



## Effect of Growth Regulator on Morpho-Physiological Attributes of Chilli: A Case Study

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### ABSTRACT

The present study was undertaken with F1 hybrid JUGNI-070 variety of chilli. This solanaceous vegetable was cultivated in order to observe the effect of four growth regulators viz; Indole-3-Acetic Acid, Gibberellic acid, Putrescine and Salicylic acid of some parameters during kharif season of 2017-2018. The parameters are plant height, number of leaves, number of stem, number of fruits and average weight of the fruits. The field allotted was divided into ten ridges Rows 1-10. In R1-R2 was allocated as control, in R3-R4 IAA @ 1 ppm was sprayed and R5-R6 Salicylic acid @ 1ppm was sprayed and R7-R8 Putrescine @ 1ppm was sprayed, R9-10 GA3 @ 1ppm was sprayed, all the plant growth regulators were sprayed once at 30 days after transplanting and the results were observed after 5th and 10th days of all rows. The data was collected from randomly selected plants of each rows and the mean data was calculated. The results obtained from each rows were compared. The results revealed showed significant improvement in overall growth and development of plant in R9 rows, where plant growth regulator GA3 was used as indicated by the increase in plant height, number of leaves and number of stems. Where as in R7-8 where Putrescine was sprayed there was no significant increase in overall growth and development of the plant only the number of bud formation was observed high i.e. whereas R5 & R6 where salicylic was sprayed but there is no growth is observed, whereas R3-4 IAA were used in which it results the stem development is observed.

**Keywords:** Agriculture; Biotic; Chilli; Dose; Energy; Forage; Gap

### INTRODUCTION

Chilli has been used since ancient times, traditionally in the form of spice. It is also used as a natural flavor and colorant in food industry as well as raw material for the pharmaceutical industry. Chilli is nutritious crop, every 100 gm of green and dry chilli yield about 229 and 297 calories of energy [1]. It is mainly cultivated for three constituents of fruits viz., capsaicin, capsanthin and oleoresin [2]. It is grown for its pungent fruits which are used both as green and ripe to impart pungency and flavour to the food. Pungency, one of the important attributes of Capsicum species is due to the presence of alkaloid 'capsaicin' in the fruit. It is used primarily in the flavouring of pickles, meats, barbecue sauces, ketchup, cheese, snack food, dips, chilli cake, salads, and sausages [3]. As a medicinal plant, the capsicum species has been used as a carminative, stomachic, stimulant, rubefacient and tonic. It prevents heart diseases by dilating blood vessels. Chilli stimulates saliva and gastric juices and aids in digestion. Oleoresin of capsicum is used in pain balms and vapour rubs. Chilli (*Capsicum annuum* L.) is an important spice

crop grown for its fruits, which are used in green as well as ripe dried form for its pungency. Chilli belongs to the genus *Capsicum* family Solanaceae. There are mainly four cultivated *Capsicum* species and they are originated from South America. Commercial cultivation of chilli is mostly confined to the tropical regions of the world, since it requires long and warm season for its growth and development. Chilli is known from prehistoric remains in Peru and was widely cultivated in Central and South America in early times. It was first introduced in India by Portuguese towards the end of 15th century. Now-a-day's chilli has become an important crop all over India. In India Chilli is cultivated in all the states including Andhra Pradesh, Karnataka, Maharashtra, Orissa, Rajasthan, Tamil Nadu and West Bengal. Andhra Pradesh alone contributes 46 per cent of total chilli production of India. India produces 9.7 lakh tonnes of dry chilli from an area of 9.08 lakh hectares [4]. No country in the world has so much area and production of chilli as in India. In the world green chilli is grown over an area of 14.53 lakh ha with a production of 190.39 lakh tones, accounting for a productivity of 131.02 q per ha [4]. Chilli has a great genetic diversity for all fruits similar diversity per se exists for all phenotypic, biochemical and reaction to biotic and abiotic stresses in chilli germplasm appears in different sizes, shapes and colour. The fruit size of some varieties is more than hundred times that of others. The shape may be elongated or round, and distal end pointed, blunt or sucked in, corrugated, leathery or smooth may be the touch on outer skin.

### MATERIALS AND METHODS

The present investigation was carried out under field condition in the "Agricultural Research Field, School of Agriculture, Lovely Professional University, Phagwara, and Punjab 144411. The Lovely Professional University is situated in the south of NH-1 (G.T. Road) in the state of Punjab at a distance of about 350 km from Delhi. The study area falls in chaheru village of kapurthala district. Chaheru village is situated between 31°15' North latitude, 75°42' East latitude at an altitude of 228 meters above the mean sea level (MSL). The district lies in the center of Punjab and is situated between two rivers Sutlej and Beas. The soil of the village is alluvial and varies from coarse loamy to fine loamy. The total agriculture land of the district is 134 hectare. Major source of irrigation is tube wells, bore wells and then pump sets. Only 3% of land is irrigated by canals.

#### Climatic Condition

The area falls under sub humid area. The temperature reaches above 40 degree Celsius during summer and during winter the temperature goes down below 10 degree Celsius. The hottest month is June and the coldest month being January. The average temperature of the district is 23.8 degree Celsius and the average annual rainfall is 719mm. the highest rainfall is recorded in month of July and driest month is November. The temperature during transplantation was 28 degrees Celsius. Chilli is warm season crop and is susceptible to frost. The optimum temperature is 20-30 degree Celsius. When the temperature goes below 15 degree Celsius it severely affects the growth and development of the crop.

The methods and materials are used as follows:

#### Rising of nursery

After the variety was selected the seeds of (f1 hybrid of jugni 070 variety) chilli were sown on the raised nursery bed. All necessary agronomic practices were followed to raise the crop.



Figure 2: Raising the nursery of chilli plant (Source: Photograph by authors, Unpublished, 2018)



Figure 3: Field preparation for transplanting of chilli plant (Source: Photograph taken by authors, Unpublished, 2018)

### Field preparation and transplanting

The weeds and previous crop stubs were removed from the field. The experimental field was well prepared by one deep ploughing and two cross ploughing were given to improve field condition. The field was divided into 10 rows. When seedlings were of 30 days old they were transplanted to main field in rows at the depth of 2.5-3.0 cm. Row to Row and Plant to Plant distance was maintained at 90 X 90cm, respectively. 54 seedlings are transplanted in each row (figure 3).

### Fertilizer application

The fertilizer was applied as per the recommended dosage given in the package of practice of chilli crop. The full dose of fertilizers of urea was given at the time of transplanting as basal dose remaining half dose of urea was given at the time of second irrigation. FYM (Farm yard Manure) and Vermi compost were given (Figure 4).



**Figure 4: FYM and Vermicompost application in chilli field (Source: Photograph by authors, unpublished, 2018)**

**Weeding:** First weeding was done at 15 days after transplanting with the help of ‘Khurpi’ and second weeding were done at 30 days old transplanting respectively. 10days after transplanting, irrigation was provided after transplanting in the field (Figure 5).



**Figure 5: Irrigation and weeding in the chilli field (Source: Photograph by authors, Unpublished, 2018)**

### **Intercultural Operation**

Earthing up was done 20 days after transplanting so that the plant can get proper support and have proper growth and development . Vermi compost was applied to all the plants 10 days after transplanting to protect plant leaves.

### **Soil Testing**

Total of four soil sample was collected from the plot number 30 which was allotted to us. The soil was then tested for pH, electrical conductivity. The soil Ec and pH was tested with the help of EC and pH meter.

### **Preparation of spray solution**

The finished spray solution were prepared by the following formula-

Amount of formulation =  $\frac{\text{Conc. required (\%)} \times \text{Volume required (lit.)}}{100}$

*a. i.* in the PGR

### PGR used

Four plant growth regulator viz. IAA, Salicyclic acid, Putriscine and Gibberellic acid are used in the experiment. The different concentrations of these plant growth regulators are used during experimentation. The first foliar spray was done when the crop was 30 days old. The observation was taken for the growth and development of the plant. The following parameters are observed at 5 and 10 days after spraying of the plant growth regulator-

- Plant Height
- Number of leaves
- Number of stems
- Number of buds
- Number of fruits
- Average weight of fruits

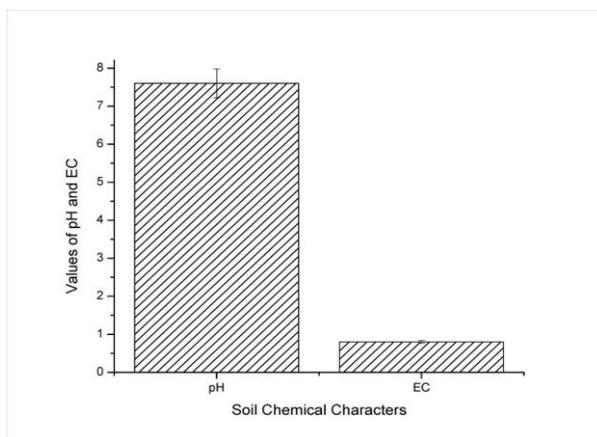
The plot allotted to us has the area of 180 meter square. The plot was then further divided into 10 rows of equal length and breadth; there are ten numbers of rows and R-3, 4 the plant was sprayed with IAA @ 1 ppm. In the R5-6 all the plants was sprayed with salicyclic acid@ 1 ppm of water and the R7-8 all the plants were putriscine @ 1ppm of water, R9-10 all the plants were sprayed with Gibberellic acid@ 1ppm and R1-2 was left as control (Table 1).

**Table 1: Calendar of the experiment**

Sl. No	Particulars	Dates
1.	Land Preparation	23/09/2017
2.	Plot size	15.9m x 9.9m
3.	Ridges Preparation	24/09/2017
4.	First irrigation	25/10/2017
5.	Transplanting in main field	26/10/2017
6.	Variety name	Jugni-070
7.	Weeding	31/10/2017
8.	Second irrigation	02/11/2017
9.	Fertilizers& Vermi-compost	27/10/2017
10.	Row to row& Plant – plant distance	90 x40cm
11.	Application of PGR'S	25/11/2017

### RESULTS AND DISCUSSION

Soil sampling and water sampling was both done from the samples collected from the field. Four samples were collected from the plot number 49 which was allotted to us. EC (electrical conductivity), OC (organic carbon), pH, AK (available potassium), AP (available phosphorous) was checked all the four samples and the average value was calculated (Figure 6).

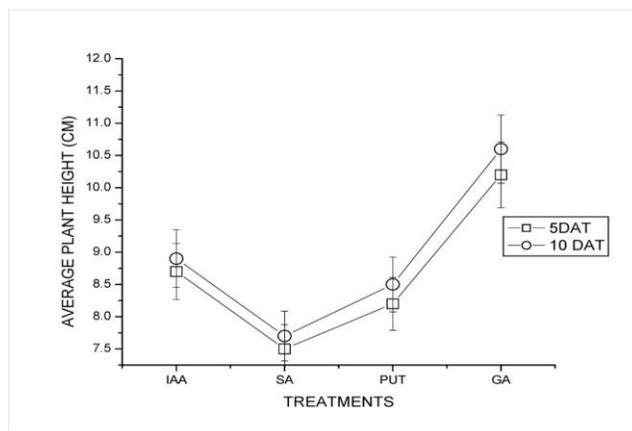


**Figure 6: Result of soil sampling of collected soil samples (Source: Drawn by authors, unpublished, 2018)**

The pH of soil is on average 7.3 which is normal to basic. The soil is open textured with high massive structure. This soil appears as brown or reddish brown, sandy loam.

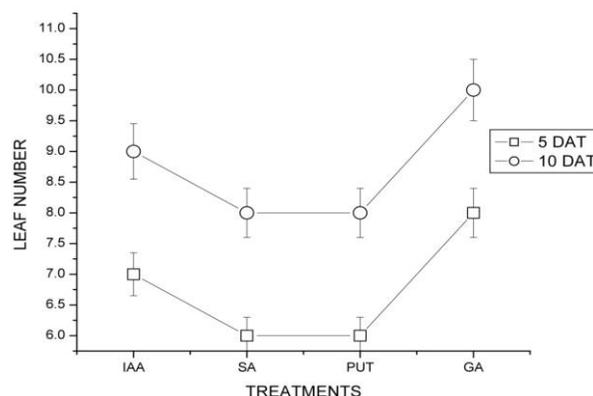
### Observation of Crop

Five plants were randomly selected from the 8 rows and observed after 5th and 10th day of first spray. The observation was based on the four parameters: height of the plants, number of leaves, number of stems and number of flower buds. The observation of a number of fruits and weight of fruits could not be done since the fruiting has not started yet. The data from all five plants were collected and the mean was taken out for each treatment so that a comparative study could be done. The data collected are represented in the graph given below. The figure 7, which is given below depicts the effect of PGR application at 30 days after transplanting. Five plants from each plot was selected and measured after 5 and 10 days of spraying. After the 5 days of spraying we can see the average plant height at R3-4 is 8.7 cm. In case of R-5, 6 the difference is from 7.5 cm as observed at 5 days of spraying. In R-7, 8 the difference is 8.2 cm. In R9-10 the average plant height is 10.2 cm. In graph 7, we can have a comparative study of the mean height of the plants where it can be observed that there is rapid growth of average plant height in R 1 - 10.



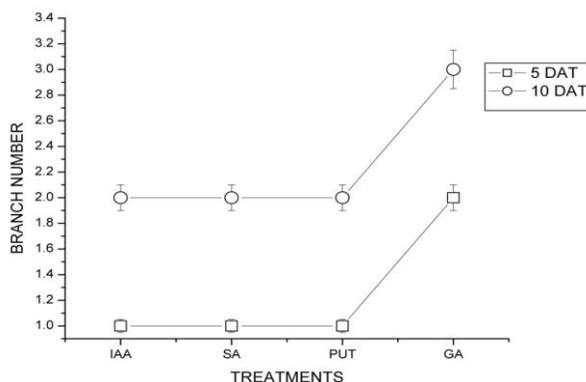
**Figure 7: Average plant height of chilli (Source: Drawn by authors et al., 2018, unpublished, LSD: 5%)**

Figure 8, which is given below depicts the effect of plant growth regulators viz. IAA, Salicylic acid, Putriscine and Gibberellic acid on the mean number of plant leaves at 30 days after transplanting. Spraying was done 30 days after transplanting and the leaves were counted at 5 and 10 days after spraying. The data were collected from five randomly selected plants at each observation and mean number was calculated. There is not much difference between the mean number of leaves between R3-10.



**Figure 8:** Average leaf number of chilli (Source: Drawn by authors *et al.*, 2018, unpublished, LSD: 5%)

Figure 9 which is given below shows the mean number of branches at different days' intervals of PGR application. The number of branches in plants also indicates the growth and development of the plant, thus healthier plants with high yields have a higher number of stems to support the maximum number of flowers and fruits. The average number of branches during the 5th and 10th days' observation after the spray was high in R 2-3 and low control. We could see this by observing the data taken at 5th and 10th day after the spraying. On the 10th day after the spray the mean number of branches in the plants of R3-10 was five which is high compared to the mean of 2.6 number of branches present in control. In R5-6 the mean number was 4 neither low nor high. By this observation, we can say that the plot R3-10 sprayed by growth regulators had increased numbers of branches than R1-2 i.e. control.



**Figure 9:** Average branch number of chilli (Source: Drawn by authors *et al.*, 2018, unpublished, LSD: 5%)

The result of this investigation clearly shows that the use of plant growth regulator 1 ppm IAA helped in rapid growth and development of the chilli crop. Whereas gibberellic acid@1 ppm helped in increasing the flowering and fruiting in chilli crop. Salicylic acid sprayed at after 30 days of transplanting showed successful results in the growth promotion of the chilli crop. The result shows that the use of gibberellic acid increased the number of flower bud formation whereas it had no effect in the overall growth and development of the crop.

### CONCLUSION

The observation was done in 10 rows divided as R1-2 in which in control, R3-4 were only IAA was sprayed, R5-6 were only sprayed with salicylic acid, R7-8 were only sprayed with putrescine R 9-10 were only sprayed with GA3 [5-8]. After carefully observing the plants from the ten rows we could find out that the plant sprayed with 1 ppm GA3 showed rapid growth in height and the number of leaves and the numbers of branches were also high in number in plants of this rows. Since flowering and fruiting has not started yet we observed the number of buds in the

plants. The number of buds formation was not affected by the spray of GA<sub>3</sub> since no drastic increase or decrease in bud formation was observed. Thus through our findings we can conclude that the use of 1 ppm GA<sub>3</sub> of water as foliar spray at 30 days after transplanting increases the rate of growth and development of the plant by rapidly increasing the height of the plant, number of leaves, number of branches whereas the foliar spray of salicylic acid of water during 30 days of transplanting rapidly increases the flower bud formation thus increasing the number of flowers. Thus plant growth regulators viz. IAA, salicylic acid, putrescine and gibberellic acid @ 1 ppm can be recommended to chilli crop [9-11].

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#### REFERENCES

- [1] Abeles FB, Morgan PW, Saltveit ME. *J Agri.* **1992**, 11, 133-137.
- [2] Akhtar H. Effect of Azotobacter inoculation and auxin precursor, L-tryptophan on the growth and yield of tomato (*Lycopersicon esculentum* L.). M.Sc. Thesis. Institute of Soil and Environmental Science, University of Agriculture, Faisalabad. **1994**.
- [3] Arshad M, Frankenberger WT. *Plant and Soil.* **1990**, 122, 219-227.
- [4] Arshad M, Frankenberger WT. *Plant Soil.* **1991**, 133, 1-8.
- [5] Davies PJ. The plant hormones: their nature, occurrence and functions. In: Davies PJ (ed.) Plant hormones and their role in plant growth and development. Martinus Nijhoff Publishers Dordrecht, the Netherlands **1987**, 1-11.
- [6] Edison HS. The effect of growth regulator IAA on the growth and development of tomato fruit. Balai Penelitian Hortikultura, Solok (Indonesia) **1991**, 30, 9-11.
- [7] Maharana T, P. Pati, GS Sahu. *Envir Ecol.* **1990**, 8, 1327-1328.
- [8] Martens DA, Frankenberger WT. *Soil Soil Biochem.* **1993**, 25, 1679-1687.
- [9] Parvez MA, Muhammad F, Ahmad MP. *J Bio Sci.* **2000**, 3(7), 1154-1155.
- [10] Patten CL, Glick BR. *Appl Environ Microbiol.* **2002b**, 68, 3795-3801.
- [11] Rovira AD. Plant root exudates and their influence upon soil microorganisms. pp. 170-184, in Baker KF and Snyder WC (eds). Ecology of soil-borne plant pathogens, Prelude to Biological Control. An international symposium on factors determining the behavior of plant pathogens in soil, University of California, Berkeley, April 7-13, 1963. University of California Press. Russell, RS 1982. *Plant root systems*. ELBS, UK, **1965**, 16-17.