Conversion of dermal proteins of an Algerian camel to an antimicrobial agent for wound healing

Tabak Messekine Djamila¹,²*, Meddah boumedienne¹, Serier Aicha², Benouguef Sadia¹ and Bensaha Fatima Zohra¹

¹Laboratoire de bioconversion, génie microbiologique et sécurité sanitaire, Laboratoire LRSBG, Faculté des sciences de la nature et de la vie, Université de Mascara, Algérie
²Laboratoire Revêtement, Matériaux & Environnement, Faculté des Sciences de l’Ingénieur Université de Boumerdes, Algérie

ABSTRACT

The aim of this study was the conversion of the camelin dermal proteins as a natural product that has an antimicrobial power used for tissue repair and wound healing. After dehairing, fleshings, deliming the animal tissues (Camelin) and purification, we supplied and fixed the essential oils of Lavandula officinalis, Eucalyptus globules and Eugenia caryophyllata. The product obtained is a spongy material communicating with open voids between the fibers. With a light yellow color, it has a remarkable antibacterial power and a very strong inhibitory activity on all the bacterial strains tested whose average diameter of the inhibition zones exceeds 15 mm. It is also characterized by a smell which can be aromatic, pleasant and spicy or fresh and spicy depending on the essential oil used.

Key words: Conversion, dermal proteins, medical purposes, essential oils, cattle, dromedary.

INTRODUCTION

The extraction technique of proteins and their preparation as biodegradable products from animal skins are well known for many years in medicine for the treatment of wound and the pathological cavity in the bones [1, 2, 3]. However, these products raise always a problem of potential transmission of an infectious agent, known or unknown, despite the severity of manufacturing and control procedures [4]. Some studies have been carried out actually to eradicate this problem from different antibiotics known by their antibacterial effect [5, 6] as it is the case for the gentamicin compress [7]. At the end of the 1980s and in the 1990s this process began to be no longer regarded as a miracle as it seems to be 40 years earlier. Indeed, many bacteria have developed a resistance to most antibiotics and, thus it seems important to find another alternative. Studies have shown that essential oils could be a particularly credible application. Antimicrobial activity of essential oils is known empirically since antiquity; their effectiveness as anti infection was scientifically established in vitro and in vivo. They have a genuine spectrum anti infectious including isolated cases of resistant infection to antibiotics [8]. Thus, our present study is looking into the preparation of an antimicrobial product based on camelin dermic protein powered by essential oils. This preparation is followed by tests of antimicrobial activity on the finished product.

EXPERIMENTAL SECTION

Dermic protein preparation

The dermic protein preparation and transformation procedure was carried out from 36 months-aged south Algerian (Timimoine region) mortal remains camelin, in the manner described below:
Cleaning: The four parts of the weight of the mortal remains skin, weighing 5kg, was soaked in 5 to 15% aqueous sodium chloride solution, containing approximately 0.2 to 1 weight parts of sodium azide as preservative agent for 1000 parts in weight of the given solution, and from 0.5 to 2% in weight of non-ionic fat, a dispersing and wetting agent (phenolic ether of nonyl polyoxyethylene), with a temperature ranging from 18° to 20° for 24 hours, in a fuller, in order to remove impurities and stains.

Removal of hair: hairs are removed with a pelain bath, containing 4 weight parts with regards to the lime mortal remain weight 0.1M, with a temperature from 18° to 20° for 48 hours. The PH of the solution must be ranging between 12 and 13.5 in a fuller.

Hypoderm elimination: the subcutaneous tissue is eliminated mechanically with a defleshing instrument. The blades of the machine cylinder plane the subcutaneous tissue, which constitute an alcaline and a lot hydrated waste, called ‘carnasse’.

Decalcifying: the defleshed tissue, consisting of 2 parts of its new weight, is soaked in a bath of a mixture of 2g/l of ammonium sulfate and 0.5g/l of sodium metabisulfite. This procedure is required to eliminate the alcaline products combined to dermic proteins [9].

The obtained pulp after these operations has a whitish color. 1 kg of it has been processed according to the Piter and Ries method [10], consisting:

Crushing: frozen from -10°C to -20°C, and finely crushed by means of a high-speed homogenizing knife. The temperature of the crushed matter has been kept below 40°C, by adding ices.

Paste cleaning: put in suspension, and shaked up strongly, at the same time, in order to wash well the paste in the 5 parts of its volume, in acetic acid at 0.5 M, and centrifuged.

Digestion: the solid centrifugal has been put in suspension in 5-litre acetic acid at 3%, containing 1g of technic pepsine for 1000 weight parts tissue, used as starting matter in 100ml of HCl at 0.01 N, in order to eliminate non-collagenous-type proteins and telopeptids. The suspension pH has been adjusted at 2.9 by means of HCl. The repeatedly shaken suspension is digested for 48 hours. The visquous camelin dermic protein solution was filtered by a G1 drawing up filter, in order to eliminate non degraded residues.

Precipitation: The camelin dermic proteins visquous filtrate was precipitated from the suspension by adding an aqueous solution of sodium hydroxide at 30% and separated by centrifugation.

Purification: Camelin dermic proteins were purified by dissolution in acetic acid at 0.5 M, and precipitation by slow adding of 3% of aqueous sodium chloride.

Dissolution: Purified camelin dermic proteins have been dissolved in acetic acid at 0.5M and diluted in water. Residual sodium chloride present in the proteins has been eliminated by cleaning using an ultrafilter. Ultrafiltration was carried on till the chloride ion will be undetectable in the eluate, after adding silver nitrate. The concentration of proteins increases to almost 2%.

Freeze-drying: the camelin dermic protein solution was filtered and poured in glass boxes, and freeze-dried at 0.5 cm in thickness.

Thermal processing: the semi-finite product has been heated to 100°C for an hour and half in a electrically heated drying classical oven, in order to improve its absorption and resistance against dampness, and then sterilized by irradiation with a dose of 2.5 billion of γ rays.

Dermic protein transformations
Feeding: It was divided into four parts. Each part is fed with essential oil, with a repeated tender shake for a half an hour to allow seepage and fixation of essential oils, except the last part which remains as a witness (see table 1).

The used oils are: lavender, clove and eucalyptus (see table 2). The used amounts depend on the efficient threshold, where the minimal inhibitive concentration is defined as the lowest oil concentration able to inhibit any bacterial growth [11]. Finally, the dampened products at 5% were packed in thermo-sealed polyethylene bags.
Table 1: Supply Formula of dermal camelin proteins

<table>
<thead>
<tr>
<th></th>
<th>% essential oil</th>
<th>% pur ethanol</th>
<th>% of water</th>
</tr>
</thead>
<tbody>
<tr>
<td>1st batch</td>
<td>Eucalyptus globulus</td>
<td>0.1</td>
<td>25</td>
</tr>
<tr>
<td>2nd batch</td>
<td>Eugenia caryophyllata</td>
<td>0.1</td>
<td>25</td>
</tr>
<tr>
<td>3rd batch</td>
<td>Lavandula officinalis</td>
<td>0.8</td>
<td>25</td>
</tr>
<tr>
<td>4th batch without essential oil</td>
<td></td>
<td>0</td>
<td>25</td>
</tr>
</tbody>
</table>

Percentages are relative to the weight of camel dermal protein (product).

Table 2: Characteristics and chemical constituents of essential oils

<table>
<thead>
<tr>
<th>Essential oil</th>
<th>Color</th>
<th>Smell</th>
<th>Main chemical constituents %</th>
<th>Aspect</th>
</tr>
</thead>
<tbody>
<tr>
<td>Eucalyptus globulus</td>
<td>Very pale yellow</td>
<td>fraîche et épicée</td>
<td>Eucalyptol 70%</td>
<td>Liquid</td>
</tr>
<tr>
<td>Eugenia caryophyllata</td>
<td>Light yellow</td>
<td>Agréable et épicié</td>
<td>Eugénol 78%</td>
<td>Liquid</td>
</tr>
<tr>
<td>Lavandula officinalis</td>
<td>Light yellow</td>
<td>Aromatique</td>
<td>Linalool Linalyl acetate 51% 19%</td>
<td>Liquid</td>
</tr>
</tbody>
</table>

Table 3: bacterial strain

<table>
<thead>
<tr>
<th>Strain name</th>
<th>Reference</th>
<th>Gram</th>
<th>Family</th>
</tr>
</thead>
<tbody>
<tr>
<td>Staphylococcus aureus</td>
<td>ATCC6538</td>
<td>+</td>
<td>Micrococcaceae</td>
</tr>
<tr>
<td>Escherichia coli</td>
<td>ATCC25922</td>
<td>-</td>
<td>Enterobacteriaceae</td>
</tr>
<tr>
<td>Pseudomonas Aeruginosa</td>
<td>ATCC10145</td>
<td>-</td>
<td>Pseudomonaceae</td>
</tr>
<tr>
<td>Klebsiella pneumoniae</td>
<td>Sauvage</td>
<td>-</td>
<td>Enterobacteriaceae</td>
</tr>
<tr>
<td>Salmonella heiderberg</td>
<td>Sauvage</td>
<td>-</td>
<td>Enterobacteriaceae</td>
</tr>
</tbody>
</table>

Biological materials

Bacterial strains: to estimate the anti-microbial activity, we have used wild strains and reference strains widely met in many human pathologies. Tests are performed on five bacteria from Pasteur Institute (Algeria) (see table 3).

Anti-bacterial activity assessment: The anti-bacterial activity assessment of various groups have been measured by the aromatogram technique, similar to that of the antibiogram used to test antibiotics [13].

We have used camelin dermic protein discs (products) of various groups of 5mm in diameter, put on the surface of a Mueller-Hinton gelosed environment. It’s a standardized environment according to WHO standard, i.e. in the way that it allows the growth of many bacteria. The environment is aseptically sown by 1ml of suspension of each strain, by using STRIS method. Boxes are pre-dried before used [13, 14]. After incubation of knead boxes in the steam room at 37°C for 24 hours. The effect of the product is seen by the formation of an inhibition halo around the disc.

The reading of the results is made by the measure of the inhibition diameter (in mm), by means of a caliper rule or a ruler. HE dilution on the camelin dermic protein discs always happens in a solvent such as glycol ethylene [15], acetone [16], ethanol at 95% [17]. The principle of this method is always the HE migration by the diffusion in the gelose. This technique inspired from that of the antibiograms, has been generalized to HE [18].

A product is said active if it has an inhibition diameter greater or equal to 15mm [19].

RESULTS

The transformation of the camelin dermic proteins allowed to obtain a product which has a biodegradable spongy structure, communicating with open spaces between fibers.

The product obtained after deliming has a little bit white color (Fig.N°1), on the other hand the semi-finished product has a very white color (Fig. N°2). After supply and fixation of various types of essential oils, the color of the product turns on clear yellow as seen on fig. N°3 with a spicy, fresh aromatic or pleasant spiced smell depending on the nature of essential oil used.

The study of the antibacterial power of products (camelin dermic proteins) fed by Eucalyptus globulus, Eugenia caryophyllata, Lavandula officinalis was made by the method of spreading on gelose. The measure of the diameter of the inhibition zones including the disc of our product (5mm) allowed to determine the antimicrobial activity of our in vitro products. Table N°4 followed by the graph N°4 show the results of the average antimicrobial activity tests of the various batch on bacterial strains of Staphylococcus aureus, Escherichia coli, Pseudomonas Aeruginosa, Klebsiella pneumoniae, Salmonella heiderberg.
Tabak Messekine Djamila et al


Fig. n°1: camel skin after deliming

Fig. n°2: camelin protein semi finished

Fig. n°3: camelin protein - finished product

Table 4: Test of the product’s antimicrobial activity

<table>
<thead>
<tr>
<th>Batches</th>
<th>Zone inhibition moyenne (mm)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Staphylococcus aureus</td>
</tr>
<tr>
<td>1\textsuperscript{st} batch Eucalyptus globulus</td>
<td>22±0.22</td>
</tr>
<tr>
<td>2\textsuperscript{nd} batch Eugénia caryophyllata</td>
<td>23.4±0.25</td>
</tr>
<tr>
<td>3\textsuperscript{rd} batch Lavandula officinalis</td>
<td>19.7±0.42</td>
</tr>
<tr>
<td>4\textsuperscript{th} batch without essential oil</td>
<td>R</td>
</tr>
</tbody>
</table>

selon ponce [19] R : résistant, non sensible (-), sensible(+), très sensible(++), extrêmement sensible(+++)

Fig. n°4: Test of the product’s antimicrobial activity
DISCUSSION

We note that the essential oils were easily absorbed into our product. According to (Dorman and Deons, on 2000) [20] active essential oils contain secondary metabolism components that are small enough to pass through the protein.

We think that the penetration and the fixation of essential oils in the product based on camelin dermic protein is caused by $\text{NH}_4^+$ of proteins which favor the fixation of the essential oil after the transfer of its proton.

The product had a very strong inhibitive activity on all the tested bacterial strains with average values of the diameter of the inhibition zones exceeding 15 mm for the 3rd lot (Lavandula officinalis). The Strains remained extremely sensitive for Staphylococcus aureus towards the 1st lot eucalyptus globulus and the 2nd lot Eugénia caryophyllata. According to Kalemba and Kunicka, the sensibility of a microorganism to essential oil depends on the properties of the essential oil and on the microorganism itself. It is well known that bacteria Gram (+) are more sensitive to essential oils, and confirm this phenomenon (Pool [21]. 2001; Brut [22]. 2004; Beckechi [23]. 2008).

The first three batches gave a broad-spectrum antibacterial activity acting as well on bacteria Gram(+) as Gram(-). The antibacterial properties of the main active components of essential oil (Eugenol, 1,8-cineole(Eucalyptol), linalool, Acetata of linalyl) are in part related to their lipophilic character leading to the accumulation in the bacterial cell walls [23, 24, 25]. This engenders a disturbance on the functioning and the permeability of the cellular membranes, a degradation of the cellular wall [26, 27], damage to the cytoplasmic membrane and the cell content leaks [28, 29, 30]. On the other hand, the 4th batch remained resistant.

CONCLUSION

In this study we have shown how dermal proteins can be obtained from a camel skin. These proteins are transformed into an antimicrobial product which has biodegradable spongy structure with a light yellow color and an aromatic, or pleasant and spicy or fresh and spicy smell depending on the nature of the essential oil used, in particular eucalyptus (Eucalyptus globulus) the clove (Eugénia caryophyllata) and lavender (Lavandula officinalis). The product has a very strong inhibitory activity on all tested bacterial strains of Staphylococcus aureus, Escherichia coli, Pseudomonas aeruginosa, Klebsiella pneumoniae, Salmonella heiderberg whose average values of the diameter of the inhibition zones is greater than 15 mm. This is particularly true for product supplied by (Lavandula officinalis).

The results are very promising, the strains remained extremely sensitive for Staphylococcus aureus towards the product supplied by Eugela Globulus or Eugenia caryophyllata. The product gives a broad-spectrum antibacterial activity acting as well on bacteria Gram(+) as Gram(-). It is easy to see that the product obtained can be used for medical purposes.

REFERENCES

[8] S Inouye, S Abe., Phytothérapie., 2007,1, 2-4
[23] C Beckchi ; F Atik-Bekkara ; DF Ouahid., 2008(abstr), *phytothérapica* p.12
[24] IM Helander ; H L Alakomi ; K Latvala., T Mattila-Sandholm., I Pol ; E J Smid ; G M Gorrsl ; A Vonwright., *Journal of Agricultural and food chemistry*. 1998. 46(9), 3590-3595
[26] A Ultee ; M H J Bennik ; R Moezelaar., *applied and Environnemental microbiology*. 2002. 68. 220-225