-co-glycolide) | 5-45 $\mu\text{m}$                 | Dopamine                                   | [101]      |
| Poly(DL-lactide-co-glycolide) | 17.2 $\mu\text{m}$                 | Nerve growth factor                        | [102]      |
| Poly(DL-lactide-co-glycolide) | 2.5 $\mu\text{m}$                  | Monosialoganglioside + Nerve Growth Factor | [103]      |
| Poly(DL-lactide-co-glycolide) | 2.6 $\mu\text{m}$                  | Brain derived Neurotrophic Factor          | [100]      |
| Poly(DL-lactide-co-glycolide) | 8-11 $\mu\text{m}$                 | Nerve Growth Factor                        | [104]      |
| Poly(DL-lactide-co-glycolide) | 50 $\mu\text{m}$                   | Nerve Growth Factor                        | [104]      |
| Poly(DL-lactide-co-glycolide) | 25 $\mu\text{m}$                   | Nerve Growth Factor                        | [103]      |
| Poly(DL-lactide-co-glycolide) | 1.8 $\mu\text{m}$                  | Ciliary Neurotrophic Factor                | [103]      |
| Poly(DL-lactide-co-glycolide) | 2 $\mu\text{m}$                    | Ciliary Neurotrophic Factor                | [105]      |

### Dendrimers

These are class of polymeric materials that are nano sized symmetrical structure consisting of tree like atoms. The word “dendrimer” originated from two words, Greek word “Dendron” meaning tree, and “meros” which mean part. Dendrimers are monodisperse macromolecules that contain symmetric branching units built around a small molecule or linear polymer core. Dendrimers structure consists of three components:

- A initiator



- interior layers or repeating units
- Exterior layer with outer interior generations

Dendrimers of different compositions are explored for drug delivery which includes Poly(aminoamine (PANAM), Poly-(Etherhydroxylamine) (PEHAM) and Poly (propyleneimine) (PPI) dendrimers [106]. PANAM have unique structures and properties. Polycationic dendrimer having primary amine on surface are Full generation PANAM and Polyanionic dendrimer that has carboxylic acids on surface are half generation [107]. Polyanionic dendrimers are less toxic than polycationic dendrimers [107]. PANAM dendrimers for CNS delivery can be synthesized [108-110]. Dendrimers are modified with spacers for bioavailability improvement, buffer capacity and half-life. Surface modified dendrimers are conjugated with specific ligands to target BBB. Surface modified dendrimers complexed with gene therapeutics or drug. Imaging agents covalently conjugated with dendrimer used for *in vivo* imaging and diagnosis. Tables 10 and 11 consist of Dendrimers generations and therapeutics applications of dendrimers. In Table 12 list of PANAM dendrimers for CNS delivery is listed.

Table 10: Dendrimers generations

| Dendrimers Generation | PANAM (No of Branches) | PPI (No of Branches) | Phosphorus Dendrimers |
|-----------------------|------------------------|----------------------|-----------------------|
| 0                     | 4                      |                      |                       |
| 1                     | 8                      |                      |                       |
| 2                     | 16                     | 8(16)                | 24                    |
| 3                     | 32                     | 16(32)               | 48                    |
| 4                     | 64                     | 32(64)               | 96                    |
| 5                     | 128                    | 64(128)              | 192                   |

Table 11: Therapeutic applications of dendrimers

| Application                       | Dendrimers                        | Drug                      | Reference |
|-----------------------------------|-----------------------------------|---------------------------|-----------|
| Solubilization                    | PPI Dendrimers                    | Amphotericin B Famotidine | [111]     |
| Enhanced cellular uptake          | Mannosylated PPI dendrimers       | Efavirenz                 | [112]     |
| Biocompatible Drug carrier        | Mannosylated PPI dendrimers       | Rifampicin                | [113]     |
| Drug Targeting                    | Mannosylated PPI dendrimer        |                           | [113]     |
|                                   | Folate conjugated dendrimers      |                           | [114]     |
|                                   | Dextran conjugated PPI dendrimers |                           | [115]     |
| Sustained release                 | Dextran conjugated PPI dendrimers | Doxorubicin HCl           | [115]     |
| Delivery of anti-HIV drug         | 4.0 G PPI                         | Zidovudine                | [116]     |
| Delivery of anticancer bioactives | 4.0 G PAMAM                       | Doxorubicin               | [117]     |

Table 12: List of PANAM dendrimers for CNS delivery

| Generation | Linkage          | Drugs delivered                            | Ligand(s)                                        | Major findings and comments                                                       | Ref   |
|------------|------------------|--------------------------------------------|--------------------------------------------------|-----------------------------------------------------------------------------------|-------|
| G4         | MAL-PEG5000-NHS  | Doxorubicin                                | Transferrin (Tf) and wheat germ agglutinin (WGA) | Reduced nonspecific uptake by the normal cells; enhanced transport across the BBB | [118] |
|            | MAL-PEG5000-NHS  | Doxorubicin                                | Transferrin (Tf) and tamosifen (TAM)             | Enhanced transport across the BBB                                                 | [119] |
|            | NHS-PEG3400-MAL  | pEGFP-N2 plasmid                           | Lactoferrin (Lf)                                 | Enhanced brain uptake and transfection efficiency                                 | [120] |
|            | Triglycine (GGG) | Quantum dots (Qdots) and YFP siRNA plasmid | epidermal growth factor (EGF)                    | Enhanced nucleic acid delivery compared to trans-IT                               | [121] |
| G5         | NHS-PEG3400-MAL  | pEGFP-N2 plasmid                           | rabies virus glycoprotein (RVG29)                | Enhanced <i>in vivo</i> biodistribution in the brain                              | [109] |
|            | NHS-PEG3400-MAL  | pEGFP-N2 plasmid                           | Transferrin (Tf)                                 | Enhanced brain uptake and brain transfection efficiency                           | [110] |
|            | NHS-PEG3400-MAL  | pEGFP-N2 plasmid                           | Angiopep-2                                       | Enhanced brain uptake and transfection efficiency                                 | [108] |
|            | NHS-PEG3400-MAL  | pORF-TRAIL plasmid                         | Angiopep-2                                       | Enhanced <i>in vivo</i> biodistribution in the brain                              | [122] |
|            | NHS-PEG3400-MAL  | pORF-TRAIL plasmid                         | Chlorotoxin (CTX)                                | Enhanced <i>in vivo</i> biodistribution in the brain                              | [123] |
| G4.5       | NHS-PEG3400-MAL  | opioid peptide DPDPE                       | Transferrin receptor monoclonal antibody OX26    | Enhanced permeability through buccal mucosa by multiple fold                      | [124] |

### Micelles

Micelles are amphilic molecules aggregate present in aqueous medium [125] they can entrap poor water soluble, lipophilic compound in their micelles core. Mostly studies micelles are Pluronic based micelles. They can deliver the drug to Brain by crossing the blood brain barrier [54,125]. Micelles composed of Pluronic P85 Shows Enhancement of upto 19 fold in bovine brain microvessel endothelial cell line [52,54]. Polymeric micelles enhanced the delivery of Ritonavir, vinblastine and paclitaxel by suppressing the MDR1 mediated Drug efflux [54,52]

Micelles does not increase brain drug level in MDR 1a/b mice because of ATP depletion [126] Table 13 consists of studies done using Micelles for solubility enhancement.

**Table 13: Solubility enhancement using micelles**

| Drug         | Amphiphilic polymer                                                                                                   | Comment                                                                                                                                    | Ref   |
|--------------|-----------------------------------------------------------------------------------------------------------------------|--------------------------------------------------------------------------------------------------------------------------------------------|-------|
| Camptothecin | Pluronic P105, d- $\alpha$ -tocopheryl polyethylene glycol 1000 succinate                                             | Increased micellar stability; increased cytotoxicity                                                                                       | [127] |
| Docetaxel    | Poly(ethylene oxide)-blockpoly(styrene oxide) (PEO-b- PSO) and PEO-b-poly(butylene oxide) (PEO-b-PBO)                 | PSO-based copolymers were associated with higher solubilizing capacities than PBO due to the aromatic structure of the coreforming polymer | [127] |
| Griseofulvin | E B copolymers (E = oxyethylene, m n B = oxybutylene, subscripts denote number-average block lengths in repeat units) | Solubilization independent of B block length when it exceeds about 15B units                                                               | [128] |
| Paclitaxel   | N-octyl-O-sulfate chitosan                                                                                            | Improved bioavailability and reduced toxicity                                                                                              | [129] |
| Paclitaxel   | mixed micelles of polyethylene glycol-phosphatidyl ethanolamine (PEG-PE) and vitamin E                                | Mixed micelles efficiently solubilized poorly soluble drug as compared to PEGPE micelles                                                   | [130] |

### Nose to Brain Drug Delivery

Through the olfactory or trigeminal nerve system that ends at olfactory neuroepithelium or respiratory epithelium drug delivery to brain can be possible through nasal route [131]. BBB can be crossed by using these nerve systems. Nose to brain drug delivery can reduce systemic toxicity but this route is inefficient. According to Illum *et al.* less than 0.1% of drug through nasal route will normally reaches to brain [132]. Poor nose to brain drug delivery can be improved using nanocarriers. Main barrier of this route is olfactory epithelium. Drug loaded nanocarriers can be used to achieve transmembrane transport across the barrier. Many studies have been done which suggests this strategy. A few studies have provided supportive data to this strategy. According to Betbeder *et al.* analgesic effect of morphine can be increased by administering morphine nasally using 60 mm maltodextrin nanoparticles [133]. This study also concludes that Intranasal nanoparticles formulation shows superior analgesic effect against subcutaneously administered morphine. Zhang *et al.* done same study using intranasally administered nanoparticles of nimodipine which shows improved results using nasal route [134]. Mucoadhesive nanoparticles can improve the olfactory drug delivery. Mainly Chitosan is used as mucoadhesive agent that can interact with junctional complexes between epithelial cells. Estradiol and risperidone were given intranasally using Chitosan nanocarriers and their therapeutic effects were stronger than intravenously administered nanocarriers [135,136]. Lectins can also be used as mucoadhesive [137]. Lectin coated PLA nanoparticles can increase the coumarin concentration in brain by two folds instead of uncoated ones.

### Neurotoxicity of Nanocarriers

CNS is highly protective system but nanocarriers can cross them which can also lead to over exposure of drug molecules and nanomaterials resulting in CNS toxicity. *In vitro* and *in vivo* neurotoxicity is reported in many studies using nanocarriers. In table shows these studies [48,138-141]. For *in vitro* neurotoxicity evaluation PC12 neuronal cell line is commonly used [138,142]. Brain damage is generally caused by increasing Reactive oxygen species levels and they can further exploited using neural stem cells in *in vitro* studies and *in vivo* studies [48,140]. Titanium oxide nanoparticles does not directly cause damage to dopaminergic neuronal cells, but can cause damage to microglial cells [48]. Table 14 consists of several studies showing neurotoxicity caused by nanocarriers. Mostly neurotoxicity studies performed using nanoparticles which are made of inorganic material. In future, toxicity of polymeric and lipid nanocarriers loaded drug should be conducted to find out potential risks along with targeted drug delivery.

**Table 14: Neurotoxicity of nanocarriers for CNS delivery**

| Tested model                | Carrier Type                            | Major Findings                                               | References |
|-----------------------------|-----------------------------------------|--------------------------------------------------------------|------------|
| PC12 Neuronal cell Line     | Manganese oxide nanoparticles           | Increased levels of Reactive oxygen species                  | [143]      |
|                             | Copper, silver, manganese nanoparticles | Toxicity of Dopaminergic                                     | [144]      |
|                             | Anionic magnetic nanoparticles          | Decrease in cell viability and altered nerve growth factor   | [145]      |
| Mouse Neural stem cells     | Zinc oxide nanoparticles                | Cell apoptosis                                               | [146]      |
| Dopaminergic neuronal cells | Titanium oxide nanoparticles            | Neuronal cell toxicity                                       | [138]      |
| Mice model                  | Ultrafine carbon black particles        | Changes inflammatory cytokines                               | [139]      |
| Rat model                   | Manganese oxide nanoparticles           | Increase in macrophage inflammatory protein-2                | [142]      |
| Fish model                  | Fullerenes nanoparticles                | Lipid peroxidation, protein damage and glutathione depletion | [142]      |

## CONCLUSION

Nanotechnology is suitable for targeted drug delivery and improves the therapeutic management of Brain diseases. Pharmaceutical industry is moving toward the nanotechnology because it can cross the BBB and improve patient health. But many issues have been reported using nanotechnology for Brain diseases. Because of complex structure of brain and toxicity produced by nanomaterials, toxicity studies are required. Nanocarrier medicine should be evaluated for both with and without drug molecules. Polymers or other materials which have been used should be in therapeutic range and should not cause any toxicity and interfere with drug molecule action. Mostly Brain diseases require emergency treatment so chronic and cumulative effect of nanoformulation should be checked on brain tissues and toxicity studies should also be done. Brain is a very vital organ; damages to this organ are very difficult to evaluate as compared to other organs such as liver, heart or kidney. Many advanced diagnostic techniques are employed to diagnose brain diseases such as Magnetic resonance imaging, positron emission tomography and computed tomography scan. Targeting drug delivery is an important science to cure chronic diseases. Many targets like LDL receptors, Insulin receptors and transferrin are important in targeted drug delivery. CNS targeting can be caused by carrier-mediated transporters and endogenous carriers also be exploited for brain targeting.

## REFERENCES

- [1] MJ Cipolla. The cerebral circulation. Integrated systems physiology: *From molecule to function*. **2009**, 1-59.
- [2] L Biddlestone-Thorpe; N Marchi; K Guo; C Ghosh; D Janigro; K Valerie; H Yang. *Adv Drug Deliv Rev*. **2012**, 64, 605-613.
- [3] W Löscher; H Potschka. *Nature Rev Neurosci*. **2005**, 6, 591-602.
- [4] U Kniesel; H Wolburg. *Cell Mol Neurobiol*. **2000**, 20, 57-76.
- [5] NJ Abbott. *J Anat*. **2002**, 200, 523-534.
- [6] W Kamphorst, AG De Boer, PJ Gaillard. Brain Drug Targeting: The Future of Brain Drug Development, Cambridge University Press, **2002**.
- [7] E Neuwelt; NJ Abbott; L Abrey; WA Banks; B Blakley; T Davis; B Engelhardt; P Grammas; M Nedergaard; J Nutt. *Lancet Neurol*. **2008**, 7, 84-96.
- [8] MA Deli. Solubility, Delivery, and ADME Problems of Drugs and Drug-Candidates. Washington: Bentham Science Publ. Ltd., **2011**, 144-165.
- [9] AH Schinkel. *Adv Drug Deliv Rev*. **1999**, 36, 179-194.
- [10] TR Stouch; O Gudmundsson. *Adv Drug Deliv Rev*. **2002**, 54, 315-328.
- [11] MA Deli. *Biochim Biophys Acta*. **2009**, 1788, 892-910.
- [12] Y Chen; L Liu. *Adv Drug Deliv Rev*. **2012**, 64, 640-665.
- [13] A Minagar; JS Alexander. *Mult Scler J*. **2003**, 9, 540-549.
- [14] AW Vorbrodt; DH Dobrogowska. *Folia Histochem Cytobiol*. **2004**, 42, 67-76.
- [15] DW Holman; RS Klein; RM Ransohoff. *Biochim Biophys Acta*. **2011**, 1812, 220-230.
- [16] G Bowman; J Kaye; M Moore; D Waichunas; N Carlson; J Quinn. *Neurology*. **2007**, 68, 1809-1814.
- [17] HC Wijesuriya; JY Bullock; RL Faull; SB Hladky; MA Barrand. *Behav Brain Res*. **2010**, 1358, 228-238.
- [18] AM Palmer. *Neurobiol*. **2010**, 37, 3-12.
- [19] M Toborek; YW Lee; G Flora; H Pu; IE András; E Wylegala; B Hennig; A Nath. *Cell Mol Neurobiol*. **2005**, 25, 181-199.
- [20] M Gottfredsson, JR Perfect. In: Book Fungal meningitis, Thieme Medical Publishers, New York, **2000**, 307-322.
- [21] JD Lee; LY Tsai; CH Chen; JJ Wang; JK Hsiao; CM Yen. *Acta Trop*. **2006**, 97, 204-211.
- [22] H Stolp; P Johansson; M Habgood; K Dziegielewska; N Saunders; C Ek. *Cardiovasc Psychiatry Neurol*. **2011**.
- [23] W Fang; Y Deng; Y Li; E Shang; F Fang; P Lv; L Bai; Y Qi; F Yan; L Mao. *Eur J Pharm Sci*. **2010**, 39, 8-14.
- [24] K Jin; Y Sun; L Xie; J Childs; XO Mao; DA Greenberg. *J Cereb Blood Flow Metab*. **2004**, 24, 399-408.
- [25] A De Boer; P Gaillard. *Annu Rev Pharmacol Toxicol*. **2007**, 47, 323-355.
- [26] M Habgood; N Bye; K Dziegielewska; C Ek; M Lane; A Potter; C Morganti-Kossmann; N Saunders. *Eur J Neurosci*. **2007**, 25, 231-238.
- [27] Y Persidsky; SH Ramirez; J Haorah; GD Kanmogne. *J Neuroimmune Pharmacol*. **2006**, 1, 223-236.

- [28] TA Brooks; SM Ocheltree; MJ Seelbach; RA Charles; N Nametz; RD Egleton; TP Davis. *Brain Res.* **2006**, 1120, 172-182.
- [29] H Maeda; G Bharate; J Daruwalla. *Eur J Pharm Biopharm.* **2009**, 71, 409-419.
- [30] V Soni; D Kohli; S Jain. *J Drug Target.* **2008**, 16, 73-78.
- [31] A Doi; S Kawabata; K Iida; K Yokoyama; Y Kajimoto; T Kuroiwa; T Shirakawa; M Kirihata; S Kasaoka; K. Maruyama. *J. Neurooncol.* **2008**, 87, 287-294.
- [32] N Tanaka; M Sasahara; M Ohno; S Higashiyama; Y Hayase; M Shimada. *Brain Res.* **1999**, 827, 130-138.
- [33] A Misra; S Ganesh; A Shahiwala; SP Shah. *J Pharm Pharm Sci.* **2003**, 6(2), 252-273.
- [34] K Gao; X Jiang. *Int J Pharm.* **2006**, 310, 213-219.
- [35] D Das; S Lin. *J Pharm Sci.* **2005**, 94, 1343-1353.
- [36] XH Tian; XN Lin; F Wei; W Feng; ZC Huang; P Wang; L Ren; Y Diao. *Int J Nanomed.* **2011**, 6, 445-452.
- [37] YC Kuo; HH Chen. *Int J Pharm.* **2006**, 327, 160-169.
- [38] DH Kim; DC Martin. *Biomaterials.* **2006**, 27(15), 3031-3037.
- [39] X Gao; B Wu; Q Zhang; J Chen; J Zhu; W Zhang; Z Rong; H Che; X Jiang. *J Control Release.* **2007**, 121, 156-167.
- [40] MK Reddy; V Labhasetwar. *FASEB J.* **2009**, 23, 1384-1395.
- [41] GP Zara; R Cavalli; A Fundarò; A Bargoni; O Caputo; MR Gasco. *Pharmacol Res.* **1999**, 40, 281-286.
- [42] GP Zara; R Cavalli; A Bargoni; A Fundarò; D Vighetto; MR Gasco. *J Drug Target.* **2002**, 10, 327-335.
- [43] S Yang; J Zhu; Y Lu; B Liang; C Yang. *Pharm Res.* **1999**, 16, 751-757.
- [44] D Pandita; A Ahuja; V Lather; B Benjamin; T Dutta; T Velpandian; RK Khar. *AAPS PharmSciTech.* **2011**, 12, 712-722.
- [45] A Lamprecht; JP Benoit. *J Control Release.* **2006**, 112, 208-213.
- [46] E Garcion; A Lamprecht; B Heurtault; A Paillard; A Aubert-Pouessel; B Denizot; P Menei; JP Benoît. *Mol Cancer Ther.* **2006**, 5, 1710-1722.
- [47] K Michaelis; M Hoffmann; S Dreis; E Herbert; R Alyautdin; M Michaelis; J Kreuter; K Langer. *J Pharmacol Exp Ther.* **2006**, 317, 1246-1253.
- [48] TC Long; N Saleh; RD Tilton; GV Lowry; B Veronesi. *Environ Sci Technol.* **2006**, 40, 4346-4352.
- [49] N Mori; A Kurokouchi; K Osonoe; H Saitoh; K Ariga; K Suzuki; Y Iwata. *Brain Res Dev Brain Res.* **1995**, 703, 184-190.
- [50] K Kakinuma; R Tanaka; H Takahashi; Y Sekihara; M Watanabe; M Kuroki. *Int J Hyperthermia.* **1996**, 12, 157-165.
- [51] X Zhang; J Xie; S Li; X Wang; X Hou. *J Drug Target.* **2003**, 11, 117-122.
- [52] EV Batrakova; S Li; DW Miller; AV Kabanov. *Pharm Res.* **1999**, 16, 1366-1372.
- [53] AV Kabanov; VY Alakhov. *Crit Rev Ther Drug Carrier Syst.* **2002**, 19.
- [54] EV Batrakova; S Li; VY Alakhov; DW Miller; AV Kabanov. *J Pharmacol Exp Ther.* **2003**, 304, 845-854.
- [55] A Marushima; K Suzuki; Y Nagasaki; T Yoshitomi; K Toh; H Tsurushima; A Hirayama; A Matsumura. *Neurosurgery.* **2011**, 1418-1426.
- [56] T Dutta; NK Jain. *Biochim Biophys Acta.* **2007**, 1770, 681-686.
- [57] RS Dhanikula; A Argaw; JF Bouchard; P Hildgen. *Mol Pharm.* **2008**, 5, 105-116.
- [58] S Soni; AK Babbar; RK Sharma; A Maitra. *J Drug Target.* **2006**, 14, 87-95.
- [59] TK Vyas; A Shahiwala; MM Amiji. *Int J Pharm.* **2008**, 347, 93-101.
- [60] A Desai; T Vyas; M Amiji. *J Pharm Sci.* **2008**, 97, 2745-2756.
- [61] HM Shubar; IR Dunay; S Lachenmaier; M Dathe; FN Bushrab; R Mauludin; RH Müller; R Fitzner; K Borner; O Liesenfeld. *J Drug Target.* **2009**, 17, 257-267.
- [62] A Himanshu; P Sitasharan; A Singhai. *IJPLS.* **2011**, 2, 945-951.
- [63] DB Vieira; LF Gamarra. *Int J Nanomed.* **2016**, 11, 5381.
- [64] S Joshi; R Singh-Moon; M Wang; DB Chaudhuri; JA Ellis; JN Bruce; IJ Bigio; RM Straubinger. *J Neurooncol.* **2014**, 120, 489-497.
- [65] A Schnyder; J Huwyler. *NeuroRx.* **2005**, 2, 99-107.
- [66] KM McNeeley; E Karathanasis; AV Annapragada; RV Bellamkonda. *Biomaterial.* **2009**, 30, 3986-3995.
- [67] H Ding; V Sagar; M Agudelo; S Pilakka-Kanthikeel; VSR Atluri; A Raymond; T Samikkannu; MP Nair. *Nanotechnology.* **2014**, 25, 055101.
- [68] H Guo; W Chen; X Sun; YN Liu; J Li; J Wang. *Carbohydr Polym.* **2015**, 118, 209-217.
- [69] P Ramos-Cabrer; F Campos. *Int J Nanomedi.* **2013**, 8, 951-960.
- [70] A Akbarzadeh; R Rezaei-Sadabady; S Davara; SW Joo; N Zarghami; Y Hanifehpour; M Samiei; M Kouhi; K Nejati-Koshki. *Nanoscale Res Lett.* **2013**, 8, 1.

- [71] A Sharma; US Sharma. *Int J Pharm.* **1997**, 154, 123-140.
- [72] W Mehnert; K Mäder. *Adv Drug Deliv Rev.* **2001**, 47, 165-196.
- [73] A Fundarò; R Cavalli; A Bargoni; D Vighetto; GP Zara; MR Gasco. *Pharmacol Res.* **2000**, 42, 337-343.
- [74] E Rostami; S Kashanian; AH Azandaryani; H Faramarzi; JEN Dolatabadi; K Omidfar. *Chem Phys Lipids.* **2014**, 181, 56-61.
- [75] Y Chen; G Dalwadi; H Benson. *Curr Drug Deliv.* **2004**, 1, 361-376.
- [76] J Diederichs; R Muller. *Pharmazeutische Industrie.* **1994**, 56, 267-275.
- [77] C Freitas; RH Müller. *Int J Pharm.* **1998**, 168, 221-229.
- [78] C Da-Bing; TZ Yang; L Wang-Liang; Q Zhang. *Chem Pharm Bull.* **2001**, 49, 1444-1447.
- [79] W Mehnert; K Mäder. *Adv Drug Deliv Rev.* **2001**, 47, 165-196.
- [80] RH MuÈller; K MaÈder; S Gohla. *Eur J Pharm Biopharm.* **2000**, 50, 161-177.
- [81] SP Vyas, RK Khar. Targeted & controlled drug delivery: Novel carrier systems, 1<sup>st</sup> edition, CBS publishers & distributors, Delhi, **2004**.
- [82] H Patel; DA Patel; PD Bharadia; V Pandya; D Modi. *Int J Pharm Life Sci.* **2011**, 2, 1006-1019.
- [83] A Yadav; H Mote. *Indian J Pharm Sci.* **2008**, 70, 170.
- [84] E Björk; P Edman. *Int J Pharm.* **1988**, 47, 233-238.
- [85] A Martinac; J Filipović-Grčić; D Voinovich; B Perissutti; E Franceschini. *Int J Pharm.* **2005**, 291, 69-77.
- [86] E Givini; G Rassu; V Sanna; M Cossu; P Giunchedi. *J Pharm Pharmacol.* **2005**, 57, 287-294.
- [87] K Nan; F Ma; H Hou; WR Freeman; MJ Sailor; L Cheng. *Acta Biomater.* **2014**, 10, 3505-3512.
- [88] D Sensoy; E Cevher; A Sarıcı; M Yılmaz; A Özdamar; N Bergişadi. *Eur J Pharm Biopharm.* **2009**, 72, 487-495.
- [89] A Kondo; H Fukuda. *J Ferment Bio Eng.* **1997**, 84, 337-341.
- [90] KJ Widder; RM Morris; GA Poore; DP Howard; AE Senyei. *Eur J Cancer Clin Oncol.* **1983**, 19, 135-139.
- [91] RS Molday; SP Yen; A Rembaum. *Nature.* **1977**, 268(5619), 437-438.
- [92] SK Jain; A Awasthi; N Jain; G Agrawal. *J Control Release.* **2005**, 107, 300-309.
- [93] AK Srivastava; DN Ridhurkar; S Wadhwa. *Acta Pharm.* **2005**, 55, 277.
- [94] A Patel; S Ray; RS Thakur. *DARU J Pharm Sci.* **2006**, 14, 57-64.
- [95] R Umamaheswari; S Jain; P Tripathi; G Agrawal; N Jain. *Drug Deliv.* **2002**, 9, 223-231.
- [96] JM Neutze; F Wyler; AM Rudolph. *Am J Physiol Renal Physiol.* **1968**, 215, 486-495.
- [97] LC Becker; NJ Fortuin; B Pitt. *Circ Res.* **1971**, 28, 263-269.
- [98] E. Ghezzi; L. Benedetti; M. Rochira; F. Biviano; L. Callegaro. *Int J Pharm.* **1992**, 87, 21-29.
- [99] D Maysinger; I Jalsenjak; AC Cuello. *Neurosci Lett.* **1992**, 140, 71-74.
- [100] S Mittal; A Cohen; D Maysinger. *Neuroreport.* **1994**, 5, 2577-2582.
- [101] A McRae-Degueurce; S Hjorth; DL Dillon; DW Mason; TR Tice. *Neurosci Lett.* **1988**, 92, 303-309.
- [102] PJ Camarata; R Suryanarayanan; DA Turner; RG Parker; TJ Ebner. *Neurosurgery.* **1992**, 30, 313-319.
- [103] D Maysinger; J Filipovic-Grcic; AC Cuello. *Neuroreport.* **1993**, 4, 971-974.
- [104] CE Krewson; R Dause; M Mak; WM Saltzman. *J Biomater Sci Polym Ed.* **1997**, 8, 103-117.
- [105] J Filipović-Grčić, D Maysinger, I Jalšenjak. In: Book Poly (L-lactide) co-glycolide microspheres with ciliary neurotrophic factor, **1998**.
- [106] AR Menjoge; RM Kannan; DA Tomalia. *Drug Discov Today.* **2010**, 15, 171-185.
- [107] L Xu, WA Yeudall, H Yang. In Tailored Polymer Architectures for Pharmaceutical and Biomedical Applications, ACS Publications, **2013**, 197-213.
- [108] W Ke; K Shao; R Huang; L Han; Y Liu; J Li; Y Kuang; L Ye; J Lou; C Jiang. *Biomaterials.* **2009**, 30, 6976-6985.
- [109] Y Liu; R Huang; L Han; W Ke; K Shao; L Ye; J Lou; C Jiang. *Biomaterials.* **2009**, 30, 4195-4202.
- [110] RQ Huang; YH Qu; WL Ke; JH Zhu; YY Pei; C Jiang. *FASEB J.* **2007**, 21, 1117-1125.
- [111] U Gupta; HB Agashe; NK Jain. *J Pharm Pharm Sci.* **2007**, 10, 358-367.
- [112] KM Kitchens; ME El-Sayed; H Ghandehari. *Adv Drug Deliv Rev.* **2005**, 57, 2163-2176.
- [113] PV Kumar; A Asthana; T Dutta; NK Jain. *J Drug Target.* **2006**, 14, 546-556.
- [114] A Agarwal; A Asthana; U Gupta; NK Jain. *J Pharm Pharmacol.* **2008**, 60, 671-688.
- [115] A Agarwal; U Gupta; A Asthana; NK Jain. *Biomaterials.* **2009**, 30, 3588-3596.
- [116] V Gajbhiye; N Ganesh; J Barve; NK Jain. *Eur J Pharm Sci.* **2013**, 48, 668-679.
- [117] C Kojima; T Suehiro; K Watanabe; M Ogawa; A Fukuhara; E Nishisaka; A Harada; K Kono; T Inui; Y Magata. *Acta Biomater.* **2013**, 9, 5673-5680.
- [118] H He; Y Li; XR Jia; J Du; X Ying; WL Lu; JN Lou; Y Wei. *Biomaterials.* **2011**, 32, 478-487.
- [119] Y Li; H He; X Jia; WL Lu; J Lou; Y Wei. *Biomaterials.* **2012**, 33, 3899-3908.

- [120] R Huang; W Ke; Y Liu; C Jiang; Y Pei. *Biomaterials*. **2008**, 29, 238-246.
- [121] Q Yuan; E Lee; WA Yeudall; H Yang. *Oral Oncol*. **2010**, 46, 698-704.
- [122] S Huang; J Li; L Han; S Liu; H Ma; R Huang; C Jiang. *Biomaterials*. **2011**, 32, 6832-6838.
- [123] R Huang; W Ke; L Han; J Li; S Liu; C Jiang. *Biomaterials*. **2011**, 32, 2399-2406.
- [124] Q Yuan; Y Fu; WJ Kao; D Janigro; H Yang. *ACS Chem Neurosci*. **2011**, 2, 676-683.
- [125] H Wong; A Rauth; R Bendayan; X Wu. *J Control Release*. **2006**, 116, 275-84.
- [126] AV Kabanov; EV Batrakova; VY Alakhov. *J Control Release*. **2003**, 91, 75-83.
- [127] Y Gao; LB Li; G Zhai. *Colloids Surf B*. **2008**, 64, 194-199.
- [128] Z Zhou; C Chaibundit; Ad'emanuele; K Lennon; D Attwood; C Booth. *Int J Pharm*. **2008**, 354, 82-87.
- [129] C Zhang; G Qu; Y Sun; X Wu; Z Yao; Q Guo; Q Ding; S Yuan; Z Shen; Q Ping. *Biomaterials*. **2008**, 29, 1233-1241.
- [130] RR Sawant; RM Sawant; VP Torchilin. *Eur J Pharm Biopharm*. **2008**, 70, 51-57.
- [131] A Mistry; S Stolnik; L Illum. *Int J Pharm*. **2009**, 379, 146-157.
- [132] L Illum. *J Pharm Pharmacol*. **2004**, 56, 3-17.
- [133] D Betbeder; S Spérando; JP Latapie; J de Nadaí; A Etienne; JM Zajac; B Francés. *Pharm Re*. **2000**, 17, 743-748.
- [134] Q Zhang; X Jiang; W Jiang; W Lu; L Su; Z Shi. *Int J Pharm*. **2004**, 275, 85-96.
- [135] M Kumar; A Misra; A Babbar; A Mishra; P Mishra; K Pathak. *Int J Pharm*. **2008**, 358, 285-291.
- [136] M Kumar; A Misra; A Mishra; P Mishra; K Pathak. *J Drug Target*. **2008**, 16, 806-814.
- [137] X Gao; W Tao; W Lu; Q Zhang; Y Zhang; X Jiang; S Fu. *Biomaterials*. **2006**, 27, 3482-3490.
- [138] SM Hussain; AK Javorina; AM Schrand; HM Duhart; SF Ali; JJ Schlager. *Toxicol Sci*. **2006**, 92, 456-463.
- [139] J Wang; MF Rahman; HM Duhart; GD Newport; TA Patterson; RC Murdock; SM Hussain; JJ Schlager; SF Ali. *Neurotoxicol*. **2009**, 30, 926-933.
- [140] X Deng; Q Luan; W Chen; Y Wang; M Wu; H Zhang; Z.Jiao. *Nanotechnol*. **2009**, 20, 115101.
- [141] G Oberdörster; Z Sharp; V Atudorei; A Elder; R Gelein; W Kreyling; C Cox. *Inhal Toxicol*. **2004**, 16, 437-445.
- [142] TR Pisanic; JD Blackwell; VI Shubayev; RR Fiñones; S Jin. *Biomaterials*. **2007**, 28, 2572-2581.
- [143] SB Raymond; LH Treat; JD Dewey; NJ McDannold; K Hynynen; BJ Bacska. *PloS one*. **2008**, 3, e2175.
- [144] HL Liu; MY Hua; PY Chen; PC Chu; CH Pan; HW Yang; CY Huang; JJ Wang; TC Yen; KC Wei. *Radiology*. **2010**, 255, 415-425.
- [145] PY Chen; HL Liu; MY Hua; HW Yang; CY Huang; PC Chu; LA Lyu; IC Tseng; LY Feng; HC Tsai. *Neuro Oncol*. **2010**, 54.
- [146] CX Deng; X Huang. *Ther Deliv*. **2011**, 2, 137-141.