



## Development of FIA system for the spectrophotometric determination of hydroquinone in pure material and pharmaceutical formulations

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### ABSTRACT

This article deals with the development of flow injection system for the spectrophotometric determination of Hydroquinone drugs. The analytical system is based on oxidation reaction using  $\text{KMnO}_4$  as oxidizing agent for conversion of HQ to P-benzoquinone (BQ) in an alkaline medium, the absorbance is measured at 610nm. All different chemical and physical experimental parameters affecting on the development and stability of the colored product were carefully studied. The parameters optimized by using  $20 \mu\text{g}\cdot\text{ml}^{-1}$  standard solution of HQ. Beer's law is obeyed over the concentration range of 1-26 and 3-125  $\mu\text{g}\cdot\text{ml}^{-1}$  of Hydroquinone with detection limits of 0.0125 and 0.25  $\mu\text{g}\cdot\text{ml}^{-1}$  of Hydroquinone for spectrophotometric and FIA- Merging zone, respectively. The color product formed by passing of a 0.1 M sodium hydroxide as carrier at flow rate of  $2.12 \text{ml}\cdot\text{min}^{-1}$  is merged with a volume of 227.65  $\mu\text{L}$  of sample and the volume of 227.65  $\mu\text{L}$  of oxidant agent to conclude a final methodology for the determination of HQ accommodating 227.65  $\mu\text{L}$  as injection sample. Under these conditions, the FIA system allowed to analyze was about 55 samples per hour. The method was successfully applied to the determination of HQ in Pharmaceutical formulations without any interference from common excipients used as additives. The results agree favorably with standard method.

**Key words:** Hydroquinone, Flow injection, oxidization reaction, spectrophotometric determination,  $\text{KMnO}_4$ .

### INTRODUCTION

Hydroquinone (HQ) is used as a developer in black and white photography [1]. An antioxidant for fats and oils a polymerization inhibitor, a stabilizer in paints, varnishes, motorfuels, oils, an intermediate for rubber processing chemicals in the production of mono and dialkyl ethers and as de-pigmenting agent [2]. Besides its positive and beneficial utilization, it bears some harmful and toxic aspects as well, which may produce serious health complications due to its release especially in water and air from mentioned and other sources. The possible health problems include irritation of skin, eyes, nose and throat, dizziness, headache, unconsciousness, tinnitus, breathing difficulties and others [3]. It has also been reported as a nephrocarcinogenic reagent [4]. It is extensively used in skin – toning preparations or skin lightening cosmetics and suggested to be effective at 1.5 -2.0% [1,5-8].

It is recommended that HQ must be used under prescription because its long –term contact in concentrations of greater than 5% can produce various side effects [6]. These side effects may be acute or chronic. Acute side effects are allergic and irritant contact dermatitis, post-inflammatory hyperpigmentation and nail discoloration. High concentrations of HQ (above 5-6%) have been implicated in persistent hypopigmentation or depigmentation, a condition known as leukoderma. Exogenous ochronosis is a major chronic side effect of HQ. This condition is characterized by reticulated, ripple-like, sooty pigmentation on the forehead, cheeks and other areas of HQ application [7]. Despite its numerous useful applications, HQ has been reported as mutagenic in animals [8] and a possible nephrocarcinogen [4]. According to other report [9] mononuclear compounds such as benzene metabolites, caffeic acid and o-toluidine should express their carcinogenicity through oxidative DNA damage.

Several techniques have been reported in the literature for estimation of HQ in cosmetics such as voltammetry [10,11] high performance liquid chromatography (HPLC) [8,12-14] in different types of samples, titrimetry [15,16], Capillary electrochromatography (CEC) [17], Fluorimetry [18], GC/MS [19], electrochemical methods [20-22], chemiluminescence [1,23-26]. Colorimetric analysis of small amount of HQ in styrene has also been reported [27, 28], flow injection analysis [29,31]. Spectrophotometric determination of HQ has been cited elsewhere [32-36]. Due to easy instrumentation, low equipments, running cost, better repeatability, it's a large throughput per hour, broad linear dynamic range and easy sample injection and preparation, the flow injection analysis technique such as FIA, rFIA, stop-flow or merging zone have clear advantages over the other techniques for the quantitative determination of environmentally toxic organic compounds in complicated matrices [37] and these techniques have some limitations such as use of expensive and or toxic ligands for complex formation and hence its expensive use for HQ complex determination in aqueous samples [32,34,38].

In contrast, our newly developed method is very fast, simple, economical and the main aim optimization is to find the experimental conditions which give the best response. Moreover, it has lower detection limits, better sensitivity and better application range for dilute aqueous samples where matrix effect minimizes the interfering effect of ions or reagents.

## EXPERIMENTAL SAECTION

### Apparatus

All spectral and absorbance measurements were carried out by using a Shimadzu UV-Visible, 1200 digital double beam recording spectrophotometer (Japan), with 1 cm quartz cells Biotech. Engineering Management CO.LTD.UV-9200(UK). A quartz flow cell with 100  $\mu$ L internal volume and 1 cm bath length (U.S.A) was used for the absorbance measurements. A one channel manifold (Figure 10) was employed for the FIA spectrophotometric determination of Hydroquinone. A peristaltic pump (YZ1515X, China) was used to transport the reagents solutions. Injection valve (6-three ways, Merging zone version, Home made) was employed to provide appropriate injection volumes of standard solutions and samples. Tubes; A- sample and reagent loops, made of teflon (0.5 mm internal diameter) B-Flexible vinyl tubing of 2mm with internal diameter tubing. Reaction coil was made of glass (I.D; 1.5 mm, length 100 cm, home made). Sodium hydroxide (figure 10) as carrier was combined with injected sample (Hydroquinone) as Loop<sub>1</sub> and reagent (KMnO<sub>4</sub>) as Loop<sub>2</sub>, then they merged with carrier (NaOH), mixed in reaction coil (RC) with length of 100 cm, injection sample loop (227.65  $\mu$ L) and reagent volume (227.65  $\mu$ L), flow rate of 2.12 ml/min, the absorbance was measured at 610 nm and at room temperature.

### Reagents

Working Hydroquinone standard material was provided from state company for Drug Industries and Medical appliance (SDI) Sammara-Iraq of (99% purity) and standard solution of 500  $\mu$ g.ml<sup>-1</sup> was freshly prepared by dissolving 0.0125 gm of hydroquinone in 5 ml sodium hydroxide 2M and then diluted with distilled water to the mark with 100 ml volumetric flask. Potassium permanganate of (99% purity) was obtained from Merck (Germany) a standard solution of 0.02  $\mu$ g.ml<sup>-1</sup> was freshly prepared by dissolving 0.158 gm of KMnO<sub>4</sub> in 50 ml distilled water. Sodium hydroxide (98% purity) from (RDL), solution of 2M was prepared by dissolving 8 gm in 100 ml distilled water, 200  $\mu$ g.ml<sup>-1</sup> of various interferences by dissolving 0.125 gm in 250 ml distilled water. Dosage forms were obtained from commercial sources.

### Procedure of Pure drug

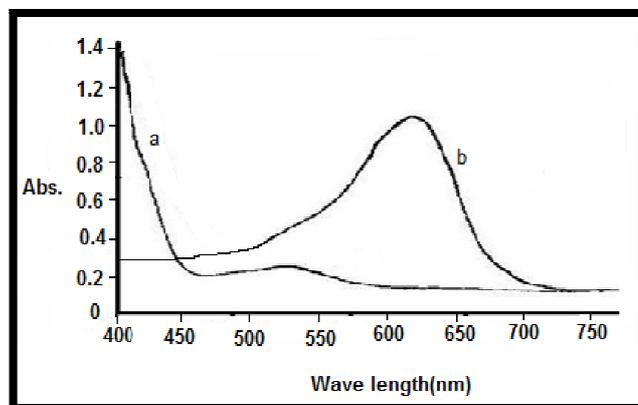
An aliquot of sample containing 20  $\mu$ g.ml<sup>-1</sup> of HQ was transferred into a series of 25 ml standard flask to cover the range of 1-26  $\mu$ g.ml<sup>-1</sup>. A volume of 1.5 ml of 0.02M KMnO<sub>4</sub> solution and 2 ml of 2M sodium hydroxide solution were added. The contents of flasks were diluted to the mark with distilled water, mixed well and left for 30 min. The absorbance was measured at 610 nm (at room temp.). The color of the oxidant formed is stable for more than 4 hr. For optimization of conditions and in all subsequent experiments, a solution of 500  $\mu$ g was used and the final volume was 25 ml (i.e. 20  $\mu$ g/ml).

### Analysis of Commercial dosage forms

For the preparation of stock solution of lightening cream, 0.25 gm of the each sample was taken in a pre-weighed beaker and 20 ml of methanol was added and thoroughly mixed using a glass rod, then completed to the mark with 0.1M NaOH. 1 ml of this solution was mixed with 1.5 ml of 0.02 M KMnO<sub>4</sub> in a 25 ml volumetric flask, this mixture was diluted with D.W. The sample thus prepared was transferred to the quartz cell and absorbance recorded in the same way as mentioned above, Cream samples were also prepared according to the method reported earlier [5] and the result were compared with those of the newly developed method.

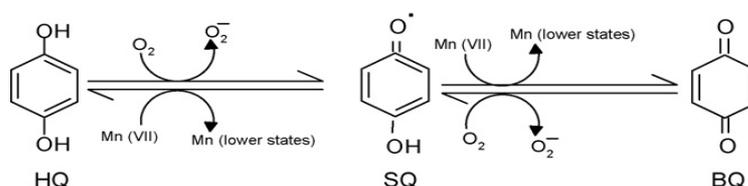
## RESULTS AND DISCUSSION

Hydroquinone (HQ) is slowly oxidized to BQ via a semiquinone (SQ) in alkaline medium [9] yield highly soluble colored which can be utilized as a suitable assay procedures for hydroquinone. The green colored product have maximum absorption at 610 nm. The blank at these wave length shows zero absorbance (Figure1).



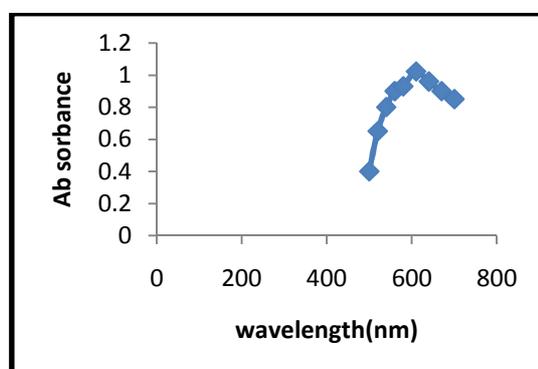
Figure(1):Absorption spectra of b ( $20 \mu\text{g}\cdot\text{ml}^{-1}$ ) of HQ treated as described under procedure and measured against a reagent blank and a the reagent blank measured against distilled water

This conversion of HQ into BQ was taken as the basis for ultra-trace determination of HQ in aqueous solutions where the formation of greenish solution is responsible for absorption due to catalytic oxidation of the HQ in to BQ by  $\text{KMnO}_4$ . The conversion of HQ into BQ brings structural changes from benzenoid to quinoidring according to chromophore theory[39]. A possible proposed mechanism for conversion of HQ into BQ via SQ can be worked out for Mn (VII) of  $\text{KMnO}_4$ , as shown in figure(2).



Figure(2): Proposed mechanism for conversion of HQ into SQ and BQ

According to this mechanism, the spectrum of HQ also disappears and the spectrum of BQ dominates. This proves the involvement of Mn (VII) of permanganate for oxidation of HQ into BQ with subsequent reduction of Mn (VII) in to lower state. The reduction of Mn (VII) into lower state is also evident from the lowering and the disappearance of Mn (VII) spectrum which is present at lower concentration of analyte in a range of 400-750nm. The role of  $\text{O}_2$  in second step is not essential but can assist Mn (VII) for its catalytic action.



Fig(3):- Change of absorbance with wavelength

### Optimization of parameters

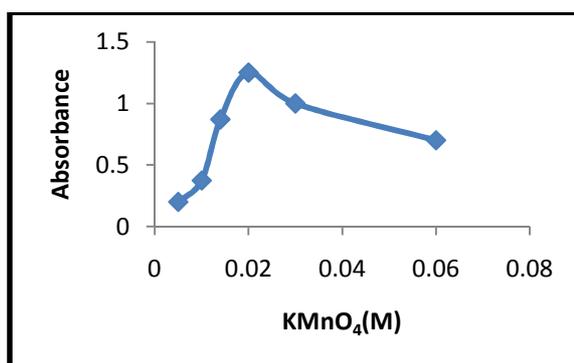
The effect of various variables on the color development was studied to establish the optimum conditions for the accurate analysis of HQ.

#### Optimization of analytical wavelength

In the absence of interferences, the wavelength chosen for a quantitative determination in the wavelength 610nm of maximum absorbance. Sometimes the adding coloring reagent may absorb very close to the same region were the determining substance does. Figure(3) describes the absorbance of 20  $\mu\text{g}\cdot\text{ml}^{-1}$  HQ solution as BQ at different wavelength after 30 min with 0.02 M  $\text{KMnO}_4$  solution.

#### Optimization of mixing amount of $\text{KMnO}_4$ solution

The effect of adding various amount of  $\text{KMnO}_4$  solution on absorbance of 0.00454 M HQ solution is given in Figure(4), the highest absorbance value was observed at 0.02M of  $\text{KMnO}_4$  after 30 min mixing of the HQ in alkaline medium (NaOH) and  $\text{KMnO}_4$  then measured at 610 nm.



Fig(4):Effect of  $\text{KMnO}_4$  on absorbance of HQ

#### Effect of order of adding reagent

Order of adding reagent plays very important role in accuracy of results and peak enhancement. In the present study it was observed that the addition of 1.5 ml of 0.02 M  $\text{KMnO}_4$  to 1 ml of 500  $\mu\text{g}\cdot\text{ml}^{-1}$  HQ, then added 2 ml of 2 M NaOH and dilution with water up to 25 ml resulted in a lower absorbance value. The greatest absorbance value was observed when we first took 1 ml of 500  $\mu\text{g}\cdot\text{ml}^{-1}$  HQ, then added 2ml NaOH solution (2M) and added 1.5 ml of 0.02 M  $\text{KMnO}_4$  through mixing and diluting this in 25 ml volumetric flask to the mark with distilled water.

#### Optimization of time for development of stable color

In UV-Vis spectrophotometry the main problem is the instability of the absorbance value. The effect of time on absorbance presented in Figure(5).

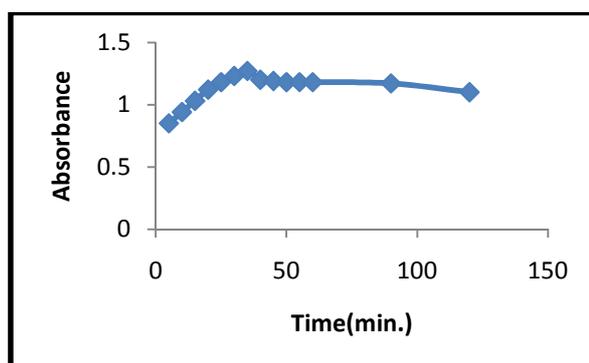


Fig.(5): Dependence of absorbance on mixing time

It is evident from fig(5) that the solution is quite stable within the range studied. However, 30-35 min. after mixing was chosen as optimum time for further study so that one could prepare, mix and process the solution according to his analytical skill. The stable color development in lesser time is due to tremendous oxidizing power of  $\text{KMnO}_4$  for conversion of HQ into BQ.

### Effect of temperature

The reaction between HQ and  $\text{KMnO}_4$  in the presence of alkaline solution (NaOH) was found to be instantaneous. However, the reaction is complete within 15 min. at room temperature ( $25^\circ\text{C}$ ), but 35min was sufficient to get maximum intensity and stability color after the addition of oxidizing agent and distilled water to final solution. The effect of temp. in range of  $5\text{--}50^\circ\text{C}$  were studied and the result obtained in figure(6) shows that the greatest absorbance at room temperature.

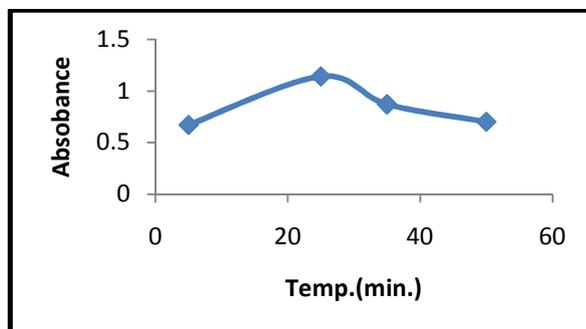
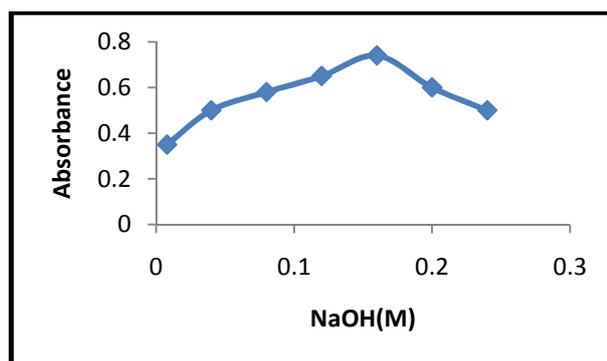


Fig.(6): Effect of temperature on absorbance

### Effect of sodium hydroxide concentration

It was found that the presence of a base led to increase the intensity of the coloured product, so 0.16M of NaOH was selected which was found that the best volume equal to 2ml of this base give high sensitivity which selected in subsequent, as shown in figure(7).



Figure(7): Effect of NaOH concentration on absorbance

### Interference study

The effect of some foreign compounds, which often found in pharmaceutical products, were studied by adding different amounts organic compounds to 1ml of  $500\ \mu\text{g}\cdot\text{ml}$  of HQ. The color was developed following the recommended procedure described earlier. It was observed that organic molecules were not interfering except glucose with the determination at levels found in dosage forms, As shown in table(1).

Table (1): Interference effect of various organic molecules during HQ determination

Interference( $200\ \mu\text{g}\cdot\text{ml}^{-1}$ )	Lactose	Maltose	Glucose	Sodium citrate	EDTA
Conc. of HQ found	-11.092	-10.586	-18.34	-1.16	2.431
E%	-11.853	-11.358	-18.945	-2.135	1.378
Rec.%	88.146	88.642	81.055	97.865	101.378

### Calibration plot

Employing the conditions described in the procedure, a linear calibration curve for hydroquinone is obtained in figure(8), which shows that Beer's law is obeyed over the concentration range ( $1\text{--}26\ \mu\text{g}\cdot\text{ml}^{-1}$ ) with correlation coefficient of 0.9997 with detection limit of  $0.0125\ \mu\text{g}\cdot\text{ml}^{-1}$ .

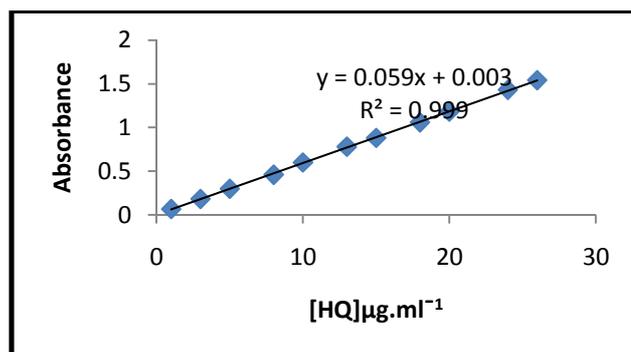


Fig (8): The calibration curve of hydroquinone using spectrophotometric method

### Structure of the colored product

The stoichiometry of the oxidation reaction between hydroquinone with  $\text{KMnO}_4$  was investigated using job's method. The result obtained in figure (9) shows that 1:1 drug to reagent was formed at 610 nm in presence of sodium hydroxide solution [40,41]

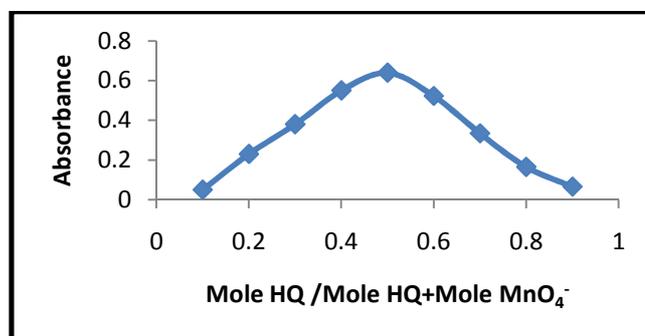


Fig (9) : Continuous variation plot of the reaction between HQ and potassium permanganate (0.02M)

### Precision and Accuracy

To evaluate the accuracy and precision of the methods, pure drug analyzed, each determination being repeated six times at three different concentration [42].

Table (2): Accuracy and precision of proposed methods

HQ taken	HQ Found	*Rec%	%E	RSD%
3	2.935	99.97	-0.0216	0.637
10	10.15	101.5	+1.5	1.269
14	14.18	101.285	+1.285	1.47

\*Average of six determinations.

The result shown in table (2) indicate that satisfactory precision and accuracy could be attained with the proposed method . The %E and RSD % values were less than 1.5 % which indicate the high accuracy.

Table (3): Application of the proposed method and pharmaceutical preparations for determination of hydroquinone drug

H Q Samples	HQ( $\mu\text{g.ml}^{-1}$ )		Rec%	Average recovery%	RSD%
	Taken	found			
Fediquin	5	4.863	99.97	99.993	0.314
	10	10.163	100.016		0.316
Hydropaque	5	4.776	99.95	99.984	0.527
	10	10.193	100.019		0.315
Hydroquinone	5	4.846	99.969	99.973	0.315
	10	9.776	99.977		0.426

1. Marked by Jordan 2. Marked by Syria 3. Marked by Syria

### Analytical Application

Proposed method have been used Fediquin(40%), Hydropaque (40%), Hydroquinone (2%) drugs containing hydroquinone and it gave good accuracy and precision as shown in table (3) , the proposed method compared with

standard method [41], since T-test and F-test shows that there was no significant differences the proposed method and standard method, the results obtained were tabulated in table (4).

Table (4) :Comparison of hydroquinone determined in pharmaceutical preparation by the proposed method with standard method

HQ Sample	Rec%	
	Proposed method	Standard method
Fediquin	99.993	99.75
Hydropaque	99.984	99.48
Hydroquinone	99.973	99.67

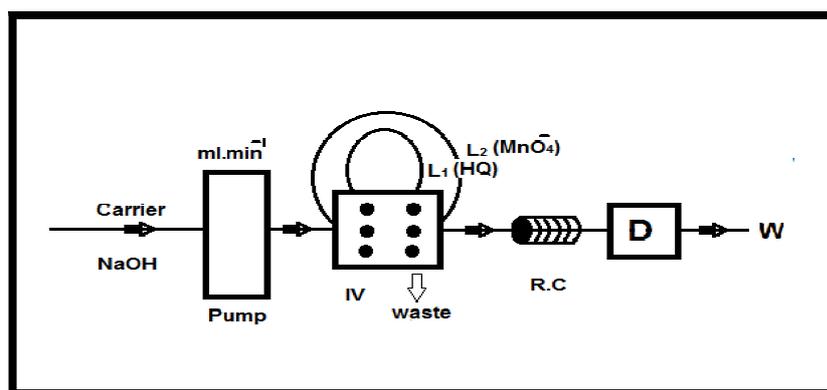
The proposed method was compared with other spectroscopic method in literature for the oxidization reaction of hydroquinone , as shown in table (5).

Table (5): Comparison of hydroquinone determination in the proposed method and other literature methods

Reagent	$\lambda_{max}$ . nm.	Limit of detection	Linear range $\mu\text{g.ml}^{-1}$	Ref.
Rhodamine B (RhB)	557	$0.16\mu\text{g.ml}^{-1}$	0.36-3.96	42
Ammoniummeta-vanadate	245.5	$7\text{ng.ml}^{-1}$	0.025-205	43
Methanol	293		0-1.3	44
Sulfuric acid	225		10-26	45
Potassium permanganate	610	$0.0125\mu\text{g.ml}^{-1}$	1-26	Proposed method

### Chemical and flow optimization

The flow injection manifold depicted in figure (10) were investigated in the relation to chemical and flow variable in order to obtain optimum conditions for system .They were optimized by making all variables constant and varying one each at a time.



Figure(10): Flowgram of the whole manifold with double loop -6- three way valve (IV; injection valve, RC; reaction coil, D; detector, W; waste)

### Choice of base

The oxidative reaction of HQ with potassium permanganate can be conducted in basic medium . Therefore , bases (NaOH , KOH ,  $\text{NH}_4\text{OH}$  ,  $\text{Na}_2\text{CO}_3$ ) were used at various concentration, sodium hydroxide was chosen as a favorable base for oxidation of drug using  $\text{KMnO}_4$  as oxidizing agent. The use of a 0.1M of sodium hydroxide as a carrier stream at  $2.12\text{ml. min}^{-1}$ ,  $227.65$  of  $2.5 \times 10^{-2} \text{MKMnO}_4$  and  $227.65\mu\text{L}$  of  $50\mu\text{g.ml}^{-1}$  of HQ.

### Effect of sodium hydroxide concentration

Various concentration (0.05-0.7M) of sodium hydroxide solution were used to obtain the optimum concentration of sodium hydroxide that can be used as carrier stream at  $2.12\text{ ml.min}^{-1}$  using  $227.65\mu\text{l}$  of  $50\mu\text{g.L}^{-1}$  of HQ  $227.65\mu\text{L}$  of  $2.5 \times 10^{-2} \text{mol.L}^{-1}$  of potassium permanganate, figure(11) shows the variation of absorbance with the molar concentration of sodium hydroxide which shows clearly that 0.1 M of sodium hydroxide is a propitious concentration.

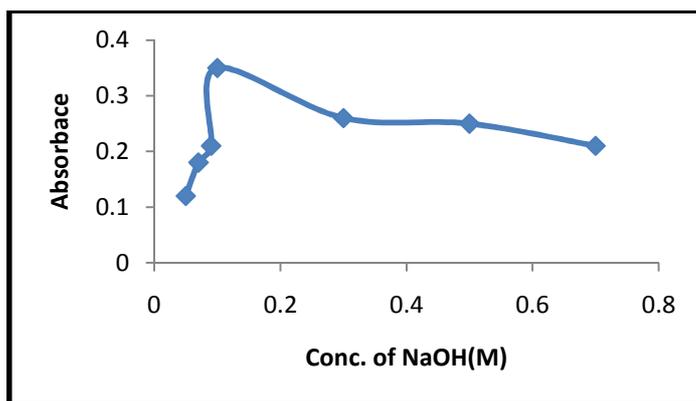
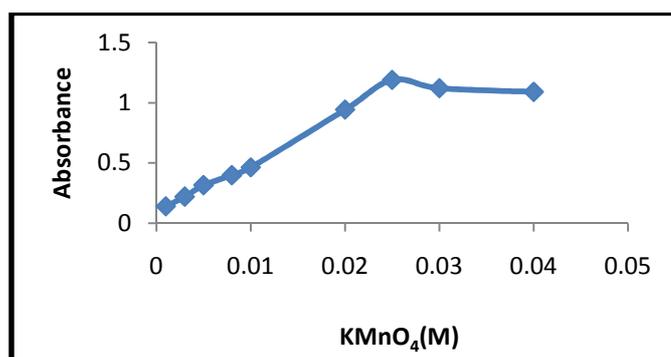


Fig (11) :Effect of variation of absorbance versus molar concentration of NaOH(M)

#### Optimization of $\text{KMnO}_4$ concentration

The effect of adding various amount of  $\text{KMnO}_4$  solution on absorbance of  $50 \mu\text{g}\cdot\text{ml}^{-1}$  HQ solution is given in figure(12). It is seen that the maximum absorbance of  $2.5 \times 10^{-2} \text{M}$  occurs in the presence of various concentrations ( $1 \times 10^{-3}$  -  $4 \times 10^{-2} \text{M}$ ) of  $\text{KMnO}_4$ , the concentration of  $2.5 \times 10^{-2} \text{M}$   $\text{KMnO}_4$  solution showing an absorbance 1.189 was taken as optimum amount for further study because a calibration curve check of HQ in case of former solution did not result in better absorbance value at lower HQ concentration. According to literature [24], application of a higher  $\text{KMnO}_4$  concentration would facilitate the determination of HQ in higher concentration range.



Fig(12): Effect of  $\text{KMnO}_4$  on absorbance of HQ

#### Physical parameters

##### Sodium hydroxide flow rate

The effect of sodium hydroxide flow rate was investigated in the range of  $(1.5-6.5) \text{ ml}\cdot\text{min}^{-1}$  to obtain the best absorbance where sodium hydroxide flow rate of  $2.12 \text{ ml}\cdot\text{min}^{-1}$  gave the highest response as shown in figure(13).

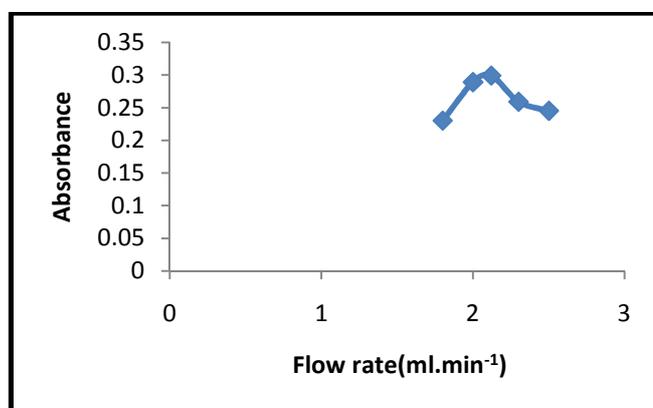


Fig (13): Effect of flow rate of NaOH on absorbance of HQ

**Effect of sample volume**

The sample volumes (157,196.25,227.65,274.75  $\mu\text{L}$ ) were evaluated using different length of sample loop and the results were plotted in figure (14). The results obtained showed that injected sample of 227.65  $\mu\text{L}$  gave the best absorbance.

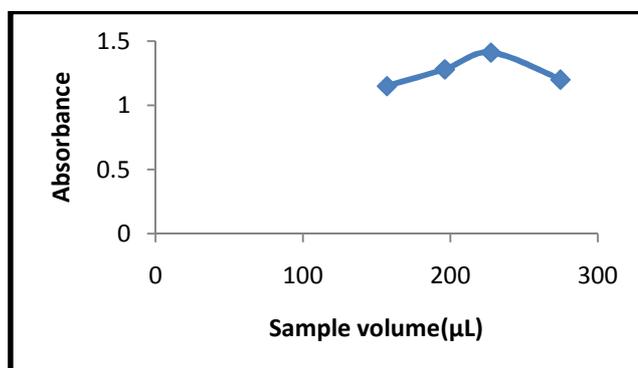
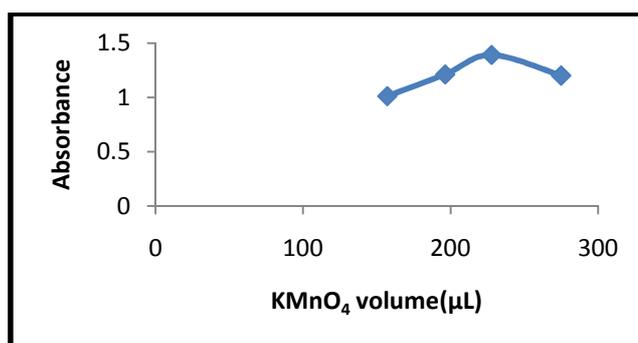


Fig (14): Variation of absorbance versus the injected HQ volume

**Effect of reagent volume**

Different potassium permanganate volumes of (157,196.25,227.65,274.75  $\mu\text{L}$ ) can be achieved by inserting different lengths of reagent loop. Figure (15) shows that 227.65  $\mu\text{L}$  is the optimum volume.



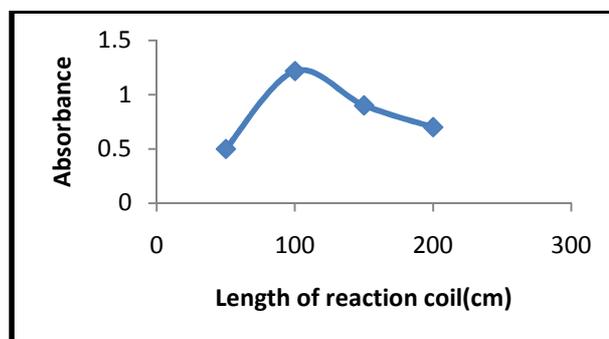
Fig(15): Variation of absorbance versus the reagent volume

**Effect of temperature**

The effect of temperature on the reaction of  $[\text{HQ-MnO}_4\text{-OH}]^-$  system was studied. It was found that there was no effect of the temperature on the reaction for temp.range( 5-50 $^{\circ}\text{C}$  ).

**Effect of mixing coil**

Different delay reaction coil length (50,100,150,200 cm) were used to measure the absorbance of colored product at  $2.5 \times 10^{-2}\text{M}$  potassium permanganate, 0.1 M base (NaOH) and  $50 \mu\text{g}\cdot\text{ml}^{-1}$  HQ. Figure(16) shows that 100cm gave the highest absorbance and was used in all subsequent experiments.



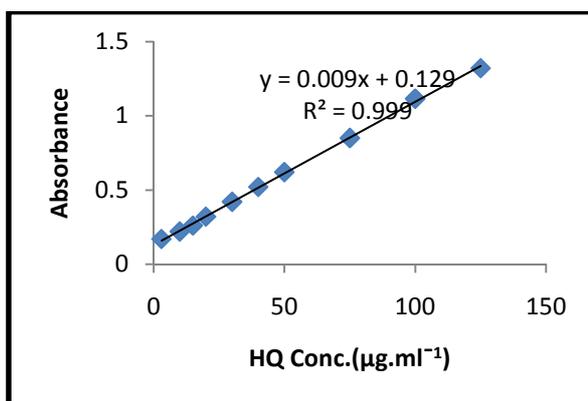
Fig(16) : Effect of the length of reaction coil in ( cm)

**Calibration curve**

Series of HQ solutions( 3,5,10,15,20,30,40,50,75,100,125) $\mu\text{g.ml}^{-1}$ . were prepared from stock solutions under the optimum conditions of reagent and manifold variables as indicated in table (6). The calibration curve is linear in the concentration rang (3-125)  $\mu\text{g.ml}^{-1}$ with adetection limit of  $0.25\mu\text{g.ml}^{-1}$ .Figure (17) shows the variation of absorbance of color product versus HQ concentration and table (7) shows the treatment of the results[46,47] in linear regression terms.

**Table (6): The optimization of the chemical and FIA parameters using Merging Zones –Flow injection analysis for determination of HQ drug**

Parameters	Optimum value
NaOH Conc.	0.1 M
KMnO <sub>4</sub> Conc.	$2.5 \times 10^{-2} \text{M}$
Flow of rate of NaOH	$2.12 \text{ ml.min}^{-1}$
Sample volume ( $\mu\text{L}$ )	$227.65 \mu\text{L}$
Oxidant volume ( $\mu\text{L}$ )	$227.65 \mu\text{L}$
Reaction coil length (cm)	100cm



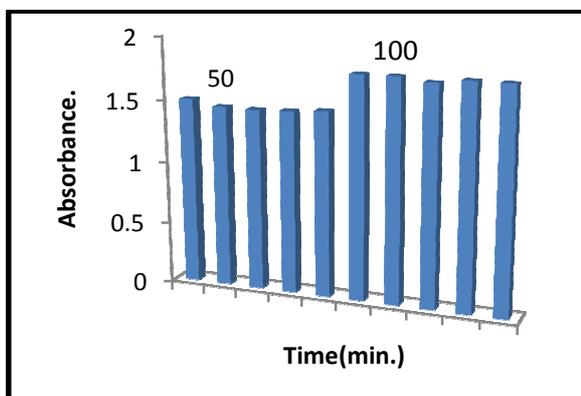
**Fig (17): The calibration curve of Hydroquinone at 0.1M NaOH as a carrier stream at flow rate  $2.21 \text{ ml.min}^{-1}$**

**Table (7): The linear equation results for the color product by the injection of  $227.65 \mu\text{L}$ . of HQ sample through the adopted system [HQ –MnO<sub>4</sub> – OH]**

Linear rang Conc. $\mu\text{g.ml}^{-1}$	Slop (b) at confidence limit 95% for (n-2) ( $b \pm S_b, t$ )	Intercept(a) at confidence limit 95% for (n-2) ( $a \pm S_a, t$ )	t from table at confidence limit 95% for (n-2)	Calculate $t = \frac{ r \sqrt{n-2}}{\sqrt{1-r^2}}$	Correlation coefficient (r)	Linearity R <sup>2</sup> %
3-125	$0.0096 \pm 0.00014$	$0.1299 \pm 0.0085$	2.31	100.964	0.9995	0.9992

**Repeatability**

Repeatability was studied at determination of hydroquinone via measurements of oxidization of drug with potassium permanganate in basic medium to form color product (green) for FIA system [HQ-MnO<sub>4</sub>-OH]. Variable concentration of HQ  $50, 100 \mu\text{g.ml}^{-1}$  were injected. Each concentration was injected successfully for five times. Figure(18) shows clearly that relative standard deviation is better than 1.5% in most cases can be obtained of a high repeatability by a short analysis time.



**Fig(18): Repeatability for five successive measurement of hydroquinone for  $50, 100 \mu\text{g.ml}^{-1}$  as injected sample**

**Determination of HQ in pharmaceutical formulations by proposed method and spectro method (40)**

Table (8) shows the agreement between the results in both methods and the paired t – test illustrates that the flow injection analysis method has no significant difference when compared with the standard adopted method ,therefore it can be regarded as an alternative determination method a part from many advantages this method had.

**Table (8) : Comparison between proposed method and spectro method for the determination of HQ in pharmaceutical formulations**

Sample	HQ found in samples (mg.ml <sup>-1</sup> )		di μg.ml <sup>-1</sup>	X̄d μg.ml <sup>-1</sup>	S <sub>d</sub>	Paired t-test = $\frac{\bar{x}_d \sqrt{n}}{s_d}$	T form table at confidence limit 95% (n-1)
	Merging zone FIA	Spectro Method					
Fediquin	0.041	0.0398	0.0012	0.00153	0.002	1.325	<4.303
Hydropaque	0.042	0.0397	0.0023				
Hydroquinone	0.021	0.0199	0.0011				

**Interference:**

The effect of some foreign compounds[40] , which often found in pharmaceutical products , were studied by adding different amount organic molecules to 5 ml of 500 μg.ml<sup>-1</sup> of hydroquinone . The color was developed following the recommended procedure described earlier. It was observed were not interfering with the determination of levels found dosage form.

**CONCLUSION**

The work described in this research comprises more than a complete new intuition for a compact home made new injection valve using Merging Zone technique .The simplicity , speed , reliability and the mode of working its well comparison with standard spectrophotometric method indicates that the method presented in this research work can be used as an alternative method for the spectrophotometric method.The work has been developed for the determination of HQ in pure material and pharmaceutical preparations in alkaline medium based on oxidation reaction using potassium permanganate as reagent for conversion of HQ to BQ [9] analysis of the former at the extent of formation of the later is very useful for analysis of HQ at ultra – trace level .It is a very good example of redox reaction and may be employed in situation involving useful oxidation products of such types .To our knowledge no what so ever this technique was used in any published work everywhere else using FIA- Merging Zone technique for determination of HQ. This method has clear edge over other methods employing expensive and or hazardous reagents ,the proposed method does not require temperature control or solvent extraction step. Moreover , the lower detection limit achieved with highly reproducible in results by this method which is not possible by other methods..

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